



TSQ Duo

Mass Spectrometer

User Guide

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Contents

	Prefaceix
	About Your System
	Related Documentationx
	System Requirements
	Safety and Special Notices
	Special Notices
	Safety Symbols and Signal Wordsxi
	Hydrogen Safety Precautions
	Using Hydrogen with a GC-MS/MS Systemxiv
	Hydrogen Connection Guidelinesxv
	Purchasing Hydrogenxvi
	Properly Storing Hydrogen xvii
	Hydrogen Safety Codes, Standards and Referencesxix
	Hazardous Substances Precautions xx
	Biological Hazard Warning Note
	Venting Toxic Gasesxxi
	Contacting Us
Chapter 1	Introduction1
	Confirming Your Instrument is Working1
	Checking Power to the System1
	Verifying the Carrier Gas Flow Rate2
	Checking Your Carrier Gas Tank Pressure
	Verify Collision Gas Tank Pressure
	Checking the Vacuum and Temperature
	Cleaning the Exterior of Your Instrument
	Configuring Your Instrument
Chapter 2	Changing the Column
	Determining the Column Type
	Replacing the Factory Installed Column
	Connecting the Column to the Transfer Line
Chapter 3	Tuning
	Accessing Auto Tune

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	Tune Types
	EI Initial Tune
	EI Standard Tune
	EI Standard Quick Tune
	EI SRM Tune
	EI SRM Quick Tune
	EI Tune Check
	EI Diagnostics Only
	EI Target Tune
	Fast Scan Tune
	Negative CI Tune
	Positive CI Tune
	Tuning the Mass Spectrometer
	Updating Tunes for New RF Lens
Chapter 4	Creating a Method53
	Accessing the Method Editor
	Creating a GC-MS method54
	Running a Sequence
Chapter 5	Optimizing Your Method91
	Changing the Chromatographic Separation91
	Finding the Best Way to Make an Injection
	Improving the Way You Prepare Samples
	Changing the Dwell Time or Scan Rate92
	Narrowing the Mass Range
	Adjusting the Transfer Line and Ion Source Temperature
	Optimizing an SRM Method94
	Modifying an Automatic Tune95
Chapter 6	Computer Settings
	System Requirements
	Computer Settings
	Excluding the Xcalibur Directory from Virus Scan
Chapter 7	Troubleshooting
	Setting Instrument Conditions for Troubleshooting
	Checking Air/Water Spectra
	Diagnostics Checks
	How to Know When Your System Needs Maintenance
	Investigating Baseline Issues
	Investigating Peak Issues
	Investigating Results Issues
	Index

Declaration

Manufacturer: Thermo Fisher Scientific

Thermo Fisher Scientific is the manufacturer of the instrument described in this manual and, as such, is responsible for the instrument safety, reliability and performance only if:

- installation,
- recalibration, and
- changes and repairs

have been carried out by authorized personnel and if:

- the local installation complies with local law regulations,
- the instrument is used according to the instructions provided, and
- if its operation is only entrusted to qualified trained personnel.

Thermo Fisher Scientific is not liable for any damages derived from the non-compliance with the aforementioned recommendations.

Regulatory Compliance

Thermo Fisher Scientific performs complete testing and evaluation of its products to ensure full compliance with applicable domestic and international regulations. When the system is delivered to you, it meets all pertinent electromagnetic compatibility (EMC) and safety standards as described in the next section or sections by product name.

Changes that you make to your system may void compliance with one or more of these EMC and safety standards. Changes to your system include replacing a part or adding components, options, or peripherals not specifically authorized and qualified by Thermo Fisher Scientific. To ensure continued compliance with EMC and safety standards, replacement parts and additional components, options, and peripherals must be ordered from Thermo Fisher Scientific or one of its authorized representatives.

EMC and Safety Standards

- ITQ and Ion Trap Series standards: EMC EN 61326-1:2006. Safety IEC 61010-1:2001, IEC 61010-2-081:2001
- Direct Probe Controller (DPC) standards: EMC EN 61326-1:2006. Safety IEC 61010-1:2001, IEC 61010-2-081:2001
- ISQ standards: EMC EN 61326-1:2013. Safety IEC 61010-1:2010, IEC 61010-2-010:2014, IEC 61010-2-081:2015
- TSQ 8000 Evo and TSQ Duo standards: EMC EN 61326-1:22013. Safety IEC 61010-1:2010, IEC 61010-2-010:2014, IEC 61010-2-081:2015

Low Voltage Safety Compliance

This device complies with Low Voltage Directive 2014/35/EU and harmonized standard EN 61010-1:2001.

FCC Compliance Statement

THIS DEVICE COMPLIES WITH PART 15 OF THE FCC RULES. OPERATION IS SUBJECT TO THE FOLLOWING TWO CONDITIONS: (1) THIS DEVICE MAY NOT CAUSE HARMFUL INTERFERENCE, AND (2) THIS DEVICE MUST ACCEPT ANY INTERFERENCE RECEIVED, INCLUDING INTERFERENCE THAT MAY CAUSE UNDESIRED OPERATION.



CAUTION Read and understand the various precautionary notes, signs, and symbols contained inside this manual pertaining to the safe use and operation of this product before using the device.

Notice on Lifting and Handling of Thermo Scientific Instruments

For your safety, and in compliance with international regulations, the physical handling of this Thermo Fisher Scientific instrument *requires a team effort* to lift and/or move the instrument. This instrument is too heavy and/or bulky for one person alone to handle safely.

Notice on the Proper Use of Thermo Scientific Instruments

In compliance with international regulations: Use of this instrument in a manner not specified by Thermo Fisher Scientific could impair any protection provided by the instrument.

Notice on the Susceptibility to Electromagnetic Transmissions

Your instrument is designed to work in a controlled electromagnetic environment. Do not use radio frequency transmitters, such as mobile phones, in close proximity to the instrument.

For manufacturing location, see the label on the instrument.

WEEE Compliance

This product is required to comply with the European Union's Waste Electrical & Electronic Equipment (WEEE) Directive 2002/96/EC. It is marked with the following symbol:



Thermo Fisher Scientific has contracted with one or more recycling or disposal companies in each European Union (EU) Member State, and these companies should dispose of or recycle this product. See www.thermoscientific.com/ rohsweee for further information on Thermo Fisher Scientific's compliance with these Directives and the recyclers in your country.

WEEE Konformität

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Conformité DEEE

Ce produit doit être conforme à la directive européenne (2002/96/EC) des Déchets d'Equipements Electriques et Electroniques (DEEE). Il est marqué par le symbole suivant:



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Preface

This guide contains detailed information about how to use your Thermo Scientific TSQ Duo triple-quadrupole GC/MS system. The TSQ Duo system provides the selectivity and sensitivity of a triple-quadrupole GC/MS while also functioning as a high-performance single quadrupole instrument. The system is designed to stay cleaner longer to maximize your instrument's uptime and improve your lab's productivity. In addition, the TSQ Duo system includes innovative Thermo Scientific software that will help users new to triple-quadrupole GC/MS/MS systems develop selected reaction monitoring (SRM) methods.

Contents

- About Your System
- Related Documentation
- System Requirements
- Safety and Special Notices
- Hydrogen Safety Precautions
- Hazardous Substances Precautions
- Contacting Us

About Your System

Thermo Scientific systems provide the highest caliber gas chromatography/mass spectrometry (GC/MS) instrumentation available on today's market.

GC/MS represents a combination of two powerful analytical techniques: GC, which acts as a separation technique, and MS, which acts as a detection technique. Complex mixtures of individual compounds can be injected into the GC, either manually or by an autosampler and then separated for presentation to the MS. The MS will generate a mass spectrum of the GC eluate and its components. The mass spectrum can then be used for qualitative identification as well as accurate and precise quantification of the individual compounds present in the sample.

A triple-quadrupole GC/MS/MS system provides the extra selectivity required for trace analysis of compounds in complex matrices.



WARNING Thermo Scientific systems operate safely and reliably under carefully controlled environmental conditions. If the equipment is used in a manner not specified by the manufacturer, the protections provided by the equipment might be impaired. If you maintain a system outside the specifications listed in this guide, failures of many types, including personal injury or death, might occur. The repair of instrument failures caused by operation in a manner not specified by the manufacturer is specifically excluded from the standard warranty and service contract coverage.

Related Documentation

The TSQ Duo system includes Help and these manuals as PDF files:

- TSQ Duo Preinstallation Guide, PN 1R120587-0001
- TSQ Duo User Guide, PN 1R120587-0002
- TSQ Duo Hardware Manual, PN 1R120587-0003
- TSQ Duo Spare Parts Guide, PN 1R120587-0004
- TSQ Duo Auto SRM User Guide, PN 1R120587-0005

To view product manuals

Open the desktop folder Manuals.

To open Help

- From the TSQ Series window, choose **Help** > **TSQ Series Help**.
- If available for a specific window or dialog box, click **Help** or press the F1 key for information about setting parameters.

For more information, visit www.thermoscientific.com.

System Requirements

System	Requirements
Hardware	 4.6 GHz processor with 16GB RAM DVD/CD-ROM drive Video card and monitor capable of 1680×1050 resolution 1000 GB hard drive Quad core processor
Software	 Microsoft[™] Windows[™] 7 SP1 Operating System (64-bit) Thermo Foundation 3.0 SP2 (Thermo Scientific software)¹ Thermo Scientific[™] Dionex[™] Chromeleon[™] 7 (release 7.2 SR3 MUa or later)².

Your data system must meet these minimum requirements.

¹Check release notes for compatibility with TSQ Series instrument control software. ²Check release notes for compatibility with Thermo Foundation and TSQ Series instrument control software.

Safety and Special Notices

Make sure you follow the precautionary statements presented in this guide. The safety and other special notices appear in boxes.

Special Notices

Special notices include the following:

IMPORTANT Highlights information necessary to prevent damage to software, loss of data, or invalid test results; or might contain information that is critical for optimal performance of the system.

Note Highlights information of general interest.

Tip Highlights helpful information that can make a task easier.

Safety Symbols and Signal Words

All safety symbols are followed by **WARNING** or **CAUTION**, which indicates the degree of risk for personal injury, instrument damage, or both. Cautions and warnings are following by a descriptor. A **WARNING** is intended to prevent improper actions that *could* cause personal injury. A **CAUTION** is intended to prevent improper actions that *might* cause personal injury or instrument damage. You can find the following safety symbols on your instrument or in this guide.

Symbol	Descriptor
	BIOHAZARD: Indicates that a biohazard <i>will</i> , <i>could</i> , or <i>might</i> occur.
	BURN HAZARD: Alerts you to the presence of a hot surface that <i>could</i> or <i>might</i> cause burn injuries.
	ELECTRICAL SHOCK HAZARD: Indicates that an electrical shock <i>could</i> or <i>might</i> occur.
	FIRE HAZARD: Indicates a risk of fire or flammability <i>could</i> or <i>might</i> occur.
RAMMABLE 2	FLAMMABLE GAS HAZARD: Alerts you to gases that are compressed, liquefied or dissolved under pressure and can ignite on contact with an ignition source. This symbol indicates this risk <i>could</i> or <i>might</i> cause physical injury.
	GLOVES REQUIRED: Indicates that you must wear gloves when performing a task or physical injury <i>could</i> or <i>might</i> occur.
	HAND AND CHEMICAL HAZARD: Indicates that chemical damage or physical injury <i>could</i> or <i>might</i> occur.
	INSTRUMENT DAMAGE: Indicates that damage to the instrument or component <i>might</i> occur. This damage might not be covered under the standard warranty.
	LIFTING HAZARD: Indicates that a physical injury <i>could</i> or <i>might</i> occur if two or more people do not lift an object.
	MATERIAL AND EYE HAZARD: Indicates that eye damage <i>could</i> or <i>might</i> occur.
	RADIOACTIVE HAZARD: Indicates that exposure to radioactive material <i>could</i> or <i>might</i> occur.

Symbol	Descriptor
	READ MANUAL: Alerts you to carefully read your instrument's documentation to ensure your safety and the instrument's operational ability. Failing to carefully read the documentation <i>could</i> or <i>might</i> put you at risk for a physical injury.
	TOXIC SUBSTANCES HAZARD: Indicates that exposure to a toxic substance could occur and that exposure <i>could</i> or <i>might</i> cause personal injury or death.
	For the prevention of personal injury, this general warning symbol precedes the WARNING safety alert word and meets the ISO 3864-2 standard. In the vocabulary of ANSI Z535 signs, this symbol indicates a possible personal injury hazard exists if the instrument is improperly used or if unsafe actions occur. This symbol and another appropriate safety symbol alerts you to an imminent or potential hazard that <i>could cause personal injury</i> .

Hydrogen Safety Precautions

Hydrogen is a colorless, odorless, highly flammable gas with the molecular formula H_2 and an atomic weight of 1.00794, making it the lightest element. Hydrogen gas presents a hazard as it is combustible over a wide range of concentrations: at ambient temperature and pressure, this ranges from about 4% to 74.2% by volume.

Hydrogen has a flash point of - 423 °F (- 253 °C) and an auto-ignition temperature of 1,040 °F (560 °C). It has a very low ignition energy and the highest burning velocity of any gas. If hydrogen is allowed to expand rapidly from high pressure, it can self-ignite. Hydrogen burns with a flame that can be invisible in bright light.



WARNING FIRE HAZARD: The use of hydrogen as a carrier gas is dangerous. Hydrogen is potentially explosive and must be used with extreme care. Any use of hydrogen gas must be reviewed by appropriate health and safety staff and all installations of hydrogen systems must be performed to applicable codes and standards. Thermo Fisher Scientific assumes no liability for the improper use of hydrogen as a carrier gas.

Before you begin using hydrogen, you should conduct a risk assessment based on the quantity of hydrogen to be used and the conditions of your laboratory. You should ask yourself:

"What hydrogen hazards associated with this project are most likely to occur?"

"What hydrogen hazards associated with this project have the potential to result in the worst consequences?"

• Try to reduce or eliminate the higher risks by using the proper ventilation to remove hydrogen gas before an ignitable concentration can accumulate. You should also consider purging the hydrogen to further reduce hazards and ensure anyone who will be working with hydrogen has basic hydrogen safety training.

- As with laboratory safety in general, be sure to wear safety glasses, laboratory coats, gloves, etc. Typically there are no specific requirements for gaseous hydrogen, other than eye protection when working with a compressed gas. If working with liquid (cryogenic) hydrogen, insulated gloves and protective shoes should be worn in addition to eye protection.
- You should post "No Smoking" and "No Open Flames" signs to identify hydrogen sources and cylinders. Maintain, inspect and leak-test all hydrogen sources regularly.
- All hydrogen shutoff valves should be clearly marked and permanent hydrogen piping should be labeled as such at the supply or discharge point and at regular intervals along its length. Where hydrogen gas piping passes through a wall, the piping should be labeled on both sides of the wall.
- There should also be contingency plans in place should an incident occur.
- The site emergency response team, as well as the local fire department, should know the location of all hydrogen storage tanks.

Using Hydrogen with a GC-MS/MS System

To use hydrogen with the mass spectrometer, you must always shut off the GC carrier gas before venting or turning off the mass spectrometer. There are three hydrogen safety screws on the mass spectrometer that **must** be in place. These are attached to your instrument at the factory.

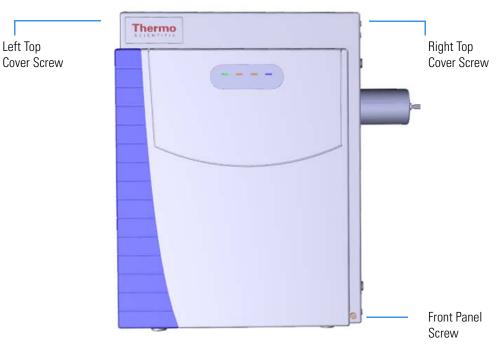


Figure 1. Hydrogen Safety Screws on the Mass Spectrometer

Before powering on the GC-MS/MS system, ensure that:

- All the covers and panels of the GC-MS/MS system are firmly attached.
- The vent valve is tightly closed if you vented the system.
- All fittings, ferrules, and o-rings are sealed.

Hydrogen Connection Guidelines

Use the following guidelines to safely connect hydrogen to your system:

• **Piping**—Hydrogen must be delivered to equipment using appropriate piping and be done in such a way as to pose essentially no hazard to end-users. Piping systems for the delivery of hydrogen should be designed and installed by a person qualified by specific training and experience with hydrogen piping systems.

Stainless steel is usually recommended because it is a safe, cost-effective material. Piping of *black iron* or copper must not be used, as the pipe can become brittle with age. Elastomeric/plastic tubing of various plastics and polymers should not be used, unless the tubing is approved for use with hydrogen. If elastomeric/plastic tubing is used for hydrogen gas delivery, the tubing should be tested for hydrogen permeability to minimize leakage.

The hydrogen piping system must be flexible enough to endure routine thermal expansion and contraction. The system should also include considerations for the most severe condition of temperature and pressure expected during service. Piping and supports must be able to withstand static loading introduced by such things as ice and snow; and dynamic loading from high wind and earthquake.

Caution should be used if burying hydrogen piping. Proper controls should be used to protect against damage and corrosion, and also to prevent Hydrogen from entering a building if there is any leakage.

• **Fittings**—All fittings must be of the proper type approved or designed for use with hydrogen gas. Use as few fittings as possible to minimize the potential for leaks. After installation, ensure that leak testing is carried out prior to system use, and on a regular basis.

There must be no PTFE tape or other things like *plumber's putty* used to enhance a seal, as this actually is a detriment to a good seal. Ideally the best installation would use stainless steel tubing with appropriate gas-tight fittings.

Welding is usually preferred for joints in hydrogen piping systems since welding provides a better connection and reduces the potential for leaks compared to mechanical fittings. Soft solder joints are not permitted for hydrogen systems (due to the low melting point of soft solder and its potential for brittle failure at cryogenic temperatures). Brazed joints are permitted, but such joints should be protected against the possibility of external fire.

Tubing connections should be clamped to barbed or press-fit type connections. Hose clamps or *jubilee clamps* must not be used.

• **Valves**—All valves must be suitable for hydrogen service and for the specific operating conditions. Valves, including regulators, must not be used for hydrogen, unless they are designed and identified for such a use. Ball valves are often chosen because of their superior leak tightness through the valve seat. Pneumatic operators are usually chosen for remotely operated valves so that potential ignition sources (electricity) are remote from the valve.

Manual shutoff valves should be provided near each point of use, within immediate reach. If a hydrogen cylinder or hydrogen generation system is located within immediate reach, a separate point-of-use shutoff valve is usually not necessary.

Line regulators that have their source away from the point of use should have a manual shutoff valve near the point of use.

An emergency gas shutoff device in an accessible location outside the use area should be provided in addition to the manual point-of-use valve in each educational and instructional laboratory space that has a piped gas supply system.

If necessary, the piping system should have uninterruptible pressure relief. The pressure relief system should be designed to provide a discharge rate sufficient to avoid further pressure increase and should vent to a safe location outside or to a ventilation system exhaust.

Purchasing Hydrogen

Use the following guidelines when purchasing hydrogen:

• **Hydrogen Generator**—Because it minimizes the amount of hydrogen present and reduces the degree of hazard, a hydrogen generator (also called an electrolyzer) is the safest way to purchase hydrogen in the quantity used in GC/MS.

However, to minimize the degree of hazard, the hydrogen generator must only be operated in a non-explosive environment because hydrogen buildup can be ignitable. This means that your ventilation system for the room or lab hood must maintain an air exchange rate that is at least two orders of magnitude greater than the maximum hydrogen production rate of the hydrogen generator. Be sure to follow the manufacturers' directions about proper use and maintenance of the regulator.

To prevent the possibility of releasing hydrogen, the hydrogen generator should be set to shut down if:

- There is a loss of flow to the ventilation system
- A hydrogen detector alarms at 25% of the lower flammable limit of hydrogen in air.

The oxygen exhausted by the electrolyzer should be vented to the outside as well.

• **Hydrogen Cylinder**—Hydrogen can be delivered in standard laboratory gas bottles or cylinders. These cylinders have a limited amount of hydrogen in them and are a safe way to transport and store hydrogen. However, compressed hydrogen gas cylinders, like all compressed gas cylinders, must be secured in an upright position, ideally with a non-combustible chain or cable. If the cylinder falls over, the valve can be knocked off and the pressurized cylinder can take off like a rocket, which leads to the release of hydrogen and possibly an explosion, severe injury, or death. Never crack a hydrogen cylinder valve to remove dust or dirt from fittings prior to attaching a regulator, as there is a risk of self-ignition.

Properly Storing Hydrogen

Storing and handling compressed hydrogen gas and cryogenic liquid hydrogen present potential health and safety hazards. Using proper storage and handling techniques is essential to maintaining a safe work environment.

Use the following guidelines when storing hydrogen:

- Store spare hydrogen gas cylinders outside and away from doors, windows, building air intake vents, structures, and vehicle routes. This precaution applies when the hydrogen is or is not in use. Indoor storage of spare hydrogen cylinders has special requirements, which is beyond the scope of this document. Documentation for each vessel should include a description of the vessel, a list of available drawings or other documents, the most recent inspection results, and the responsible person's name.
- Prevent spare cylinders from toppling by wrapping them with chains. The chains should also be protected against corrosion and excessive heat.
- Separate spare hydrogen cylinders from oxidizing gases (such as oxygen) with a 5 ft (1.5 m) tall fire barrier with a half-hour fire rating or place the cylinders at least 20 ft (6 m) apart.
- When moving hydrogen cylinders:
 - Remove the regulator and replace the cylinder valve cap before moving.
 - Move cylinders on cylinder carts or with other appropriate transport devices.
 - Never roll or drop a cylinder and never lift a cylinder by its protective cap.
- Bulk hydrogen systems include either gaseous or liquid hydrogen in fixed installations; in some gas systems a semi-permanent trailer (tube trailer) can be used. Storage vessels for compressed hydrogen gas or liquid hydrogen should be designed, constructed, tested, and maintained in accordance with applicable codes and standards. Bulk hydrogen systems represent a level of complexity again which is beyond the scope of this document; however some general guidelines are provided.

- The bulk hydrogen storage system should not be located beneath electric power lines, close to other flammable gases/liquids, or close to public areas. It should be readily accessible to authorized personnel and delivery equipment, but protected from physical damage or tampering.
- As liquid hydrogen systems also have a cryogenic hazard, additional safety considerations for the use of cryogenic liquids might be necessary.

Hydrogen Safety Codes, Standards and References

The following list of safety codes, standards and references is in no way an exhaustive list. In fact, there might be federal, state or local codes that apply to your specific location. Check with all appropriate agencies with jurisdiction before installing or using a hydrogen system.

- Air Products Safetygram #4 Gaseous Hydrogen
- ANSI/AIAA standard for hydrogen safety guidelines is AIAA G-095-2004, Guide to Safety of Hydrogen and Hydrogen Systems
- ASME B31.1, Power Piping Code
- ASME B31.3, Process Piping Code
- ASME B31.8, Gas Transmission and Distribution Systems
- BCGA Code Of Practice CP4 Industrial Gas Cylinder Manifolds and Gas Distribution Pipework
- BCGA Code Of Practice CP33 The Bulk Storage of Gaseous Hydrogen at Users' Premises
- CGA G-5, Hydrogen
- CGA G-5.4, Standard for Hydrogen Piping Systems at Consumer Locations
- CGA G-5.5, Hydrogen Vent Systems
- CGA G-5.6, Hydrogen Pipeline Systems
- CGA G-5.8, High Pressure Hydrogen Piping Systems at Consumer Locations.
- FM Global Property Loss Prevention Data Sheets 7-50: Compressed Gases in Cylinders
- FM Global Property Loss Prevention Data Sheets 7-91: Hydrogen
- IGC Doc 121/04/E, Hydrogen Transportation Pipelines System Design Features
- NASA
- NSS 1740.16 Safety Standard For Hydrogen And Hydrogen Systems Guidelines for Hydrogen System Design, Materials Selection, Operations, Storage, and Transportation
- NFPA 52, Vehicular Fuel Systems Code
- NFPA 55, Standard for the Storage, Use, and Handling of Compressed Gases and Cryogenic Fluids in Portable and Stationary Containers, Cylinders, and Tanks, 2005 Edition
- NFPA 68, Standard on Explosion Protection by Deflagration Venting
- NFPA 70, National Electrical Code

- NFPA 497, Recommended Practice for the Classification of Flammable Liquids, Gases, or Vapors and of Hazardous (Classified) Locations for Electrical Installations in Chemical Process Areas
- NFPA 13, Standard for the Installation of Sprinkler Systems
- NFPA 45, Standard on Fire Protection for Laboratories Using Chemicals
- NFPA 55, Standard for the Storage, Use, and Handling of Compressed Gases and Cryogenic Fluids in Portable and Stationary Containers, Cylinders, and Tanks
- NFPA 68, 2007 Standard on Explosion Protection by Deflagration Venting
- NFPA 69, Standard on Explosion Prevention Systems
- NFPA 91, Standard for Exhaust Systems for Air Conveying of Vapors
- NFPA 255, Standard Method of Test of Surface Burning Characteristics of Building Materials
- OSHA 29CFR1910.103 1910.103 Hydrogen

Hazardous Substances Precautions



WARNING Before using hazardous substances (toxic, harmful, and so on), please read the hazard indications and information reported in the applicable Material Safety Data Sheet (MSDS). Use personal protective equipment according to the safety requirements.

Biological Hazard Warning Note

In laboratories where samples with potential biological hazards are handled, the user must label any equipment or parts which might become contaminated with biohazardous material.



The appropriate warning labels are included with the shipment of the instrument. It is the user's responsibility to label the relevant parts of the equipment.

When working with biohazardous materials, you are responsible for fulfilling the following mandatory requirements:

- Providing instructions on how to safely handle biohazardous material.
- Training operators to be aware of potential hazards.

- Providing personal protective equipment.
- Providing instructions for what to do if operators are exposed to aerosols or vapors during normal operation (within the intended use of the equipment) or in case of single fault situations such as a broken vial. The protective measures must consider potential contact with the skin, mouth, nose (respiratory organs), and eyes.
- Providing instructions for decontamination and safe disposal of relevant parts.



WARNING The user or operator is responsible for the safe handling of hazardous chemicals or biological compounds including (but not limited to) bacterial or viral samples and the associated waste, according to international and local regulations.

Venting Toxic Gases

When analyzing toxic compounds, be aware that during the normal operation of the GC some of the sample might be vented outside the instrument through the split and purge flow vents; therefore, be sure to vent the exhaust gases to a fume hood. Consult local environmental and safety regulations for instructions in exhausting fumes from your system.

Contacting Us

There are several ways to contact Thermo Fisher Scientific for the information you need.

* To find out more about our products

Go to www.thermoscientific.com/en/products/mass-spectrometry.html for information about our products.

✤ To get local contact information for sales or service

Go to www.thermoscientific.com/en/support-landing/support.html.

- * To suggest changes to documentation or to Help
 - Fill out a reader survey online at www.surveymonkey.com/s/PQM6P62.
 - Send an e-mail message to the Technical Publications Editor at techpubs-austin@thermofisher.com.

Introduction

Use the information in this chapter to determine whether your TSQ Duo system is working properly and to check its basic systems.

IMPORTANT You will likely want to change the GC column before setting up a method. See Chapter 2, "Changing the Column," for instructions on changing the column.

Contents

- Confirming Your Instrument is Working
- Checking Power to the System
- Verifying the Carrier Gas Flow Rate
- Checking Your Carrier Gas Tank Pressure
- Verify Collision Gas Tank Pressure
- Checking the Vacuum and Temperature
- Cleaning the Exterior of Your Instrument
- Configuring Your Instrument

Confirming Your Instrument is Working

After installing a new column, confirm that your GC/MS-MS system has power, the carrier gas flow and collision gas delivery pressure are correct, the gas tanks have enough pressure, and the system is leak-free and has reached vacuum and temperature.

Checking Power to the System

To confirm that the TSQ Duo system is powered on, check that the power light on the front panel is lit and solid green. See Figure 1. If it is not, the instrument is not powered on.

✤ To power on the TSQ Duo instrument

Lift up the power switch located on the upper left side panel of the instrument near the back.



Figure 1. Front Panel of the TSQ Duo Instrument

♦ To power on the TRACE 1300/1310 GC

Reach over the top right of the instrument and pull up on the large plastic ribbed power switch on the back.

To confirm that a TRACE 1300 is powered on

Confirm that the power light on the status panel is solid green.

* To confirm that a TRACE 1310 is powered on

Confirm that the touchscreen main menu has appeared.

Verifying the Carrier Gas Flow Rate

Once you confirm that the system is powered on, verify that the carrier gas rate is correct.

* To check the carrier gas flow rate on the TRACE 1310 GC

- 1. Choose Instrument Control and then Front/Back inlet.
- 2. Display the column flow.

If the actual and set point amounts in **Col. Flow** are the same, then you have good carrier gas flow. If the amounts are different, refer the Troubleshooting section of the *TRACE 1300/1310 GC Series User Guide*.

- ***** To check the carrier gas flow rate on a TRACE 1300
- 1. Go to Start > Thermo Chromeleon > Chromeleon and open the Chromeleon Console.

Command

- 2. Click Instruments in the left-hand menu.
- 3. Select **Command** from the top menu.
- 4. A status page for all the configured instruments.

Figure 2. Checking Instrument Status in Chromeleon Software

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Instruments

5. Verify the column flow rate beside in the status pane.

Checking Your Carrier Gas Tank Pressure

Make sure you have enough pressure in the carrier gas tank to accommodate the number of samples you plan to run. If the pressure is too low, you might run out of gas in the middle of a run, which could compromise the results of your data.

* To check your carrier gas tank pressure

- 1. Locate your carrier gas tank. It might be in a different room, depending on how your lab is set up.
- 2. Look at the pressure gauge on the tank.
- 3. Ensure the pressure is more than 100 psi at the primary (or first) regulator stage. If it is not, you may want to replace the tank if you have to run a lot of samples.

Verify Collision Gas Tank Pressure

Check that the input pressure of your collision gas is at 59–61 psig for Argon or 56–58 psig for Nitrogen and that the tank is connected to the mass spectrometer.

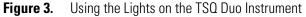


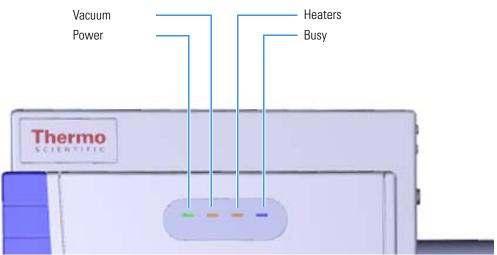
CAUTION Collision gas input pressure must remain constant for proper instrument performance. The regulator used to supply the collision gas must be able to deliver 60 ± 1 psig. Ensure that the regulator is marked clearly at 60 psig and is stable enough to supply constant pressure at 60 ± 1 psig.

Checking the Vacuum and Temperature

Use the lights on the front of your TSQ Duo system to check the vacuum and temperature of the instrument.

To check the vacuum, look at the Vacuum light. See Figure 3. When the light is a solid green, the mass spectrometer is under sufficient vacuum. If the Vacuum light is slowly blinking orange, you have not yet achieved vacuum. If the Vacuum light is blinking orange quickly, you have a large leak that has prevented the instrument from achieving vacuum. In this case, you must turn the power off and find and fix the leak. Most likely, the column nut must be tightened, the column was not installed correctly, or the vent valve was not completely closed. See Troubleshooting for more information.





To check the temperature, look at the Heaters light. When the Heaters light is a solid green, the GC-MS/MS system is at temperature. If the Heaters light is blinking orange, the ion source, ion optics and/or transfer line are not at temperature. If the Heaters light is not lit, the heaters are not turned on.

Note Until the Vacuum light is a solid green (high vacuum is achieved), the heaters will not power on and the Heaters light will not be lit.

You may also use the TSQ Series Dashboard to check the vacuum of your system. If your system has achieved sufficient vacuum, a green check mark appears next to **Vacuum** on the dashboard.

-		
TSQ Series - TSQ Duo I	Dashboard C	
Ai	ir/Water Spectrum	
Calib	bration Gas Spectrum	
	Custom Spectrum	
K	Auto Tune	
×	View Tune Report	
J.	Tune Types	
K	Manual Tune	
	AutoSRM	
Status Analyzer Power	Maintenance	
TSQ Duo		
Status: Idle		
Act	tual Set-Point	
✓ MS transfer line temp.:	250 °C 250 °C	
✓ Ion source temp.:	200 °C 200 °C	
✓ Vacuum:	ок	
Foreline pressure:	76 mTorr	Vacuum
lon gauge pressure:	Gauge not present	Vaduuri
Turbo-pump speed:	100 %	
Collision gas on:	No	
CI reagent gas flow:	0.0 mL/min 0.0 mL/min	
Instrument Control	Shut Down	

Figure 4. Checking System Vacuum on the Dashboard

Cleaning the Exterior of Your Instrument

When the exterior of your instrument gets dirty, wipe it with a clean, dry, lint-free cloth.

Configuring Your Instrument

Initially, the field service engineer will configure your TSQ Duo instrument. However, if you have reinstalled the instrument's software or you have a new computer or device, follow these instructions to reconfigure it.

✤ To configure your TSQ Duo instrument

- 1. From the Start menu on your computer desktop, choose **Start | All Programs | Thermo Chromeleon 7 | Services Manager**.
- 2. The Chromeleon Services Manager window opens. See Figure 5.

Figure 5. Chromeleon Services Manager

🗞 Chromeleon Services Manager	? ×
Instrument Controller Service The local Instrument Controller is running idle.	Stop Instrument Controller Image: Start service on system start Configure instruments
Other Chromeleon services All other Chromeleon services are running.	

- 3. Click **Configure Instruments**. The Chromeleon Instrument Configuration Manager opens. See Figure 6.
 - Figure 6. Finding Available Devices

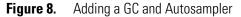
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For Help, press F1	OK

4. Right-click the PC icon and select Add Instrument. See Figure 7.

How to			
			Messages Instrument Controller TSQ-PC [Expert]
Undo	Ctrl+Z	1 A	Instrument Controller TSO-PC connected. 1:23:00 PM User TSO (from TSO-PC) has connected Chromeleon Instrument Configuration Manager to this controller.
Cut	Ctrl+X	· ·	1:24:28 PM CmDDKDrv - Chromeleon DDK Host Driver, Version 7:2 SR1 Build 6772 (RC) Copyright © 2009-2014 Thermo Fisher Scien rights reserved.
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Disconnect			2002-014 Themo Paner adentino inc. All right reserved.
Add Instrument		Adds a new instrument to controller	
Add Sharable Interface			
Add Module			
Rename	F2		
Properties	Enter		

Figure 7. Adding an Instrument

 The Add Module to Instrument dialog box opens. Under Manufacturers, select Thermo Scientific > GC: Modules to add your system's gas chromatograph and autosampler. See Figure 8



Sinstrument Configuration - Chromeleon Instrument Configur	ation Manager			E
File Edit View Controller Help				
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		OK Cancel	/	a

- 6. Click OK.
- 7. To add the mass spectrometer, right-click the PC icon again and select Add Instrument. The Add Module to Instrument dialog box opens. Under Manufacturers, select Thermo Scientific > Mass Spectrometry. Select Mass Spectrometer from the Modules list. See Figure 9.

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			DKCancel		

Figure 9. Adding the Mass Spectrometer

- 8. Click OK.
- 9. Click **Mass Spectrometer** on the left menu. The **Mass Spectrometer Configuration** page opens. On the **General** page, set the pressure units.

Figure 10. Setting the Pressure Units

Instrument Configuration - Chromeleon Instrument Configuration I	lanager 😐 😐 😐
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Source 1 Source 1	Messages Instrument Controller TSO-PC [Exper]
For Help, press F1	OK

Note You only need to set up the pressure units if you have an ion gauge or convectron gauge installed on your system. The readbacks from these components will display in the units set in this dialog box.

10. Set the remote start, which tells the TSQ Duo system when the GC has started a run. When you configure the GC, you can tell it what to send out to the instrument. Make sure the value in this dialog box matches what you set on the GC. The default is **Active Low**. See Figure 11.

Figure 11. Setting the Remote Start

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[92.168.0.150] (Front, Back) npler }]	Device Name MSDevice	Device Type TSQ Series v Simulation File v	;]]
	General Maintenance Interva Pressure units: Remote start: TSQ Series:	Hardware Inject Synchronization	ermo Fisher Scie

11. Check **Always Show Method Portability Between Instrument Methods** if you need to create methods for older model TSQ 8000 or TSQ 8000 Evo systems.

Figure 12. Enabling Method Portability

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68.0.150] (Front, Back)	General					n Manager to this co Copyright © 2009-201
	Device Name MSD	evice	Device Type	TSQ Series	-	
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	, , , , , , , , , , , , , , , , , , ,	Always show method po	rtability between instru	Iment models.		
		Commun	ications			
				-		

12. Click **Communications** to reset the network IP address and assign a TCP port. See Figure 13. This tells the instrument method how to find the TSQ Duo system if you changed the IP address using Instrument Configuration. For security purposes, you may also want to modify the TCP port. Consult your local IT Department for help.

Figure 13. Setting up the Network

Communications	X
Network address: TCP port:	192 168 1 234 30
ОК	Cancel

13. Click the **Maintenance Intervals** tab to set the number of days until you plan to perform maintenance on certain components of your GC/MS system. See Figure 14.

Figure 14. Setting the Maintenance Intervals

.150] (Front, Back) General			n Manager to this o Copyright © 2009-20
Device	Name MSDevice	Device Type TSQ Series	
🗖 Sin	nulate	Simulation File	_
Ter	mporarily used in other application	Hardware Inject Synchronization	ermo Fisher Scienti
	Read	Configuration	
	General Maintenance Interval	s	
	Enabl	e Days	
	Foreline pump oil:	125	
	Fill calibration	365	
	Turbo pump oil/bearing:	520	
	Filament 1:	7 1,000	
	Filament 2:		
	Ion source:		
	Multiplier:	1.000	
	Reset All		

- 14. Select the Foreline Pump Oil check box to enable the maintenance intervals. Then set the number of days after which you want to be reminded to check the oil. The manufacturer recommends changing the oil every 125 days. Select the Turbo Pump Oil/Bearing check box to enable the maintenance reminder. Then set the number of days after which you want to be reminded to check the oil. Refer to the Turbo Pump manual for the manufacturer's recommended maintenance intervals.
- 15. Select the **Filament 1** check box to enable the maintenance reminder. In a leak-free system, expect the filament to last between 30-360 days, depending on usage.

- 16. Select the **Filament 2** check box to enable the maintenance reminder. Then set the number of days after which you want to be reminded to check filament 2. In a leak-free system, expect the filament to last between 30-360 days, depending on usage.
- 17. Select the **Ion source** check box to enable the maintenance reminder. Then set the number of days after which you want to be reminded to check the ion source. The time between cleaning depends very strongly on your analysis. You will have to determine the correct length of time between source cleanings.
- 18. Select the **Multiplier** check box to enable the maintenance reminder. Then set the number of days after which you want to be reminded to check the electron multiplier.
- 19. Click OK to return to the main Instrument Configuration home page.
- 20. Select **TRACE 1300 Series GC II** from the left side menu to configure the GC handshaking parameters.
- 21. Under the Signals tab, set the handshaking parameters as shown in Table 1.

Remote Start In	High to Low
Inhibit Ready In	When High
End of Run Out	High to Low
Start of Run Out	High to Low
GC Ready Out	When Low

Table 1. GC Handshaking Parameters

	A Instrument Controller TSQ-PC		instrument Cor	ומטוופר ו-שע-איט ובאויפרון
Baçki	1:23:00 PM User TSQ (from TS		omeleon Instrument	5770 (DO) 0
TRACE 1300 Series Cont	iguration			
General Instrument Signals Auxiliary Hea	Oven Front Inlet Back Inlet Front Inle	ont Detector Back Dete	ctor Auxiliary Carrie	ers Valves & Events
Remote Start In:	High -> Low			
End Of Run Out:	High -> Low			
Inhibit Ready:	None			
GC Ready Out:	When Low 💌			
Start Of Run Out:	High -> Low 💌			
Prep-Run Out:	When Low 💌			
Get			ОК	Cancel Help
	1			

Figure 15. GC Handshaking Parameter Configuration

22. Click OK once you have entered the correct GC handshaking parameters.

Note For the remainder of the GC configuration settings, refer to the *TRACE 1300* and *TRACE 1310 Series GC User Guide*. Refer to your autosampler user documentation for the correct autosampler configuration settings.

- 23. Close the Instrument Configuration window.
- 24. Check the status of the configured instruments in the **Status Pane** of the TSQ Series Dashboard.

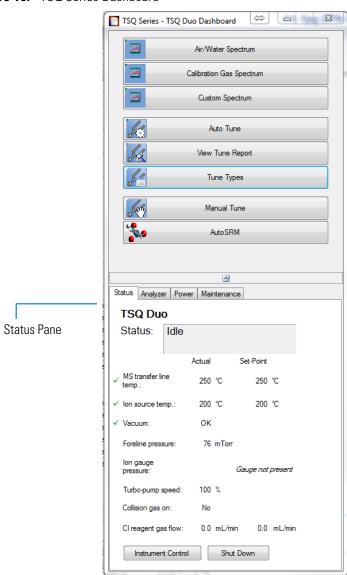


Figure 16. TSQ Series Dashboard

2

Changing the Column

The TSQ Duo GC-MS/MS system ships with a factory-tested 15 m \times 0.25 mm ID TG-SQC column, which the field service engineer uses to qualify the instrument. This column is for system qualification purposes only and not for regular testing and should be replaced. For best results, choose a column that will give you the best possible resolution, analysis speed, and quantitation.

Contents

- Determining the Column Type
- Replacing the Factory Installed Column
- Connecting the Column to the Transfer Line

Determining the Column Type

When determining the type of column for your particular needs, here are a few things to consider:

• **Column Material**—Columns made out of fused silica are economical and widely used. Columns made out of this material have a wide range of stationary phases and are available in many sizes that can be used with a mass spectrometer.

Large diameter columns made of steel are widely used in process gas analysis, but they are not typically used on mass spectrometers. There are also metal-clad, fused silica columns, which have the advantages of fused silica, but the metal makes them resistant to breakage. These columns are less common and more expensive.

• **Stationary Phase**—The stationary phase is the most important consideration when selecting a column. The interaction between the stationary phase and the analyte determines how well the analytes separate from each other (resolution) and also affects how quickly the separation occurs (analysis time). Choose a stationary phase that is compatible with the nature of your analytes and the maximum GC oven temperature that you will use.

- Internal Diameter—The smaller the diameter of the column, the better the separation. However, smaller diameter columns do not have as much capacity for matrix or analytes. As a result, smaller diameter columns are subject to overloading, which leads to retention time shifts and peak shape changes. Larger diameter columns can accept larger concentrations of material, but will require longer columns or slower GC oven temperature ramps (which increase the analysis time) to match the separation power of smaller-diameter columns. Typical column sizes for GC/MS have inside diameters (ID) of 0.25 mm. Smaller ID columns, such as the 0.18 and 0.10 mm, are becoming increasingly popular. Additionally, 0.32 and 0.53 mm ID columns are commonly used.
- **Film Thickness**—With larger film thicknesses, there is more capacity for the analyte. This capacity can aid in the separation of high concentration samples and in the separation of very volatile samples because thicker stationary phases allow more opportunities for the analytes to interact with the stationary phase. The optimal film thickness depends on the internal diameter of the column, and the maximum total capacity needed for sample analysis.

Thick films with small internal diameters give very strong interactions with the analytes, which can result in longer analysis time and peak tailing. Large ID columns with thin films have very little interaction with the analytes, which will result in very fast analysis times with little separation. Typical film thicknesses are 0.25 μ m for a column with an ID of 0.25 mm. Other common film thicknesses are 0.1, 0.5, and 1.0 μ m.

Be careful when selecting columns for mass spectrometers. Some columns with large inner diameters that work fine with other GC detectors may need lower head pressure when operated with vacuum at the outlet. This lower head pressure can allow air diffusion into the column through the carrier gas flow module's split and purge valves.

• Length—The length of the column affects how much time the analyte has to interact with the stationary phase. Longer columns typically have better resolutions and higher capacities, but longer analysis times. Longer columns are also more expensive. Typical column lengths are 15 or 30 m for GC/MS, but 100 m columns may be needed for very complex mixtures like gasoline. Very short columns (2.5, 5, and 10 m) are also available.

Tip The Thermo Scientific[™] TG-5MS column is suitable for many applications.

Note Contact your local sales representative to order a Thermo Scientific column. You can also refer to our catalog or visit our Web site at www.thermoscientific.com/columns.

Replacing the Factory Installed Column

***** To replace the factory-installed column in the TSQ Duo system

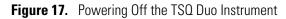
Note If you are running samples, stop the acquisition before powering off the system.

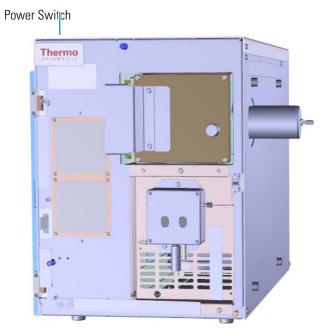
1. Cool down the GC oven and injector. See the GC documentation for information.

2. Open the TSQ Series Dashboard and click Shut Down.

During the shutdown procedure the vacuum and heaters lights will remain off. Once the procedure is complete and the instrument is ready to be powered off, the power light will turn amber and start blinking rapidly. At this point it is safe to power off the TSQ Duo system.

3. On the left side of the instrument, push down on the power switch to power-off the TSQ Duo system.





- 4. Open the front door of the instrument.
- 5. Turn the vent knob counterclockwise to vent the system.
- 6. Wait 5 minutes for the instrument to vent.
- 7. Remove the current column:
 - a. Make sure the heated zones of the GC are cooled down. Refer to the GC documentation for instructions.
 - b. Turn off the carrier gas and if used, the detector gas. See the GC documentation for information about using detector gases.
 - c. Open the front door of the GC.



WARNING BURN HAZARD: The injector, oven, and transfer line may be hot. Allow them to cool to room temperature before touching them.

- d. Unscrew the transfer line nut and remove the column from the transfer line.
- e. Unscrew the injector nut and remove the column.
- f. Remove the column from the column rack and from the GC.
- 8. Connect the new column to the injector inside the GC.

Note Wear clean, lint- and powder-free gloves when you handle the column and injector ferrule.

- a. Unwind the column enough to easily connect its ends to the injector and detector.
- b. Wipe about 100 mm (4 in.) of the column with a tissue soaked in methanol.
- c. Insert the column through the injector retaining nut and ferrule (larger end up). If the M4 retaining nut is used, slide it on the column through the side cut. Wipe the column again with a tissue soaked in methanol.

Tip Slide a notched septum on the column before the injector retaining nut to make it easier to measure the proper distance between the nut and end of the column.

- d. Use a scoring wafer to score and break the column about 1 cm (0.4 in.) from the end. Use a magnifying glass to check for an even, flat cut. Repeat if necessary.
- e. Insert a notched septum on the column to hold the retaining nut at this position. Thread the retaining nut into the injector but do not tighten.
- f. Ensure that the end of the column is the proper distance (splitless = 5 mm, split = 10 mm, PTV and PTVBKF = 30 mm) from the back of the injector nut.
- g. Adjust the column position so that the septum contacts the bottom of the retaining nut. Use your fingers to tighten the retaining nut until it starts to grip the column.
- h. Tighten the column nut finger-tight until it starts to grip the column plus a quarter turn.
- i. Remove the notched septum from the column.
- 9. Set up the GC parameters:
 - a. Set the oven and injector temperature to 50 °C (122 °F).
 - b. Set the carrier gas flow to 1.0 mL/min.
 - c. Turn off vacuum compensation, which is located on the Carrier menu of the GC.
 - d. Use the column flowmeter connector to verify that there is flow through the column. If you do not have a flowmeter, dip the column outlet in a small vial of methanol. Bubbles indicate there is flow through the column. If there is no flow, check that the carrier gas is on, the GC inlet is pressurized, and the column is not plugged. If there is still no flow, consult the GC documentation or contact Technical Support.
 - e. Allow the column to purge for at least 10 minutes. If you used methanol to detect column flow, remove column from methanol during purge time.

- f. Insert the column into the fitting of the column flowmeter connector that blocks the column flow.
- 10. Perform a column leak check:
 - a. On the TRACE 1310, select the **Leak Check** icon in the **Maintenance** menu. Otherwise, perform the leak check through the Chromatography Data System. Refer to the *TRACE 1300 and TRACE 1310 Series GC User Guide* for instructions.
 - b. Start the leak check.

The split and purge valves of the selected channel are automatically closed, and the channel is pressurized with carrier gas to the leak check setpoint.

The system monitors the pressure for one minute. If the pressure does not drop more than the maximum allowed sensitivity value, then the leak check will pass. If the leak check does not pass, use the leak detector to find and fix any leaks.

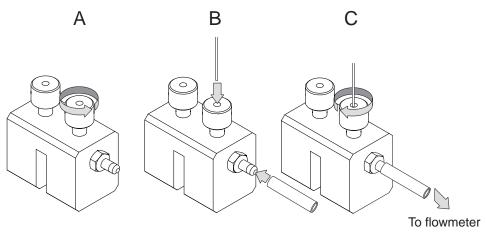
Tip Leaks can be caused by not tightening the fitting on the column flowmeter connector. Check the fitting before looking for the leak elsewhere.



CAUTION INSTRUMENT DAMAGE: Do not allow the column flowmeter connector to exceed 80 $^{\circ}$ C (176 $^{\circ}$ F). Otherwise, it will melt and damage the instrument.

- c. Repeat the leak check until no leaks are indicated.
- 11. Calibrate the carrier gas flow (column evaluation):
 - a. Carefully push the capillary column end into the flowmeter section of the column flowmeter connector.





- b. Connect the flowmeter to the dedicated fitting on the column flowmeter connector.
- c. If you have a TRACE 1310, select the **Back** or **Front Column** icon in the **Configuration** menu. Otherwise, perform the column evaluation through the

Chromatography Data System. See the *TRACE 1300 and TRACE 1310 User Guide* for instructions.

- d. Select **Column** and input the column's physical characteristics.
- e. If a pre-/post column is present, set the length and nominal internal diameter of the pre-/post column in the same valid ranges for the column. The following two lines are added to the menu.

Note For the most reproducible results, you should conduct a more detailed column evaluation. However, the following steps, while recommended, are not required.

- f. Start the column evaluation. According to the physical characteristics of the column, the system calculates and displays the relevant column K-factor. At the end of the routine, a message will indicate that the evaluation was successful.
- g. Expect a K-factor of approximately 0.7 0.9 for a 15 m, 0.25 mm i.d. column (1.3 2.0 for a 30 m, 0.25 mm i.d. column). If the column does not report a K-factor within this range or within 0.1 units of the previous stored value, check for a leak or broken column using the leak detector. The K-factor is a measured resistance for the column. A K-factor that is too low may indicate a leak in the system, while a K-factor that is too high may indicate a blockage.

Fix any issues found and rerun column evaluation until an appropriate K-factor is achieved.e a leak in the system, while a K-factor that is too high might indicate a blockage.

- 12. Disconnect the column flowmeter:
 - a. Disconnect the column from the column flowmeter connector.
 - b. Remove the clear plastic component, including its fittings, from the oven and set them aside.
 - c. Close the GC door.
- 13. Condition the column before inserting it into the TSQ Duo system. Column conditioning consists of passing a carrier gas flow through the column heated to a programmed temperature as described in the column manufacturer's instructions.
 - a. If there are no conditioning instructions, perform the column conditioning by setting a final temperature 10 °C–20 °C below the column's recommended maximum temperature.



CAUTION INSTRUMENT DAMAGE: The material released from the column (column bleed) during conditioning may contaminate the ion source if the column is inserted into the transfer line during the high-temperature stage of conditioning.



WARNING FIRE HAZARD: Do not use hydrogen as the carrier gas for conditioning your column. It could vent into the oven and present an explosion hazard.

b. Run the slow temperature program that is recommended by the manufacturer. A typical program would hold the column at 40 °C (104 °F) for 15 minutes, and then ramp at 10 °C/min (50 °F/min) up to 10–20 °C below the maximum allowed column temperature. Hold the column at this temperature for two hours.



CAUTION INSTRUMENT DAMAGE: Never exceed the column manufacturer's maximum operating temperature.

Connecting the Column to the Transfer Line

When connecting the column to the transfer line, you may use either the spring loaded transfer line nut with the graphite Vespel[™] ferrule or the regular transfer line nut

Note For best results, we recommend you use the spring loaded transfer line nut.

* To connect the column using the spring loaded transfer line nut

Note If you use a graphite Vespel ferrule with your column, Thermo Fisher Scientific recommends using the spring loaded transfer line nut with it. See the spare parts guide for ordering information.

- 1. Lower the oven temperature and allow it to cool.
- 2. If the TSQ Duo system is running, shut down and vent it. See the instrument's hardware manual for instructions.



CAUTION BURN HAZARD: The injector, detectors, oven, and transfer line may be hot. Allow them to cool before touching them.

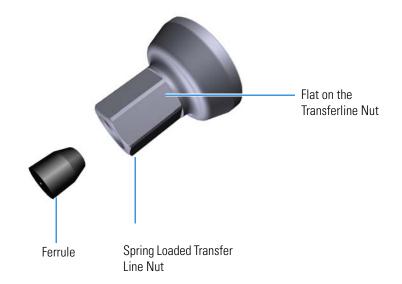
3. Unwind about one turn of the column from the column outlet end.

Note Wear clean, lint- and powder-free gloves when you handle the column and transfer line ferrule.

- 4. Wipe approximately 300 mm (12 in.) of the column with a tissue soaked in methanol.
- 5. Choose an appropriate ferrule for the outer diameter of your column.
- 6. Insert the column through the spring loaded transfer line nut and ferrule, entering through the tapered end of the ferrule.

7. Wipe the column again with a tissue soaked in methanol.

Figure 19. Transfer Line Nut and Graphite Vespel Ferrule Orientation



- 8. Insert the column into the measuring tool (see Figure 20), which is in the MS Toolkit, so that it is even with the lines at the end of the column. Figure 21 indicates proper positioning of the column in the tool for accurate measuring.
- 9. Use a scoring wafer to score and break the column. Use a magnifying glass to check for an even, flat cut. Repeat if necessary.
- 10. Use a 5/16 in. wrench to hold the column measuring tool steady.

Figure 20. Column Measuring Tool



- 11. While holding the column measuring tool steady, tighten the spring loaded transfer line nut with a 1/4" wrench until the column just stops moving in the ferrule.
- 12. Turn the spring loaded transfer line nut 1 flat backward so the column is able to move in the ferrule with slight resistance.
- 13. Line up the outlet of the column with the arrows on the end of the column measuring tool.

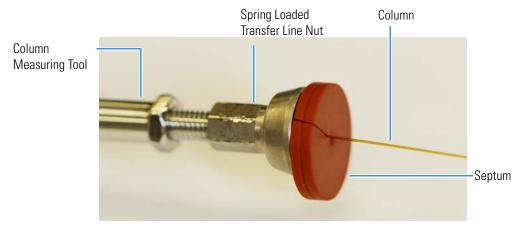


Figure 21. Lining Up the Column in the Column Measuring Tool

14. Place a septum with a notch cut into it behind the transfer line nut. The septum marks the place on the column where it should exit the nut.

15.





16. Pull the column back from the spring loaded transfer line nut. Do not move the septum from its position on the column.

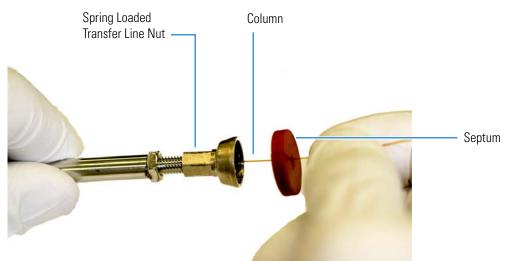


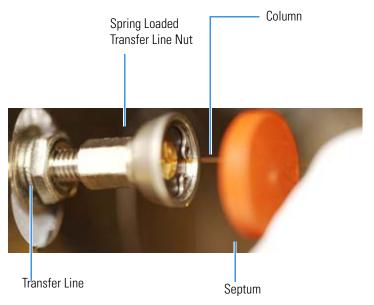
Figure 23. Pulling the Column Back from the Spring Loaded Transfer Line Nut

- 17. Loosen the transfer line nut from the column measuring tool.
- 18. Remove the column, transfer line nut and ferrule from the column measuring tool, making sure not to move the septum from its location on the column.

Note The ferrule should still be able to move on the column. Use the septum to mark the correct location where the column should exit the nut.

19. Insert the column into the transfer line.

Figure 24. Inserting the Column into the Transfer Line



- 20. Tighten the spring loaded transfer line nut until it is just secure enough so that you cannot move it.
- 21. Loosen the nut by turning it exactly 1 flat backward.

22. Position the column in the transfer line using the cut septum to measure the correct length you should insert the column.

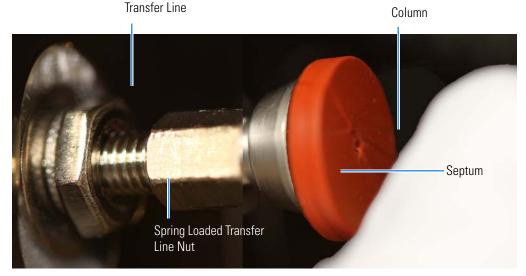


Figure 25. Positioning the Column in the Transfer Line

- 23. Tighten the spring loaded transfer line nut 1 flat forward—back to where it is secure enough in the transfer line that you cannot move it.
- 24. Tighten the spring loaded transfer line nut 1 additional quarter turn.
- 25. Remove the cut septum.
- 26. Condition the graphite Vespel ferrule:
 - a. Raise the oven temperature to the maximum temperature you will operate the GC.
 - b. Wait 10 minutes.
 - c. Lower the oven temperature to 40 °C (104 °F) and allow it to cool before continuing.



WARNING BURN HAZARD: The oven may be hot. Allow it to cool to room temperature before opening it. The injector will still be hot, so do not touch it.

- 27. Close the front door of the GC.
- 28. Restore working conditions.
 - a. Raise the oven temperature to the initial temperature that you will use.
 - b. Turn on vacuum compensation on the GC.
- 29. Power on the TSQ Duo instrument. See Chapter 1, "Introduction," for instructions.
- 30. Once the instrument is pumped down and able to scan, view air water spectra and look for evidence of leaks with a large m/z 28 signal. If you observe a leak, stop scanning and gently tighten the nut in small increments until no leaks appear when scanning.

* To connect the column using the regular transfer line nut

Note For best results, we recommend you use the spring loaded transfer line nut. See "To connect the column using the spring loaded transfer line nut" on page 21.

- 1. Lower the oven temperature and allow it to cool.
- 2. If the TSQ Duo system is running, shut down and vent it. See the instrument's hardware manual for instructions.



CAUTION BURN HAZARD: The injector, detectors, oven, and transfer line may be hot. Allow them to cool before touching them.

3. Unwind about one turn of the column from the column outlet end.

Note Wear clean, lint- and powder-free gloves when you handle the column and transfer line ferrule.

- 4. Wipe approximately 300 mm (12 in.) of the column with a tissue soaked in methanol.
- 5. Choose an appropriate ferrule for the outer diameter of your column.

Note If the maximum oven temperature in your method is ≥ 290 °C (554 °F), Thermo Fisher Scientific recommends using a spring loaded transfer line nut with a graphic Vespel ferrule or a SilTiteTM nut and ferrule. By cycling the oven at and above this temperature, expansion and contraction of the graphite Vespel material can cause leaks in the transfer line.

6. Insert the column through the transfer line nut and ferrule, entering through the tapered end of the ferrule. Wipe the column again with a tissue soaked in methanol.

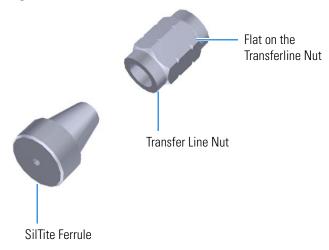


Figure 26. Transfer Line Nut and SilTite Ferrule Orientation

- 7. Insert the column into the measuring tool, which is in the MS Toolkit (See Figure 27), so that it is even with the lines at the end of the column. Figure 28 indicates proper positioning of the column in the tool for accurate measuring.
- 8. Use a scoring wafer to score and break the column. Use a magnifying glass to check for an even, flat cut. Repeat if necessary.
- 9. Use a 5/16 in. wrench to hold the column measuring tool steady.

Figure 27. Column Measuring Tool

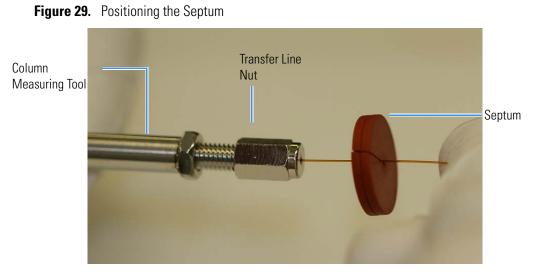


- 10. While holding the column measuring tool steady, tighten the transfer line nut with a 1/4" wrench until the column just stops moving in the ferrule.
- 11. Turn the transfer line nut 1 flat backward so the column is able to move in the ferrule with slight resistance.
- 12. Line up the outlet of the column with the arrows on the end of the column measuring tool.



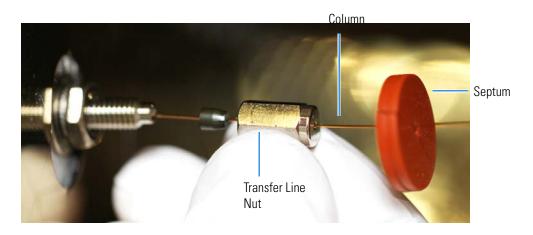
Figure 28. Lining Up the Column in the Column Measuring Tool

13. Place a septum with a notch cut into it behind the transfer line nut. The septum marks the place on the column where it should exit the nut.



14. Pull the column back from the transfer line nut. Do not move the septum from its position on the column.

Figure 30. Pulling the Column Back from the Transfer Line Nut



- 15. Loosen the transfer line nut from the column measuring tool.
- 16. Remove the column, transfer line nut and ferrule from the column measuring tool, making sure not to move the septum from its location on the column.
- 17. Insert the column into the transfer line.
- 18.

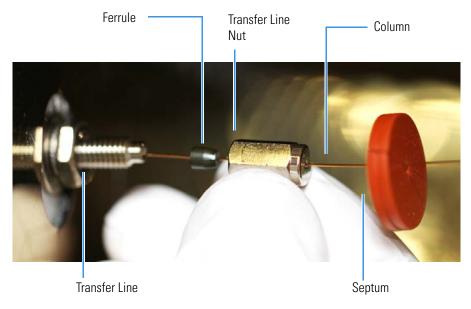
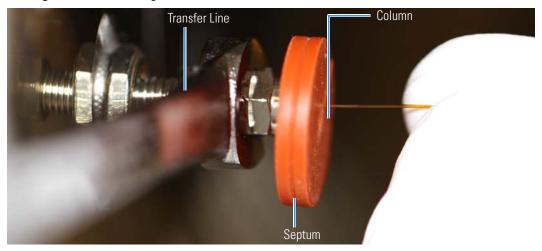


Figure 31. Inserting the Column into the Transfer Line

- 19. Tighten the transfer line nut until it is just secure enough so that you cannot move it.
- 20. Loosen the nut by turning it exactly 1 flat backward.
- 21. Position the column in the transfer line. Use the septum as a guide to measure the correct length you should insert the column. Be careful not to change the location of the septum on the column.

Figure 32. Positioning the Column in the Transfer Line



- 22. Tighten the nut 1 flat forward—back to where it is secure enough in the transfer line that you cannot move it.
- 23. Tighten the nut 1 additional quarter turn.
- 24. Remove the cut septum.

Note If you are using a SilTite ferrule, follow the instructions that come with SilTite ferrules. If you are using a graphite Vespel ferrule, they require conditioning to ensure a leak-tight seal. See the spare parts guide for information about ordering these ferrules.

- 25. Condition the graphite Vespel ferrule:
 - a. Raise the oven temperature to the maximum temperature you will operate the GC.
 - b. Wait 10 minutes.
 - c. Lower the oven temperature to 40 °C (104 °F) and allow it to cool before continuing.



WARNING BURN HAZARD: The oven may be hot. Allow it to cool to room temperature before opening it. The injector will still be hot, so do not touch it.

- d. Retighten the transfer line nut.
- 26. Close the front door of the GC.
- 27. Restore working conditions.
 - a. Raise the oven temperature to the initial temperature that you will use.
 - b. Turn on vacuum compensation on the GC.
 - c. Power on the instrument. See Chapter 1, "Introduction," for instructions.
 - d. Once the instrument is pumped own and able to scan, view air water spectra and look for evidence of leaks with a large m/z 28 signal. If you observe a leak, stop scanning and gently tighten the nut in small increments until no leaks appear when scanning.

Tuning

Tuning will improve the performance of your TSQ Duo system. For optimum stability, you must wait until the power, vacuum, and heaters lights on the front of the instrument are a solid green. These lights indicate that the instrument has reached vacuum and that it is at the last set temperature. If the system has been powered off for a period of time (that is, a cold system), the system components take up to 4 hours to thermally stabilize after reaching the temperature setpoint. If you did not vent the instrument (that is, the system is hot), the components take approximately 10 minutes to thermally stabilize after reaching the temperature setpoint.

Contents

- Accessing Auto Tune
- Tune Types
- Tuning the Mass Spectrometer
- Updating Tunes for New RF Lens

You can tune the TSQ Duo system by using the TSQ Series Dashboard.

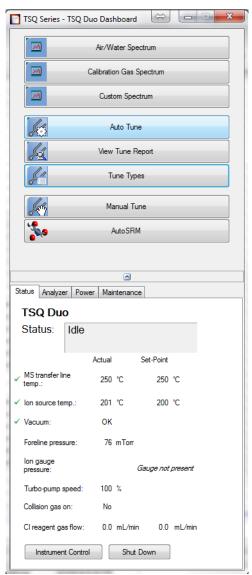
IMPORTANT Be sure to give the MS system enough time to stabilize. Otherwise, you may see mass drift, mass spectral changes, or changes in the fragmentation of your data.

Note If you are running samples with heavy matrix, running the samples before the source has had time to stabilize at high temperature will prematurely dirty the ion volume and optics.

Accessing Auto Tune

- To access TSQ Series Auto Tune
- Double-click the TSQ Series Dashboard shortcut on your desktop to open the TSQ Series Dashboard and then click Auto Tune. See Figure 33.

Figure 33. TSQ Series Dashboard



Tune Types

This section describes the available tune types for the mass spectrometer.

EI Initial Tune EI Standard Tune EI Standard Quick Tune EI SRM Tune EI SRM Quick Tune EI Tune Check EI Diagnostics Only EI Target Tune Fast Scan Tune Negative CI Tune Positive CI Tune

El Initial Tune

EI Initial Tune—Creates a default tune file. It requires a clean instrument, and it sets the repeller to 0 V and the quadrupole voltages to low values. This tune is used to reset parameters for all the other tunes after cleaning the ion source. As a result, use this tune only when the ion source is clean. The EI initial tune should also be used when changing from an SRM tune to a standard tune. The EI initial tune has higher resolution and lower sensitivity than the EI standard tune. This tune starts with the tune file stored in the instrument at the factory. It then performs a mass calibration and leak check, sets the repeller to 0 V, and tunes the lenses. The quadrupole offset voltage is set to a low value to improve resolution, which is also tuned. The detector gain is calibrated to generate 300,000 electrons for every ion that strikes the detector. Additionally, this tune generates spectra that are the closest in appearance to the factory tune.

El Standard Tune

EI Standard Tune—Provides EI tuning and is used to completely retune the system. It takes the longest amount of time to run, but it has the advantage of reoptimizing nearly all the parameters affecting the signal. This type of tune performs a mass calibration, tunes the lenses and resolution, and performs a leak check. It adjusts the detector sensitivity to generate a m/z 219 ion with an intensity of 20,000,000 counts. Unless your SOP requires it, this is not the best tune to use on a daily basis because of the length of time required to run it.

Figure 34 shows a typical tune report for an EI Standard Tune on a system using helium as a carrier gas.

						Ins	trument Name: TSQ S	Series
une Comple	ted - FriOct	21 16:09:49	2016			Ins	trument ID: 10099200	00
une File: A	utoTune_El_	2016-10-21-	16-09-49.tsq8tune			Us	er: TSQ8141213	
une Type:	El Standard	Tune (built-i	n)					
starting Tune	File: (I a	st Saved Tun	e)					
raiting tune	(La		-/					
						El	Ion Source Type	EI
m/z 69.07 Int 25,518,6		/z 131.07 16,632,001	m/z 219.00 Int 19,297,05/	m/z 41 Int 899		z 502.00 Po	larity	Positive
FWHM 0.6	4 FV	VHM 0.64	FWHM 0.62	FWHM	0.52 FW	HM 0.49 Ele	ctron Lens Voltage	15 V
						Ele	ctron Energy	70 eV
8		15			8	En	ission Current	50 µ A
Λ		٨	Λ			lon	Guide Frequency	1680.9
1		11	1 1.			Q1	Frequency	1092.7
11						Co	llision Cell Frequency	1833.7
		11				Q3	Frequency	1089.3
						Mu	tiplier Voltage	1614.6 V
		11		1 1		De	tector Gain	1.7 × 10 ^ 5
		11				MS	Transfer Line	250.0 °C
11						Fi	ament Selection	1
1.4		J.h.	Jh		N		Source Temperature	200.0 °C
er es	TI 125	100 1 102 1 121 122	218 218 220 217 219 2	411 412	413 499 3	di sta so4 Fo	reline Pressure	78 mTorr
						lon	Gauge Pressure	4.7 × 10 ^ -8 Torr
Mass	pectrum N	lot Acquire Abundar	ed noe Relative Ab	undance	isotope Mass T	heoretical Isotope Mass	Isotope Abundance	Isotope Ratio
69.00	69.00	16,300,8	809	100.00	70.05	70.00	192,688	1.18
130.97	130.99	10,821,5	507	66.39	132.00	131.99	351,091	3.24
218.96	218.99	12,352,8	842	75.78	220.01	219.99	508,655	4.12
413.97	413.98	505,3	12.23	3.10	415.01	414.98	42,802	8.47
413.37	501.97	381,2	248	2.34	503.04	502.97	34,894	9.15
501.98	Air/ Water Check		(%18/69)	29.47		Leak Check: 0.539	6 of reference Pass	
501.98	Check	H20	(
501.98	Oheck:		(%28/69)	3.97				
501.98	Check:	N2		3.97 2.00				
501.98	Check:	N2 02	(%28/69)					

Figure 34. Typical El Standard Tune Report

Typical results for an EI Standard Tune are listed below.

- Peak Intensities:
 - Base Peak is *m/z* 69 or 219
 - Base Peak ~ 20,000,000
- Water Background: *m/z* 18:69 < 240%
- Repeller Voltage:
 - Helium carrier gas = 3–8 V
 - Hydrogen carrier gas = 7–15 V
- Foreline Pressure: < 100 mTorr

• Ion Gauge Pressure: <5e-5 Torr

Note Foreline and ion gauge pressures are dependent on column flow rate.

• Leak Check: < 10%

Run the EI Standard Tune if you suspect a system problem. The following conditions could indicate an issue:

- Increased detector gain—Detector gain is related to multiplier voltage, so if the detector gain is increased, multiplier voltage will also increase.
- Leak check change—Leak check results change over time base on instrument conditions. Recently vented systems exposed to air should be lower than 10% after one day of pumping down. Assuming the system is leak free, the instrument leak check should constantly decrease over time until stabilizing.

El Standard Quick Tune

EI Standard Quick Tune—A shortened version of the EI Standard Tune recommended for use when subsequent maintenance tuning is needed after an EI Standard Tune. This tunes the repeller in conjunction with Q3 to overcome the effects of matrix buildup in the source. It starts with the last saved tune and sets the detector sensitivity to generate a m/z 219 ion with an intensity of 20,000,000 counts.

EI SRM Tune

EI SRM Tune—Provides EI tuning and completely retunes the system. This tune reoptimizes nearly all the parameters affecting the signal. This type of tune performs a mass calibration, tunes the lenses and resolution, and performs a leak check. Unless your SOP requires it, this is not the best tune to use on a daily basis because of the length of time required to run it. This tune is more sensitive for high mass than the EI Standard Tune, making it more sensitive to many SRM transitions. It sets the detector sensitivity to generate a m/z 219 ion with an intensity of 20,000,000 counts and verifies the correct mass calibration.

SRM tunes adjust the resolution of each mass at 50% peak height by design. This tune may provide slightly larger peak intensities than a Standard tune because it allows a slightly wider peak through Q1 and Q3. These tune types are intended for EI SRM analysis only and not EI Full Scan data acquisition.

Figure 35 shows a typical tune report for an EI SRM Tune on a system using helium as a carrier gas.

				I SQ Se	ries Tune R	esults		
							strument Name: TSQ \$	Sector Sector
25. CO. 200		21 16:58:39					strument ID: 1009920	00
Tune File: Ai	utoTune_El	_2016-10-21-	16-58-39.tsq8tun	B		L	Jser: TSQ8141213	
Tune Type:	EI SRM Tu	ne (built-in)						
Starting Tune	File: (La	st Saved Tun	e)					
	1			31 77.00			I Ion Source Type	EI
m/z 69.07 Int 23,865,3		/z 131.00 15,495,983	m/z 219.00 Int 18,506,60			m/z 502.00 F	olarity	Positive
FWHM 0.7	0 F\	VHM 0.70	FWHM 0.6	FWH	VI 0.62 F	WHM 0.60	lectron Lens Voltage	15 V
						E	lectron Energy	70 eV
							mission Current	50 u A
٨		٨	٨		1		on Guide Frequency	1680.9
11		1					1 Frequency	1092.7
		11					Collision Cell Frequency	1833.7
			1				13 Frequency	1089.3
			1 11			A	luitiplier Voltage	1583.7 V
							etector Gain	1.4 × 10 ^ 5
							IS Transfer Line	250.0 °C
						F	lament Selection	1
JL		Jh	Jh		2		on Source Temperature	200.0 °C
	1 12s 1	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	zle ' zle ' zbo 217 219			sdi 'sbz 50 502 504 F	oreline Pressure	79 mTorr
52 36609 -	1000	222 - 222		1525 15	98. 1988	k	on Gauge Pressure	4.7 × 10 * -6 Torr
ull Scan Sj Mass	pectrum f	Not Acquir Abunda	nce Relative A	bundance	Isotope Mass	Theoretical Isotope Mass		Isotope Rate
68.98	69.00	16,567,4	450	100.00	69.90	70.00	289,783	1.75
130.96	130.99	10,841 ,	989	65.44	131.95	131.99	394,088	3.6
218.96	218.99	13,180,8	335	79.56	220.04	219.99	501,896	3.81
413.96	413.98	1,184,0	024	7.15	415.06	414.98	99,455	8.4
502.21	501.97	801,0	020	4.83	503.11	502.97	127,918	1 5.97
Air/ Water Check:		H20	(%18/69)	25.90		Leak Check: 0.54	1% of reference Pass	
All/ Waler V		N2	(%28/69)	3.82				
All/ Water (
All/ Waler		02	(%32/69)	2.00				
All/ Water ((%32/69) (%44/69)	2.00 0.97				

Figure 35. Typical El SRM Tune Report

Typical results for an EI SRM Tune are listed below.

- Peak Intensities:
 - Base Peak is *m/z* 69 or 219
 - Base Peak ~ 20,000,000

Note Isotope Abundance and Isotope Ratios are only valid when using a tune type that tunes the resolution at 10% peak height.

- Water Background: *m*/*z* 18:69 < 240%
- Repeller Voltage:
 - Helium carrier gas = 3-8 V
 - Hydrogen carrier gas = 7–15 V

- Foreline Pressure: < 100 mTorr
- Ion Gauge Pressure: <5e-5 Torr

Note Foreline and ion gauges are optional devices. Their pressures depend on column flow.

• Leak Check: < 10%

EI SRM Quick Tune

EI SRM Quick Tune—A shortened version of the EI SRM Tune that is recommended for use when subsequent maintenance tuning is needed after an EI SRM Tune. This tunes the repeller in conjunction with Q3 to overcome the effects of buildup in the source. It starts with the last saved tune and sets the detector sensitivity to generate a m/z 219 ion with an intensity of 20,000,000 counts.

SRM tunes adjust the resolution of each mass at 50% peak height by design. This tune may provide slightly larger peak intensities than a Standard tune because it allows a slightly wider peak through Q1 and Q3. These tune types are intended for EI SRM analysis only and not EI Full Scan data acquisition.

EI Tune Check

EI Tune Check—Used to check how well your last tune is performing. As it is the fastest tune type, it allows you to quickly update the detector sensitivity as the system gets dirty. This allows the sample intensity to remain constant for longer periods without running a full retune of the instrument. The daily tune check performs a leak check, makes sure the mass calibration is correct, and sets the detector sensitivity to generate a m/z 219 ion with an intensity of 20,000,000 counts. If your SOP allows it, you can use this tune to rapidly verify that the previous lens tune is still generating good spectra.

El Diagnostics Only

EI Diagnostics Only—Runs a complete set of diagnostics, including a leak check, and generate a report. No tuning is performed. Uses the parameters from the last saved tune.

El Target Tune

EI Target Tune—Starts with the last saved tune and adjusts the ion ratios of the calibration gas to those expected of classic single quadrupole MS analysis. This tune is intended to be run after full calibration using an EI Standard Tune. If your SOP was developed in response to regulatory requirements for classic single-quadrupole MS analysis, or you require classic single quadrupole spectra for spectral library matching, use this tune. It also sets the detector sensitivity to generate a m/z 219 ion with an intensity of approximately 20,000,000 counts and verifies correct mass calibration.

Note If you are starting with a clean source, first run the EI Initial Tune as a reset tune. After this tune is complete, follow it with your preferred tune.

Fast Scan Tune

Fast Scan Tune—This tune retunes the system with fixed Q1 voltages necessary for fast scanning. By increasing the ion energies and shortening ion flight times through the mass analyzer, this tune provides increased ion signal required to tune resolution and perform mass calibration at a high scan rate. This tune may cause high mass ions to exhibit more fronting than the other built-in tune types. This tune performs a leak check and sets the detector gain to 300,000. It does not tune the detector gain. Run a fast scan tune when scanning above 10,000 amu/s to improve the mass calibration for high mass ions.

Negative CI Tune

CI- Tune—Used to analyze samples with negative CI. The standard NCI tune performs a mass calibration, then tunes the lenses and sets the resolution. This type of tune assumes you are using methane as the CI reagent gas and tunes the system with a 1.0 mL/min flow. This tune does not set the detector gain. The built-in CI- tune will start with the most recent AutoTune_NCI tune file, so have an appropriate tune file saved on the instrument's PC.

Note Chemical ionization tunes are very different from the electron ionization tunes. Do not use a CI tune unless your instrument has a CI ion volume and CI reagent gas installed.

Note If the instrument was last used in EI mode and tuned with a high repeller voltage before switching to a clean CI ion Source, a tune file with a low repeller voltage should be loaded in manual tune and saved to the instrument before tuning in CI mode.

Figure 36 shows a typical CI- Tune report where methane is the CI reagent gas.

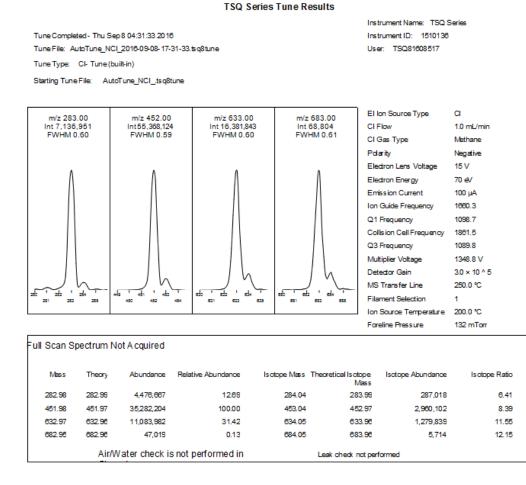


Figure 36. Typical CI- Tune Report with Methane as a CI Reagent Gas

Typical results for a CI- Tune using methane as the reagent gas are listed below.

- Peak Intensities:
 - Base Peak: 452 or 633
 - Base Peak ≥ 6,000,000
- CI Gas Flow: 1.0-4.0 mL/min Methane
- Emission Current: 50–100 μA

Note Emission current is an input value, and it should match the value set in the tune.

- Multiplier Voltage
 - Normal Performance: < 2200 V
 - Replace Multiplier: ≥ 2200 V
- Foreline Pressure: < 400 mTorr

• Ion Gauge Pressure: < 1e-4 Torr

Note Foreline pressure fluctuates with CI reagent gas flow rate. As the CI reagent gas flow rate increases, the foreline pressure also increases. Ion gauge pressure also increases if an ion gauge is installed on the system.

• Isotope Ratios: m/z 453:452 = 5.8-11.8%

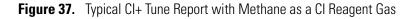
Positive CI Tune

CI+ Tune—Used to analyze samples with positive CI. The standard PCI tune performs a mass calibration, then tunes the lenses and sets the resolution. This type of tune assumes you are using methane as the CI reagent gas and tunes the system with a 1.5 mL/min flow. This tune does not set the detector gain. The built-in CI+ tune will start with the most recent AutoTune_PCI tune file, so have an appropriate tune file saved on the instrument's PC.

Note Chemical ionization tunes are very different from the electron ionization tunes. Do not use a CI tune unless your instrument has a CI ion volume and CI reagent gas installed.

Note If the instrument was last used in EI mode and tuned with a high repeller voltage before switching to a clean CI ion Source, a tune file with a low repeller voltage should be loaded in manual tune and saved to the instrument before tuning in CI mode.

Figure 37 shows a typical CI+ Tune report where methane is the CI reagent gas.



			TSQ Se	ries Tune R	esults		
	itoTune_PCI_;	4 01:52:18 2015 2015-07-14-15-8 It-in)				Instrument Name: TSQ Instrument ID: 1504062 User: TSQ8150543	
Starting Tune	File: AutoT	une_PCI_2015-	08-05-11-48-33.tsq8tune				
	m/z 219.0 Int 7,487,4 FWHM 0.0	58	In	n/z 414.00 t 13,095,158 WHM 0.60		El Ion Source Type CI Flow CI Gas Type Polarity	Cl 1.5 mL/min Methane Positive
z/e 217	2/18 2/19	da da	ali aliz aliz	d'a d's	4/2	Electron Lens Voltage Electron Energy Emission Current Ion Guide Frequency Q1 Frequency Collision Cell Frequency Q3 Frequency Multiplier Voltage Detector Gain MS Transfer Line Filament Selection Ion Source Temperature Foreline Pressure Ion Gauge Pressure	1091.8 1288.1 V 3.0 × 10 ^ 5 250.0 °C 1
ull Scan Sp	ectrum No	t A cquired				ion dauge riessure	4.0 × 10 **-5 101
Mass	Theory	Abundance	Relative Abundance	Isotope Mass	Theoretical Isoto	pe Isotope Abundance	Isotope Ratio
218.96 413.98	218.99 413.98	4,715,452 8,472,996	55.65 100.00	220.02 415.04	219 414	99 223,043 98 681,800	
	Air/W	ater check is	not performed in		Leak check no	ot performed	

Tip If you intend to use ammonia reagent gas, attach methane to one CI reagent gas port and ammonia to the other port. Tune the instrument using methane, then switch to the ammonia port. Allow plenty of time for the new reagent gas to purge the CI tubing before starting your analysis.

Typical results for a CI+ Tune using methane as the reagent gas are listed below.

- Peak Intensities:
 - Base Peak: 414
 - Base Peak ≥ 1,000,000
- CI Gas Flow: 1.5-4.0 mL/min Methane
- Emission Current: 25–50 µA

Note Emission current is an input value, and it should match the value set in the tune.

- Multiplier Voltage
 - Normal Performance: < 2200 V

- Replace Multiplier: ≥ 2200 V
- Foreline Pressure: < 400 mTorr
- Ion Gauge Pressure: < 1e-4 Torr

Note Foreline pressure fluctuates with CI reagent gas flow rate. As the CI reagent gas flow rate increases, the foreline pressure also increases. Ion gauge pressure also increases if an ion gauge is installed on the system.

- Isotope Ratios:
- m/z 415:414 = 5.8-11.8%

Note To add a tune type to the list, see Modifying an Automatic Tune.

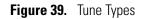
Tuning the Mass Spectrometer

- To tune your mass spectrometer
- 1. In the Tune Types field, select EI, CI, or Both, depending upon which ionization mode you are using.
- 2. Select a tuning category. See Figure 38.

Figure 38. Tuning Categories

🔣 TSQ Series Autotune	? 🔀
Tune types: 🔘 El 🔘 Cl 💿 Both	
Category:	Display report when complete
Now running:	Show spectra
Action:	Start

3. Select the tune type you want to use:



🙆 TSQ Series	Autotune	? <mark>×</mark>
Tune types:	C EI CI 🖲 Both	
Category:		
El Initial El SRM (El SRM) El SRM] El Standa	ostics Only (built-in) Tune (built-in) Quick Tune (built-in) Tune (built-in) ard Quick Tune (built-in) ard Tune (built-in) t Tune (built-in)	Display report when complete
Now running	j:	Show spectra
Action:		
		Start

4. Select the **Display Report When Complete** check box so that you can view the tune report after running the tune.

Figure 40. Displaying a Report

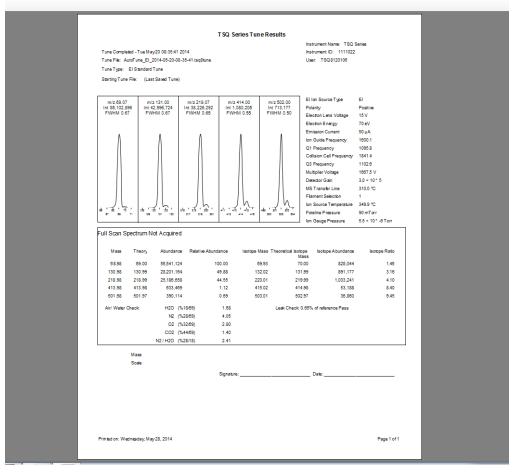
🔣 TSQ Series Autotune	? <mark>×</mark>
Tune types: ◎ EI ◎ CI ◎ Both Category: All	
El Diagnostics Only (built-in) El Initial Tune (built-in) El SRM Quick Tune (built-in) El SRM Tune (built-in) El Standard Quick Tune (built-in) El Standard Tune (built-in) El Tarnet Tune (built-in) V	Display report when complete
Now running:	✓ Show spectra
Action:	Start

- 5. Select the Show Spectra check box to show the spectra while the system is tuning.
- 6. Click the **Start** button to begin tuning.

Note Make sure the power options on your computer are not set to go into Standby mode while you are acquiring data for your tune. Otherwise, it will interrupt your tune.

Once the tune completes, your tune report opens in the *Tune Results Viewer*. If you did not select the **Display Report When Complete** check box, you can click **View Tune Report** on the dashboard and view the report.

Figure 41. Sample Tune Report



- 7. Click **Report Options**. In the dialog box, select the charts and reports you want to display and change the name of your instrument.
- 8. Select a mass for **Calculate Relative Abundance Using**. The default is mass 69, which will reference all masses to mass 69. Some masses may have over 100% abundance. If you do not wish to have a relative abundance above 100%, select Base Peak. This option will select the reference mass as the highest abundance mass in the tune.
- 9. Click OK.

	Report Options	Ĩ	164 109
	Lab name: OK	ncy	186 108
	Instrument name: Cancel		141 5.8
	Calc relative abundance using: 69		310 1
: ;;;;	Include signature line	ture	250 71
	Optional reports		3.5
mΝ	Diagnostics Device settings Tune type settings		
o	Custom report	ance	
ю	Use custom report definition	,901	
39	File name:	,654	
39		,346	
		,035	
)0 39	Diagnostics Device settings Tune type settings Custom report	,90 ,65 ,34	01 54 46 35

Figure 42. Tune Report Options

10. To print your tune report, click **Print Report** to open a print dialog box and print your report.

Note Only tune the system as often as your application requires. Repeated tuning on an otherwise functioning system can cause abnormalities as each tune reoptimizes all parameters affecting the signal.

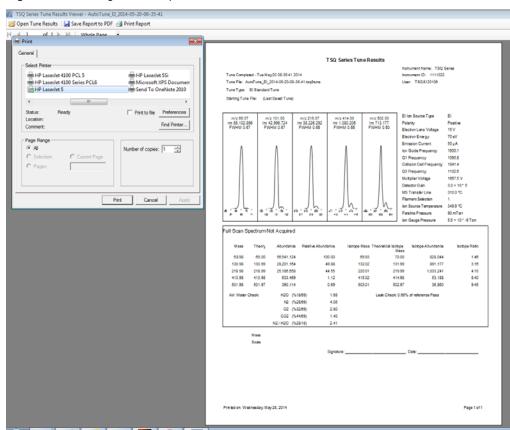


Figure 43. Printing a Tune Report

In the Tune Results window, you can open tune results, print reports, or change the way you view the report.

11. To save the report, click the **Save** icon **□** and save it as a Microsoft Excel[™] file or an Adobe[™] Acrobat[™] PDF file.

You may find it useful to compare this tune with a previous tune report of the same tune type. Some changes in peak height are normal, but if the difference is significant, see Troubleshooting.

- 12. To browse to another tune report on your computer, click **Open Tune Results** in the top of the window.
- 13. Click **Open** to open a previously saved tune report.

🔾 🛛 🕌 🔸 Computer 🔸 OS (C:) 🔸 Users 🔸	Public Public Documents				• 4 Search TuneResults
Irganize 🔻 New folder					II •
Favorites	Name	Date modified	Туре	Size	
Downloads	AutoTune_E1_2014-05-20-08-35-41.tsg8tuneres	5/20/2014 8:35 AM	TSQ8TUNERES File	85 KB	
E Recent Places	(Not Saved) AutoTune EL 2014-05-20-08-05-18.tsg8tun	5/20/2014 8:05 AM	TSQ8TUNERES File	23 KB	
E Desktop	(Not Saved) AutoTune El 2014-05-20-07-59-28.tsg8tun	5/20/2014 7:59 AM	TSOBTUNERES File	25 KB	
	Initial_EI_2014-05-20-07-53-54.tsq8tuneres	5/20/2014 7:53 AM	TSQ8TUNERES File	67 KB	
libraries	AutoTune_NCI_2014-05-13-15-58-59.tsq8tuneres	5/13/2014 3:59 PM	TSQ8TUNERES File	81 KB	
Documents	AutoTune_PCI_2014-05-13-15-30.tsq8tuneres	5/13/2014 1:15 PM	TSQ8TUNERES File	70 KB	
🚽 Music	Initial_EL_2014-05-13-11-44-36.tsq8tuneres	5/13/2014 11:44 AM	TSQ8TUNERES File	63 KB	
Pictures	(Not Saved)_Initial_EL_2014-05-13-11-19-27.tsq8tuneres	5/13/2014 11:19 AM	TSQ8TUNERES File	25 KB	
Videos	(Not Saved)_AutoTune_PCL2014-05-13-11-05-03.tsq8t	5/13/2014 11:05 AM	TSQ8TUNERES File	63 KB	
	[Not Saved]_AutoTune_PCI_2014-05-13-09-22-57.tsq8t	5/13/2014 9:22 AM	TSQ8TUNERES File	68 KB	
E Computer	(Not Saved)_AutoTune_PCI_2014-05-13-09-08-47.tsq8t	5/13/2014 9:08 AM	TSQ8TUNERES File	65 KB	
🏝 OS (C:)	(Not Saved)_AutoTune_PCI_2014-05-13-08-53-07.tsq8t	5/13/2014 8:53 AM	TSQ8TUNERES File	68 KB	
Removable Disk (Ht)	(Not Saved)_AutoTune_PCI_2014-05-12-16-53-24.tsq8t	5/12/2014 4:53 PM	TSQ8TUNERES File	67 KB	
	(Not Saved)_AutoTune_PCI_2014-05-12-16-36-17.tsq8t	5/12/2014 4:36 PM	TSQ8TUNERES File	73 KB	
Network	(Not Saved)_AutoTune_PCI_2014-05-12-15-33-10.tsq8t	5/12/2014 3:33 PM	TSQ8TUNERES File	73 KB	
	(Not Saved)_AutoTune_PCI_2014-05-12-15-18-38.tsq8t	5/12/2014 3:18 PM	TSQ8TUNERES File	73 KB	
	AutoTune_EL_2014-05-12-12-49-43.tsq8tuneres	5/12/2014 12:49 PM	TSQ8TUNERES File	68 KB	
	AutoTune_EL_2014-05-04-22-06-07.tsq8tuneres	5/4/2014 10:06 PM	TSQ8TUNERES File	105 KB	
	AutoTune_II_2014-04-02-18-51-26.tsq8tuneres	4/2/2014 6:51 PM	TSQ8TUNERES File	97 KB	
	(Not Saved)_AutoTune_El_2014-02-24-17-21-12.tsq8tun	2/24/2014 5:21 PM	TSQ8TUNERES File	102 KB	
	AutoTune_EL_2014-02-01-09-53-31.tsq8tuneres	2/1/2014 9:53 AM	TSQ8TUNERES File	68 KB	
	AutoTune_EL_2014-01-23-09-40-05.tsq8tuneres	1/23/2014 9:40 AM	TSQ8TUNERES File	99 KB	
	AutoTune_EL_2014-01-23-08-59-39.tsq8tuneres	1/23/2014 8:59 AM	TSQ8TUNERES File	88 KB	
	AutoTune_EL_2014-01-22-12-58-37.tsq8tuneres	1/22/2014 12:58 PM	TSQ8TUNERES File	88 KB	
	AutoTune_EL_2014-01-22-12-10-01.tsq8tuneres	1/22/2014 12:10 PM	TSQ8TUNERES File	88 KB	
	AutoTune_II_2014-01-22-11-42-44.tsq8tuneres	1/22/2014 11:42 AM	TSQ8TUNERES File	88 KB	
	Diagnostics_El_2014-01-22-10-59-17.tsq8tuneres	1/22/2014 10:59 AM	TSQ8TUNERES File	84 KB	
	AutoTune_EL_2014-01-20-15-26-23.tsq8tuneres	1/20/2014 3:26 PM	TSQ8TUNERES File	25 KB	
	AutoTune_EL_2014-01-20-15-18-14.tsq8tuneres	1/20/2014 3:18 PM	TSQ8TUNERES File	52 KB	
	AutoTune_El_2014-01-20-15-04-25.tsq8tuneres	1/20/2014 3:04 PM	TSQ8TUNERES File	103 KB	
	(Not Saved)_AutoTune_EL_2014-01-20-14-32-02.tsq8tun	1/20/2014 2:32 PM	TSQ8TUNERES File	98 KB	
File name:					Tune result files

Figure 44. Loading Tune Results

If the sensitivity and resolution are adequate for running your samples, you are ready to develop or run a method.

Updating Tunes for New RF Lens

If you have purchased a new lens 3/RF lens (PN 1R120574-0103), you must update the ion guide frequency in the Manual Tune utility and retune your instrument.

- * To update the ion guide frequency using the Manual Tune utility
- 1. Open the TSQ Series Dashboard.
- 2. Click Manual Tune on the dashboard. See Figure 45.

Custom Spectrum	
Auto Tune	
View Tune Report	
Tune Types	
Method Editor	Manual Turn
Manual Tune	— Manual Tune
AutoSRM	
Status Analyzer Power Maintenance	
TSQ 8000 Evo	
Status: Idle	
Actual Set-Point	
✓ MS transfer line 250 °C 250 °C temp.:	
✓ Ion source temp.: 199 °C 200 °C	
Vacuum: OK	
Foreline pressure: 100 mTorr	
lon gauge pressure: 5.0×10 ⁻⁶ Torr	

Figure 45. Opening Manual Tune on the TSQ Series Dashboard

- 3. The Manual Tune utility opens.
- 4. Select Frequency Tune on the top menu. See Figure 46.

```
Figure 46. Accessing Frequency Tune
```

Frequency Tune

lass & Res	olution Tune Freque	ncy Tune Target Tu	une Factors						
Spectra: Scan rate: Det. gain:		lonization mode: c lon polarity: Cal. gas level:	EI Positive Off	CI gas type: None CI gas flow: 0 <i>Changing CI gas typ</i>	mL/min e may take two minutes.	Start Scan			
٦						Q3 RF	2.1		Rep
8-							9 amu	5.559	
8-							1100 amu	741.909	- -
~ 1									
8-									
telative abundance 40 50 60									
abund.									

5. From the **Device** drop-down menu, select **Ion Guide**. See Figure 47.

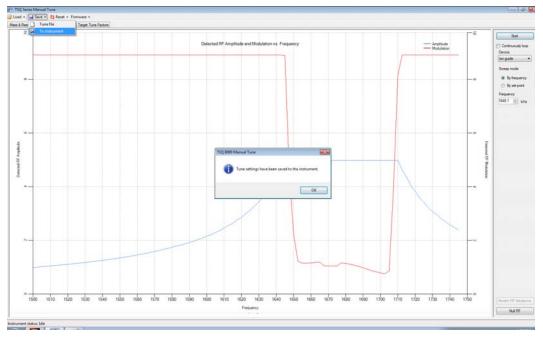
Device Menu

Figure 47. Locating the Device Menu

S TSQ Series Manual Tune Coad ▼ 🚽 Save ▼ 🚺 Reset ▼			Firm
Aass & Resolution Tune Frequency Tune Target T	une Factors		
	Detected RF Amplitude and Modulation vs. Frequency	Amplitude Modulation	Start Continuously loop Device Qued 3 Collision cell Qued 3 © By set point Frequency 1102.6 € kHz RF Imbalance Qued 0.00

- 6. Click **Start**. The system will detect the ion guide frequency.
- 7. When the instrument has finished detecting the ion guide frequency, go to **Save | To Instrument** on the top menu. A dialog box confirming that the tune settings have been saved to the instrument will open. See Figure 48.

Figure 48. Saving the New Ion Guide Frequency to the Instrument



8. To confirm that the intensities are still correct, select **Mass & Resolution Tune** on the top menu to go back to the Manual Tune home page.

9. Choose **3** from the **Spectra** drop-down menu. See Figure 49.

Figure 49. Selecting the Correct Number of Spectra to Scan

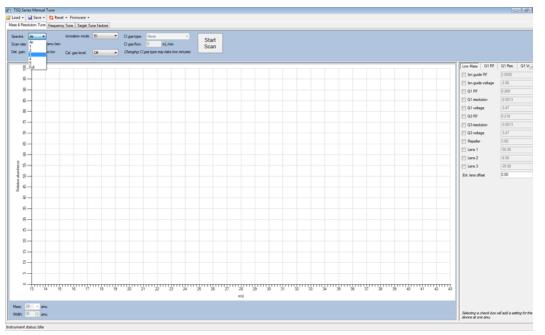
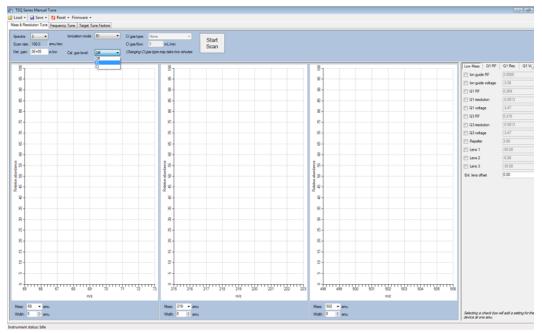




Figure 50. Setting the Calibration Gas Level to El



11. Click **Start Scan**. Check the intensities for the masses 69, 219, and 502. Set the masses using the **Mass** drop-down menu below each spectrum. See Figure 51.

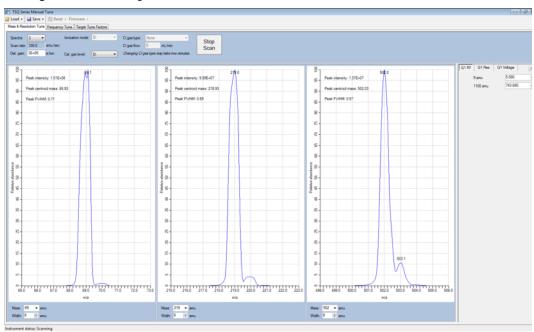


Figure 51. Checking the Mass Intensities

Note The intensities might be lower than they were at the previous frequency set for the ion guide until the lenses are tuned in AutoTune.

12. Once you have confirmed the intensities are sufficient for tuning, retune the system.

3 Tuning Updating Tunes for New RF Lens

Creating a Method

Once you have tuned the TSQ Duo system, you can create a method for each of its components. Use these methods to indicate to the GC/MS system how to collect your data.

Contents

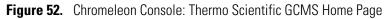
- Accessing the Method Editor
- Creating a GC-MS method
- Running a Sequence

Accessing the Method Editor

To create a method for the TSQ Duo mass spectrometer, the TRACE 1300 or 1310 GC, and your autosampler, go to **Start > Thermo Chromeleon > Chromeleon 7** and open the Chromeleon software application The **Chromeleon Console** opens to the **Thermo Scientific GCMS Home Page**. See Figure 52.

4

nstruments		O Launch eWorkfle	w • 🛞 Smart Sta	rtup - 🛞 :	Smart Sh	tutdown - 🅼 Take Control 🚫 Monitor Baseline - 🍈 Command 🛛 🖓 Detach 🗁 «Autogenerated» -	
9 💽 🚖	Filter 🌱	Thermo Scientific	GCMS Home	Sampler	Frontiniet	Oven MSDevice Etter Audt Queue	
TSQ-PC 8][594/64] ☆			Injection deta Data vaut: Sequence: Current injection: Type:	E		Parton time Dayrodici Vector No: Vector and	
		Correct	General settin Overal Status: Wating for:		and8y		
		1384962					
		1000000					
	1	800000				Spectral plot could not be created.	
		600000				No spectra selected	
		400000					
		-69253	5.0	10.	0	18.0 29.0 29.0 30.0	
		Date	Time R	tention C	Device	Message	
		8/5/2014	1:37:49 PM			User TSQ has disconnected Chromeleon Instrument Configuration Manager from this controller.	
		 8/5/2014 8/5/2014 	1:24:27 PM	_		MassSpecDriver - Mass Spectrometer Driver, Version 1 20 00. RC version - Thermo Fisher Scientific confidential - Not for sale! Copyright # 2005-2014 Thermo Fisher Scientific Inc. Jul rights reserved. Initiation driver Mass Spectrometer #2.	
instruments		0 85/2014	1:24:26 PM			hitializing driver Mass Spectrometer #2.	
🚰 Data		0 85/2014	1:24:26 PM			hisalizing driver CmDDKDrv. 1.	



Note All of the configured instruments are shown in the left pane of the Chromeleon Console window under **Instruments > TSQ-PC**. If your instruments are not shown, you must configure them. See "Configuring Your Instrument" for instructions.

Creating a GC-MS method

✤ To create a GC-MS method

1. Open the Chromeleon Console. Select **TSQ Duo** from the left menu. The TSQ Duo window opens. See Figure 53.

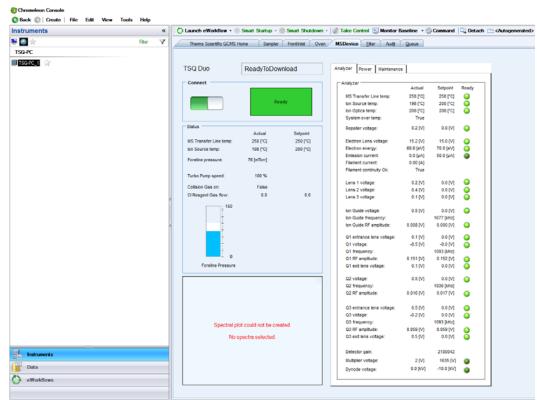


Figure 53. MS Device Window

Note Instructions for setting the most common parameters for the TriPlus RSH Sampling System follow. Refer to your autosampler documentation for more detailed information about settings.

2. Go to **Create > Instrument Method**.

trument E Folder	*	🗘 Launch eWorkflow -	Smart Startup -	Smart Shutdown
Sequence Sequence from Worklis Sequence from Worklis	Filter 🍸	Thermo Scientific GCMS	Home Sampler	FrontInlet Oven
Instrument Method ISQ-PC_1 Report Create a new Inst	ment Method	TSQ Duo	ReadyToDov	vnload
Injection Query Spectral Library		Connect		
View Settings			Re	ady
eWorkflow				
Electronic Report		Status		
			Actual	Setpoint
		MS Transfer Line temp:	250 [°C]	250 [°C]
		Ion Source temp:	201 [°C]	200 [°C]
		Foreline pressure:	76 [mTorr]	
		Turbo Pump speed:	100 %	
		Collision Gas on:	False	
		CI Reagent Gas flow:	0.0	0.0

Figure 54. Creating an Instrument Method

3. The Instrument Method Wizard – Select Instrument window opens. See Figure 55.

Figure 55. Instrument Method Wizard – Select Instrument

Instrument Method Wizard - Select Instrument			
e Instrument Method Witand guides you through the creation of instrument methods. To start, select the instrument where the method will sur.			1
Select an Instrument			
₩ 0 ☆		Fibr	$\mathbf{v} = \mathbf{v}$
TSQ-PC_T ■ TSQ-PC_T ☆			
a teach is			
	< Back Next >	Cancel	Help

- 4. Select the instrument you want to configure and click Next.
- 5. The Instrument Method Wizard System: General Settings window opens. See Figure 56.

	izard - System: General Settings			
eneral Settings for System				
Run Time				
Please specify the run tir	me of the method:			
20.000	(0.000100000.000 min)			
Diagnostic Channels				
Select diagnostic chann	nels to be used:			
Select diagnostic channel	nels to be used:	Select all channels		
	nels to be used:	Select all channels Deselect all channels		
	rels to be used:			
	rels to be used:			
	rels to be used:			
	rels to be used:			
	rels to be used:			
	rels to be used:			
	rels to be used:			
-	rels to be used:			
	rels to be used:		Next >	Cancel

Figure 56. Instrument Method Wizard – System: General Settings

- 6. You can set the GC Run Time for the method here, or load it with the other GC settings later. Click **Next**.
- 7. The Instrument Method Wizard TriPlusRSH: Settings window opens. See Figure 57.

nstrument Method Wizard	The resident of country of	
-	🚽 Save Template 🛛 🚰 Load Custom 🛛 📨 Edit Modules	1
ethod	ave remplate Coad Custom I ge calt Modules	
i.	lules, and click Create New Method button	
 GC Liquids 	GC Headspace SPME	
	Create New Method	
Injectors		
Injector port A - ID:	SSL Back	
Injector port B - ID:	None	
Injector port C - ID:	None	
Injector port D - ID:	None	
Syringes		
Syringe - ID:	• LS1 (10μL, 57mm) -	
Wash stations		
Wash station - ID:	 Standard Wash 1 (Standard wash station) 	
Internal standard / Solvent s		
Solvent station - ID:	• None	
Int standard station - ID:	• None	
Cooled/heated trays		
Primary tray - ID:	None	
Secondary tray - ID:	None	
		< Back Next > Cancel Help

Figure 57. Instrument Method Wizard – TriPlus RSH: Settings

- 8. Choose one of these three options: GC Liquids, GC Headspace, or SPME.
- 9. Enter the ID information for your samples. You must fill in any field with an asterisk (*)
 - a. Injectors—Enter the injection ID into the corresponding injector port.
 - b. Syringes—Enter the syringe ID information.
 - c. Wash Station—Enter the wash station ID information.
 - d. (Optional) Internal Standard/Solvent Stations—Enter ID information for your solvent station or interval standard station.
 - e. (Optional) Cooled/Heated Trays—Enter ID information for your primary and, if used, secondary trays.

10. Click Create New Method.

11. Use the following instructions to configure the GC Liquids general method settings. See Figure 58.

tings for TriPlusRSH.			
New 🛛 🚰 Load Template 🚽 Save Template 🛛 🚰 Load C	ustom 🛛 📝 Edit Modules		
eneral Washes Sync Advanced Injector pot Injector A (SSL Back) Injector mode Mode: Basic Rapid mode Rapid mode Syntge type Syntge volume (µL): 100 Plunger stokes: 3 Ar and filling mode: At a			
Sampler yial depth in vial Sample vial depth (mm): 30.0 Bottom sense Height from bottom (mm): 0.2	Sample viscoaity Sample type: Non visc	ous • Air	

Figure 58. General GC Liquids Settings on the TriPlus RSH Sampling System

- a. Injector Type—Choose Single or Double for each injector.
- b. Injection Mode—Choose Basic, Enrichment, Enrichment Needle Solvent Option, Internal Standard Double, Internal Standard Post, Needle Solvent Wash, Solvent Flush Double, or Solvent Flush Post.
- c. Rapid Mode—Choose Disable, After Sample Rinse, After Bubble Elimination, or After Sample Asp. in Home.
- d. Syringe Type—Enter your syringe volume in μ L. Enter the syringe needle length in mm.
- e. Sample Volume—Enter the sample volume to be injected into the GC. Typical values are between 0.5 and 5 μ L.
- f. Plunger Strokes—Select the number of plunger strokes to use when drawing up the sample. Air bubbles in the syringe change the amount of sample injected, which can cause signal variation in different runs. To prevent this from occurring, increase the number of plunger strokes to reduce the chance of an air pocket in your syringe. Typical values are between 3 and 10.
- g. Air and filling mode—Choose Auto to use the default or Custom to change the parameters.
- h. Sample Viscosity—Select Viscous if your sample is viscous or Non Viscous if your sample is non-viscous. With a viscous sample, the syringe is filled more slowly than if it was non-viscous. Since the amount of time saved is so small, setting this option to Yes might be easier.

- i. Sampling Depth in Vial—Select the Bottom Sense check box and enter the height from the bottom of the vial (in mm) at which the tip of the syringe needle will be placed in the sample vial when it is being filled.
- j. Injection Depth—Select Standard or Minimum to indicate how the sample is introduced into the GC. If you select Standard, the autosampler inserts the needle all the way into the injection port. If you select Minimum, the autosampler barely enters the injection port.
- k. Pre-inj Dwell Time(s)—Use this field to enter the time (in seconds) that the needle will be in the injection port before the plunger injects the sample.
- 1. Post-inj Dwell Time(s)—Use this field to enter the time (in seconds) that the needle will be in the injection port after the plunger injects the sample.
- 12. Configure the settings under the **Washes** tab (see Figure 59):

Figure 59. GC Liquids Washes Settings

New 😰 Load Template 🙀 Save Template 🎬 Load Custom 🕼 Edit Modules General Washes Sync. Advanced		
General Washes Sync Advanced	Basic	
Washes		
Number of solvents(s): Single Wash station: Standard wash station		
Pre-injection Solvent: A v - v - v		
Cycles: 1 -		
Solvent volume (µL): 1.0 -		
Rinse		
Rinses: 1 Rinse volume (µL): 1.0 -		
Post-injection Solvent: B V - V - V		
Cycles: 1 V		
Solvent volume (µL): 1.0 -		
	Sample	
	Air Air	
	I I	

- a. Number of Solvents—Choose Single or Multiple depending on the number of solvents you will use for your method. You may choose up to four solvents: A, B, C, or D.
- b. Wash Station—Choose Standard Wash Station or Large Wash Station. Refer to the autosampler documentation for more information about the wash stations.
- c. Pre-injection—Use the Solvent and Cycles fields to set the number of solvent purges that will occur before the autosampler touches your sample. Always include some sample rinses, either before or after injection, to make sure you do not have sample carryover from one injection to the next. Configure the settings so that the syringe is

purged with the same solvent that was used in location A, B, C, or D or with solvents A and B or C and D. Typically, there is 0 or 1 cycles of pre-injection purges.

- d. Rinse—Use the Rinses list to select the number of times the syringe is rinsed with your sample before each injection. Rinses help ensure the sample being injected is not diluted by the residual rinse solvents. By purging the syringe with your sample before injection, the dilution is minimal. The standard setting is between 1 and 3 rinses. If you have a very limited amount of sample, however, you may want to set this field to 0 to conserve the sample.
- e. Post-injection—Use the Solvent and Cycles fields to set the number of solvent purges that will occur after the autosampler touches your sample. You should always have some sample rinses, either before or after injection, to make sure you do not have sample carryover from one injection to the next. You can have the syringe purged with the same solvent that was used in location A, B, C, or D or with solvents A and B or C and D. Typically, there are 1 to 5 cycles of post-injection purges.
- 13. Click Next. The Instrument Method Wizard GC Inlets(TRACE1300Series): FrontInlet Flow/Pressure Options window opens.

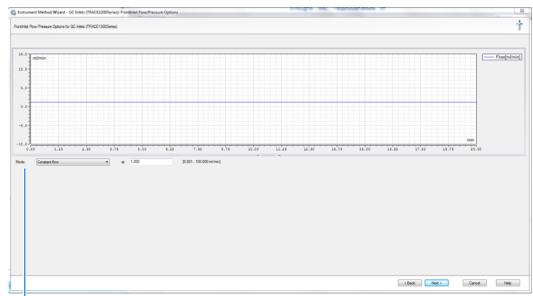


Figure 60. Instrument Method Wizard – Front Inlet Flow/Pressure Options

Carrier Gas Mode

14. Set the Mode to Constant Flow, Constant Pressure, Programmed Flow, or

Programmed Pressure. The gas flow and the oven temperature work together to determine how well the analytes are separated and how long the analysis will take. If you use constant pressure, as the column is heated in the oven, the flow rate will fall as the viscosity of the carrier gas increases. If you use constant flow, the carrier gas pressure will increase as the column temperature increases to keep the flow constant. Constant flow is more common. Typical flow rates are 1-3 mL/min. The pressure depends on the column

length and internal dimensions, so there is not a typical value. Because the outlet of the column is in the TSQ Duo instrument, which is under vacuum, be sure the vacuum compensation is on to ensure accurate flow rates.

15. Click Next. The Instrument Method Wizard – GC Inlets(TRACE1300Series): FrontInlet Options window opens. See Figure 61.

Figure 61. Instrument Method Wizard – GC Inlets(TRACE1300Series): Front Inlet Options

🛓 Instrument Metho	od Wizard - GC Inlets (TRACE1300Series): FrontIr	let Options	
FrontInlet Options for G	GC Inlets (TRACE1300Series).		
Temperature Settings C Enable temperature Temperature: 250 Inlet Parameters Operating mode Spliti	s control (2) (2) [0400 °C]	Utilities Vapour Volume Calculator Column Row Calculator	
Split flow control			
Split flow 40.0 Split ratio 33			
Splitless time 1.00			
Purge flow control Purge flow 5.00	•		
Constant septum p			
Stop purge for 200 Surge pressure 102 Surge duration 0.80 Vacuum compense Enable gas saver m Gas Saver Row 25.0 Gas savertime 150	200 Q [0.050010.0000 bar] 1 Q [0.0999.99 min] ation mode Q 1 Q [5.0500.0 mi/min]		

- 16. If you have a split/splitless inlet (SSL) injector, click the **S/SL-Front** or **S/SL-Back** tab to configure the injector port settings. In the inlet area, do the following:
 - a. Select the **Enable Temperature Control** check box and set the temperature high enough to volatize all the analytes in your sample. The material should be injected into the port to vaporize and move into the GC column quickly. Higher temperatures can lead to thermal decomposition of some analytes, so you must optimize the injector temperature for your analysis. The SSL temperature can be set up to 400 °C (752 °F). A typical value would be 250 °C (482 °F).
 - b. Select the **Split Flow Control** check box to dilute high concentrations of sample. The split flow is the amount of gas that is swept through the injector to the exhaust port. Higher values will give more dilution. The split flow reduces the amount of contamination that builds up in your system. The split flow ratio is the ratio of the split flow to the carrier gas flow. It is effectively the dilution ratio of the sample. This setting is typically turned on and set to a flow of 50 mL/min. However, more carrier gas is used, so for your analysis, lower split flows may be more acceptable. If you set the split ratio, the software calculates the correct split flow. The reverse is true also.
 - c. For best results, check the **Constant Septum Purge** box to use the septum purge, which means additional carrier gas will go through the injector. The default purge

flow value is 5 mL/min. This reduces the buildup of contaminants in the injector, on the column, and in the TSQ Duo instrument. If you perform a splitless injection, even if the split flow is set, the split flow turns off for the splitless time. The septum purge turns off for the stop purge time. After these times, the split flow and septum purge are reactivated.

- d. If your analysis requires a higher flow to quickly sweep the analytes into the column, which may be needed with high temperature injectors and thermally labile compounds, use the surge pressure to increase the column flow for the surge duration time.
- e. Because the outlet of the column is in the TSQ Duo instrument, which is under vacuum, be sure the vacuum compensation by checking the **Vacuum Compensation** checkbox is on to ensure accurate flow rates.
- f. In an effort to reduce the amount of carrier gas used, select the **Enable Gas Saver Mode**. When used, the split flow is reduced to the gas saver flow after the gas saver time. It is not recommended to use a flow of less than 20 mL/min because contaminants can build up in the injector, column, and TSQ Duo instrument, which can affect the system performance.
- 17. Click Next. The Instrument Method Wizard GC Inlets(TRACE1300Series): GC Oven Settings window opens. See Figure 62.

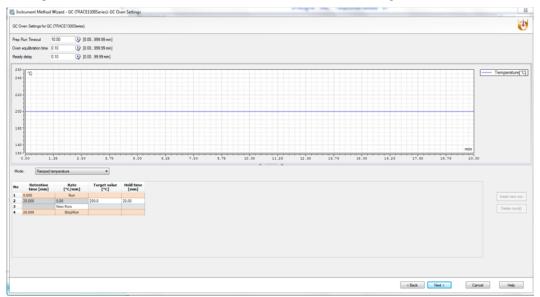


Figure 62. Instrument Method Wizard – GC Inlets: GC Oven Settings

18. Set the oven temperatures in this window. There is always at least one temperature and time in any GC temperature program. In the Initial row, enter the initial temperature, which must be 4 °C (7 °F) above room temperature and less than the maximum operating temperature of your GC column. If you set the initial temperature to a value below this limit, the GC will not reach the initial temperature. If you set the temperature above the limit, the GC column will be damaged. You can set the initial hold time to a value between 0 and 999.99 minutes. The typical initial temperature is at least 10 °C (18 °F)

above the boiling point of your sample solvent, and the initial time is usually long enough for the solvent to move through the column.

- 19. Select a maximum of 32 temperature ramps, each with its own ramp rate, final temperature, and hold time. Each temperature ramp begins at the previous ramp's temperature after the hold time has expired. A typical program has one or two ramps. The GC temperature profile is the primary method for separating your analytes from each other, the solvent, and the matrix. Your temperature profile must be optimized for your analysis needs.
- 20. Select the prep-run timeout. The prep-run timeout is the maximum amount of time that the GC will wait before it gives up on an injection. As an example, with the default value of 10 minutes, if the GC is ready to receive an injection, but does not receive it after ten minutes, the GC will stop waiting. This usually occurs in case of an error.
- 21. Set the oven equilibration time. The equilibration time is a delay between when the GC is at temperature and when the GC reports as being ready. The equilibration time is typically set to 0.5 minutes.



CAUTION INSTRUMENT DAMAGE. Be sure not to overheat the GC column or it may contaminate the instrument.

- 22. (Optional) Enable the cryogenic option to cool the oven. If this option is selected, then the minimum allowed temperature in a temperature ramp will fall from 0 to -99 °C (32 to -146 °F). The GC also allows the use of a post-run column cleaning. This is not typically used because the material that is purged from the column in this step would go into the TSQ Duo, which can lead to contamination. If you want to use this feature, set the GC oven temperature, as well as the amount of time to remain at that temperature after the analytical run is complete. You can also set the amount of pressure used to push the carrier gas through the column. Refer to the *TRACE 1300/1310 Series GC User Guide* for more information.
- 23. If you have a Programmable Temperature Vaporizer (PTV), click the **PTV-Front** or **PTV-Back** tab to configure it. The PTV is a low thermal mass injector that allows the instrument to rapidly heat or cool the inlet. You can use the PTV page to program the temperature of the injector. Refer to the GC documentation for details about the PTV or other types of injectors.

Note The user interface reflects the current configuration of your GC. If you add, remove, or change inlets or detectors, redo your instrument method according to the new GC configuration.

- 24. To add an inlet or detector to the Method editor user interface:
 - a. Attach the inlet or detector to the GC. Refer to the GC documentation for instructions.

- b. Add the inlet or detector to the current instrument configuration. See Chapter 1, "Introduction." for instructions to configure the GC.
- 25. Click Next.
- 26. From the Method Type list, select **Acquisition-General** or **Acquisition-Timed.** See Figure 63. An Acquisition method is used to collect data.

Figure 63. TSQ Series Method Type Setup

🚔 Instrument Method Wizard - MSDevice (TSQ Series): MS settings	×
MS settings for MSDevice (TSQ Series).	
TSQ Series	
Method Setup	
Method type: Acquistion - General Vise general acquistion methods to acquire any data type.	
Acoustion - Timed	Instrument model for method
MS transfer line tr Martenance onization mode: El	
Ion source temp.: 250 🕆 1C CI gas type: Methane v	Out from distances TOO Due Timed
Acquisition threshold: 1000 12 Cl gas flow: 100 12 mL/min Stop after: 1000 12 mL/min	n -
Scans	Groups
Time (min) SRM, SIM or Scan Masses Product Mass Neutral Loss Mass Collision Energy Dwell or Scan Times (sec) Tune File Name	Time Total Scan (min) Time (pec)
*	
<	
View Tune Report	
	< Back Next > Cancel Help
	Carles Nep

Note The **Maintenance** method is used to bake out or cool down the instrument during a sequence. This can be useful if you know you want to perform these tasks and you want to do them in an automated way as part of your sequence.

27. Use the **Method Setup** pane to set the temperatures and acquisition threshold in General Acquisition mode. See Figure 64 on page 66.

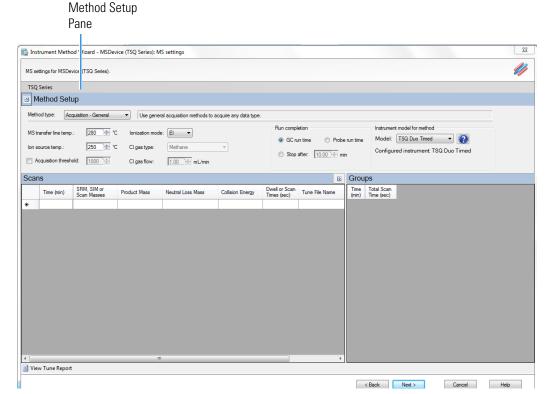


Figure 64. General Acquisition: Setting Temperatures and Acquisition Threshold

a. Set the **MS Transfer Line Temperature**. This field represents the temperature of the transfer line, which is the tube that contains the column as it leaves the GC oven and enters the TSQ Duo system. The maximum allowable temperature is 350 °C.



CAUTION You will damage the column and contaminate the TSQ Duo instrument if you set the transfer line temperature above the maximum allowed temperature for the column.

- b. Set the **Ion Source Temperature**. You can enter a value between 0 and 350 °C. The optimal temperature depends on the analyte. Higher temperatures will keep the ion source cleaner, but will lead to increased fragmentation, which may reduce sensitivity.
- c. (Optional) Select the **Acquisition Threshold** check box and enter a value for the minimum peak height for the data file, if needed. If your peak has an intensity that is below this threshold, it will not be stored. This setting may help reduce noise, but it may also alter the reported isotope ratios because the smaller isotope signals will be preferentially reduced.
- d. Select EI from the Ionization Mode pull-down menu.

Note Although CI settings are present in the TSQ Series software, chemical ionization is not available on the TSQ Duo, and the CI settings are inactive.

Note The following parameters apply only to the **Acquisition-General** method type only. **Acquisition-General** is the recommend setting for full-scan only MS methods, or simple SIM or SRM methods. If you need to develop a complex SIM or SRM method with a large compound list or overlapping retention times, refer to the instructions for **Acquisition-Timed**.

- 28. For Acquisition-General methods only, do the following:
 - a. In the **Run Completion** group, select the condition under which the run will end.
 - GC Run Time—Select this option if you want the MS run to end when the GC run is complete.
 - Probe Run Time—Select this option if you have a probe controller installed and you want the MS run to end with the probe run is complete.
 - Stop After—Select this option to set the number of minutes you want the MS to run. The end of the run can be between 0 and 1000 minutes. With this option you can stop the acquisition when all the compounds of interest have eluted, but the GC is still at an elevated temperature to keep the column clean. Thermo Fisher Scientific recommends that you select this option to save burn time on the filament.
 - b. In the Scans pane, click a scan row to enter scan information under each column. See Figure 65.

	Method Sei	tup	• Use pener	al acquisition m	albode to an	en inn men dala	hne						_
MS	is transfer line tem source temp.: Acquisition three	p.: 280 🚖 1	C Ionization mode			upure any data	- Run @	completion GC run time Stop after:	● Probe rur 10.00 🕀 min	time Mo	ument model for method del: TSQ Duo Timed	1	
ca	ans										1	Gro	ups
	Time (min)	Scan Name	SRM, SIM or Scan Masses	Product Mass	Neutral Loss Mass	Collision Energy	Dwell or Scan Times (sec)	SIM Widths (amu)	Tune File Name	Data Type	Precursor (Q1) Resolution	(min)	Total Scan Time (sec) 0.3370
	2.50	Full scan example	50-300				0.2		AutoTune_El	Centroid	Normal	• 5	0.3370
		SIM example	43, 44				0.05, 0.08	0.50, 0	AutoTune_El	Centroid	Normal	•	
	5.00	SRM example	292.9	257.91		20	0.05		AutoTune_El	Centroid	Wide (1.5)	-	
			292.91	192.05		32	0.05		AutoTune_El	Centroid	Nomal	-	
											Wide (1.5) Widest (2.5)		

Figure 65. Setting Acquisition-General Scan Information

Note If some of the following columns are not shown in the Scan pane, you can right-click and display them. You can also reorganize the columns by clicking the heading of a column and dragging it to the left or right.

i. Use the **Time (min)** column to set the time that the MS begins to acquire the scan group. Set the time for the first group late enough to allow the solvent to move through the system, and early enough to ensure the first analyte will be captured in the segment. Set subsequent segment times so that the start time will occur between compounds of interest. You may set up different types of scans in a single group. The scans will occur in descending order.

Your method may allow sequential full scans and SIM scans of a compound. As an example, in this method, the mass range of 50–300 will be scanned in 0.2 seconds. Immediately after the full scan finishes, the SIM scans begin. The SIM scans occur sequentially. Two different SIM masses, at 43 and 44, will be looked at for 0.05 and 0.08 seconds, sequentially. These simultaneous full scan and SIM scans will begin 2.5 minutes into the GC run. You will get a complete set of scans every 0.337 seconds. At 5 minutes into the GC run, the scans change to a set of SRM scans. The first SRM transition is 292.9 to 257.91 with a collision energy of 20 eV. The second SRM transition is 262.91 to 192.93 with a collision energy of 32 eV. Both SRM scans have a dwell time of 0.05 seconds. You will get a complete set of scans every 0.105 seconds.

Note The faster the MS scans, the more data points across a peak it will acquire. However, the precision of individual scans may be reduced.

All of these scans use the most recent AutoTune_EI file. You can also use a specific tune file for each of the scans.

- ii. Use the **SRM, SIM, or Scan Masses** column to tell the MS which masses to scan.
 - In full-scan mode, enter the start and end mass separated by a dash. You must put each full-scan range on a separate line.
 - For SRM methods, enter precursor masses, product masses, and collision energies of the ions you wish to monitor. Separate scan filters will be created for each SRM transition during acquisition.

Note Each line in a scan must only contain a full-scan range, individual SIM masses, or an SRM transition. Scan types cannot be mixed in a single line. Set up simultaneous scans by giving them the same time in the **Time (min)** column.

If you would like to combine multiple SRM transitions in a single scan filter, highlight the rows you want to combine and then right-click and select
 Group SRM Scans | Replace Selected Scans. See Figure 66. You can also use this function to ungroup a group of SRM scans into individual rows.

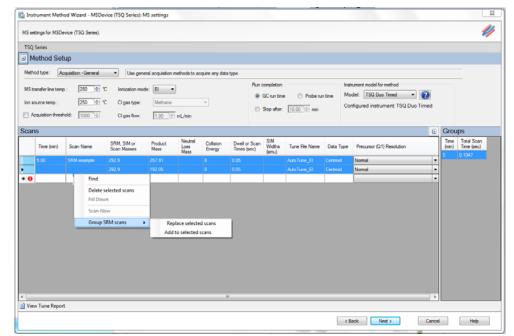


Figure 66. Grouping SRM Scans

iii. Enter the product ion mass (for SRM methods) in the **Product Mass** column. The product ion mass is the result of fragmenting the precursor ion with collision gas. If you want to perform a precursor ion scan, enter the mass of the target precursor ion into this column.

Note Entering a product ion mass changes a scan to SRM.

- iv. Use the **Neutral Loss Mass** column when you want to perform a neutral loss scan. Enter the mass of the target precursor ion in this column. The scan mass range must be at least 2.5 amu wide and must begin at 1 + the precursor ion mass.
- v. In the **Collision Energy** column, enter the collision energy (for SRM methods) in eV. The collision of a precursor ion with the collision gas at a particular energy fragments the precursor ions into product ions. By varying the collision energy, a single precursor ion can be fragmented into several product ions. An SRM transition is defined by a precursor ion being fragmented into a particular product ion mass using a certain collision energy. The Collision Energy column is unusable unless you define a precursor and product ion mass.

Note The system controls the collision gas so it will be ready when an MS/MS scan is requested.

vi. In SIM or SRM mode, use the **Dwell or Scan Times** column to define the amount of time (in seconds) that the MS will look at your SIM ion mass or your SRM product ion mass. If you are in full-scan mode, the **Dwell or Scan Times** column determines the amount of time to scan across the designated range. Set this value to have 5–20 scans across your GC peak. If you have too few scans, the

GC peak area is inaccurate. If you have too many scans, the MS signal becomes less precise. This column must be set to a value between 0.0005 and 5 s for SRM and SIM methods. The default is 0.2 s. For full scan methods, the minimum scan time depends on the mass range so that the scan rate does not exceed 13,000 amu/s.

vii. Use the **Tune File Name** to select a tune file to be used for this scan. Select the autotune file you created in Chapter 3, "Tuning," . If you wish to use the same tune file for multiple scans, you can fill down this column by right-clicking on a tune file name.

Note Selecting a tune file to open without a date-time stamp opens the most recent tune of the tune type selected. To use a specific tune file, select a tune file of that prefix with the date-time stamp that you want.

Tip Use a tune file without the date-time stamp to create your method.

viii. (Optional) Use the **Scan Name** column to enter a description of the scan. You may use the name as a label to indicate the compound used with the scan.

Note Scan Name, SIM Widths, Data Type, and Q1 Resolution columns are not shown by default. To view them, right-click one of the column headers to pull up the menu to display the hidden columns

- ix. Use the **SIM Widths** column to set the width range of the SIM window. The range of values can be between 0.01 and 10. The default is 1 amu, which means the MS will collect all the ions from your SIM mass ±0.5 amu.
- x. Use the Data Type column to select whether you want to collect Profile, Centroid, or Nominal mass spectra. Centroided mass spectra are the most common because they are used by most of the libraries and provide the smallest data files. Profile mass spectra provide detailed mass spectral peaks, which result in large data files. Profile mass spectra are not available for SRM scans. when you want to perform fast scanning (up to 20,000 amu/s), select Nominal from the drop-down list under Data Type.
- xi. Use the Precursor (Q1) Resolution column to open the peak width of the precursor mass for SRM scans. The options are Normal, Wide (1.5 at 50%), or Widest (2.5 at 50%). The default setting is Normal, which uses the tuned resolution values from the tune file specified in the Tune File Name column. Selecting Wide or Widest increases the precursor peak width to 1.5 and 2.5 amu (at 50%) respectively. Opening the precursor peak width will increase the ion signal into the collision cell and may improve the sensitivity of your method; however, it may also increase the background noise level.
- xii. Use the Product (Q3) Resolution column to open the peak width of the precursor mass for SRM scans. The options are Normal, Wide (1.5 at 50%), or Widest (2.5 at 50%). The default setting is Normal, which uses the tuned resolution values from the tune file specified in the Tune File Name column.

Selecting Wide or Widest increases the precursor peak width to 1.5 and 2.5 amu (at 50%) respectively.

c. In the **Groups** pane on the right, review the information in each row. See Figure 67. As you create scans in the Scans pane, information in the groups pane is automatically displayed.

Note By default not all columns are displayed in the Groups pane. Right-click a heading to display them. You can also reorganize the columns by clicking the heading of a column and dragging it to the left or right.

≥	Grou	ps				
	Time (min)	Tot: ✓	el Scan Chrom Chrom Filter Llee T Chrom. Filter	une File on Current	Emission Current (µA)	Use Last T Detector G
	2		Filament On: Yes		50	
		~	Emission Current			2010/10/10/10/10/10/10/10/10/10/10/10/10/
		~	Detector Gain			1212
			Electron energy: Use last tune value.			10101
			Calibration Gas: Off			101 IO
		_				NI-NI-NI-NI-NI-NI-NI-NI-NI-NI-NI-NI-NI-N
						1010
						24 PA 124
						001010
						10101
						101 IO
						21 PC
						1010

Figure 67. Accessing the Groups Pane

- i. The **Time (min)** column indicates the time that the MS begins to acquire data after the GC starts for that group. It is typical to have enough of a time delay for the start of the first group to allow the solvent to get through the column before starting acquisition. The Time column in the group pane cannot be modified. If necessary, modify the scan time in the Scans pane.
- ii. The **Total Scan Time (sec)** column indicates the sum of all the scans in each segment. See Figure 68. The total scan time also contains the stabilization time that occurs between each scan. This method, beginning 2.5 minutes into the GC run, gives you a complete set of scans every 0.337 seconds. At 5 minutes into the GC run, the scanning completely changes. Now the scans will repeat every 0.105 seconds until the GC run is complete. As the total scan time is a calculated value, it is not editable.

nstrument Method Wizard - MSDe	vice (TSQ Series):	MS settings										
settings for MSDevice (TSQ Series).												4
iQ Series												
Method Setup												
sthod type: Acquistion - General	• Use oer	veral acquistion m	ethods to acquire any	data type								-
					completion		Instrument	model for method				_
S transfer line temp .: 200 🐨		ode: El 💌				Probe run tim		TSQ Duo Timed	• 📀			
source temp.: 250 🔶	C Cligas type:	Methane	v .		Stop after: 10.0			d instrument TS	Q Duo Time	d		
Acquisition threshold: 1000	CI gas flow:	1.00	mL/min	0	stop after: 10.0	<u>10 (12)</u> min						
ans	■ Gro	une										
	Time	Total Scan	Chrom. Chrom. F	iter Filament	Use Tune File	Emission	Use Last Tuned	MS/MS Gain	Detector	Use Tune File	Bectron	Calbratio
Time (min) Scan Name	Scan Ma (min)	Time (sec) 0.1047	Fiber On Peak W	dth (sec) On	Emission Current	Current (µA) 1 50	Detector Gain	Multiplier	Gain 1.00E+005	Electron Energy	Energy (eV)	Gas Off
0												
fiew Tune Report												

Figure 68. Determining Groups Total Scan Time

- iii. Use the **Chrom Filter On** column to enable the chromatographic filter. This filter smooths spectral data as it is acquired, which may increase the signal-to-noise ratio by a factor of two or more. The chromatographic filter is most useful when at least four full scans are acquired across a GC peak. This setting is typically left on.
- iv. Use the **Chrom Filter Peak Width** column to set the peak width to match the width of the GC peak (in seconds). If the peak width is set too large, signal intensity may be reduced. The default value is 1 s.
- v. Use the **Filament On** column to turn the filament on and off in the selected segment. Turning off the filament increases the lifetime of the filament and keeps the ion source clean longer. However, no data is collected. Use this column if you had analytes eluting before the solvent peak. You can create a segment to turn off the filament during the solvent peak to preserve the filament.
- vi. Use the **Emission Current** column to set the emission current used during the acquisition. For optimal analytical performance and stability, use the emission current at which the system was tuned. However, if you want to use a different emission current, clear the **Use Tune File Emission Current** check box and enter a value in the **Emission Current** (μ A) column. A high emission current will lead to the production of more ions, but the interaction of too many ions in the source can cause a degradation in the resolution and signal. The margin of error is ± 0.5 μ A.
- vii. Use the **Use Last Tuned Detector Gain** column to indicate that you want to use the detector gain set in Auto Tune. If you do not need to use the gain set in Auto Tune or saved to the instrument in Manual Tune, then you can set the gain manually. Higher gains give larger signals, but may shorten the lifetime of the detector when concentrated samples are detected. The **MS/MS Gain Multiplier**

column only applies to groups with MS/MS events. The MS/MS gain multiplier only applies if the **Use Last Tuned Detector Gain** column is selected. The default MS/MS Gain Multiplier is 7 and is intended to give similar signal levels as a SIM scan on the precursor ion. If you enter a specific gain, the MS will use that gain for the SRM scan without the multiplier.

29. The following settings apply to Acquisition-Timed methods only.

Figure 69. Acquisition-Timed Method Settings

Instrument Method W	Vitard - MSDevice (TSQ Series	s): MS settings			
MS settings for MSDevice	(TSQ Series)				
TSQ Series					
Method Setup					
Method type: Acquir	ton - Timed 🔹 Use to	imed acquisition methods to acc	aure trived SRM or S	M data.	
MS transfer line temp :	200 10 10 Ionization	node D		Instrument model for method	
lon source temp.	250 10 YC Cigas too			Model TSQ Duo Timed 🔹 🕢	
Acquisition threshold				Configured instrument: TSQ Duo Timed	
	(Trans. St.)	· Day of erven			
Scans					
None	RT Ion Polari		Link Product Mase	to external file Collecon Energy	E Scan settings
•	Pit Ion rolan		PHODUCE MARK	Color Degy	Resulting total scen time 0.300 MeC
-		100			SRM/SIM Time 0.300 MC
					Lowest dwell time: 0.001 sec
					Window optimization
					2 Optimize
					Desired non dwell time: 10.019 ms
					Desired window 0.044 min
					Mranut window 03/4 min
					Feik with
					Mr. baseline peak width: 30/0- sec
					Desired scars per peak. 10 P
					SPM Residen
					Set resolution for each unque transition scan
					Procursor (21) Normal +
					Product (Q3) Normal ·
					Acquisition options
					Now for sequentity acquisitor and has
					Allow dwell time prioritization (High priority multiplier)
					Full scan Use full scan with mass range 50-550
					This can are call with mass random 20220
View Tune Report					
					KBack Next Cancel Help
					Tep Tep

a. In the **Scans** pane above list of scans, select **SIM** or **SRM** from the **Scan Type** list. If you want to link to an external method, click **Link to External File**. You may link to a .csv or .xml method file.

Note In order to edit the scans within the TSQ Duo method editor, clear the **Link to External File**.

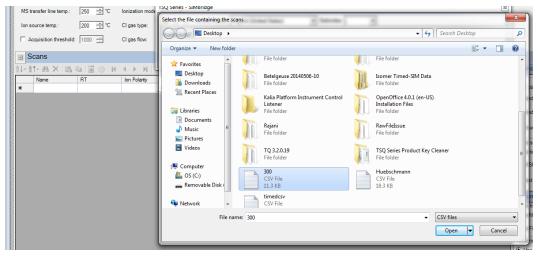
 After clicking Link to External File, the SIMBridge dialog box opens. Choose the language of your method file from the Source Locale drop-down menu. See Figure 70.

MS settings for MSDevice (TSQ Series)				4
TSQ Series				
Method Setup				
Nethod type Acquistion - Timed	Use timed acquisition methods to	to acquire timed SRM or SIM dat		
MS transfer ins temp: 200 (4) % Ion source temp: 250 (4) % Acquisition threehold: 1000 (4)	Ionization mode Ci gas type: Hethane Ci gas type: 100-10 eL/e	-	Instances code for network Model: [50 Das Treed] Configured instrument 150 Das Treed	
Scans				
Name PT	ten Pürety Mess	Product Name Coll English Clobert Strength English Clobert Strength English Clobert Strength English Clobert Strength English Clobert Strength English Clobert Strength English France English Clobert Strength English C		SIM Readon SIM Readon Peccary DD, <u>Immed</u> Peccary DD, <u>Immed</u> Aquation spine Aquation spine Aquation spine Aquation spine Ad scan I de for an eth mass region Stat free. Spine Ref acts refine mestings Tarvine Tarvine Tarvine Tarvine Tarvine Tarvine Under the generation currer. Deter pain Under the spinet spinet Deter pain Under spinet Deter pain Under spinet Under spinet Deter pain Under spinet Deter pain Deter pain D

Figure 70. Setting the Source Language of Method Files using SIMBridge

c. Browse to your file. See Figure 71.

Figure 71. Linking to an External File using SIM Bridge



- d. Click **Open** to open the method in SIMBridge.
- e. If necessary, change the method headings in your original file to match those in the method editor. A green check mark appears when your method is validated. See Figure 72.

ensfer line temp : 250 🕂 'C Ionization	TSQ Series - SIMBridge	e										
	S D B Source	locale: English (Unite	d States)	Delimiter:	•							
urce temp.: 200 🛨 °C Cligas type				J 1					-1			
quisition threshold: 1000 🛨 CI gas flow	Name of file: C:105er	rs\Alvin/Desktop\300.csv										
ans	Import map									-		
- MXILL BIR O N 4 >)	Column in file Map									Scan settings		
	Precentor Pages	212.0	2							Time summary		_
lame RT Ion Polarit	Product Produ	uct Mass 💌 182.0	2								0.300 9	
	CE Colle	ion Energy • 1	D							Resulting total scan time	0.300 9	
	Start Time Start	time 💌 3	5							SRM/SIM Time:	0.300 *	ec
	Stop Time End t									Lowest dwell time:		
	Pesticide Name									Lowest dwell time;	0.001 9	ec
										Window optimization		
										☑ Optimize		
										Desired min dwell time:	10.0÷ ms	
										Desired minoreal and	10.0 10	
										Desired window	0.6 🛨 min	
	Databa 200 same	ada farmad								Desired window Minimum window	0.6 🛨 min 0.3 🛨 min	
	Results - 300 compour			1		1						
	Name	RT Window	min) Ion Polanty Ma							Minimum window Peak width	0.3 <u>+</u> min	1
	Name Actorifien	RT Window 4 1	Postive 21	2.02 182.02	10	0	0	3.5		Minimum window		1
	Name Actonifen Actonifen	RT Window 4 1 4 1	Positive 21 Positive 26	2 02 182 02 1 03 194 02	10 15	0	0	3.5 3.5		Minimum window Peak width	03 ± min	1
	Name Acionifen Acionifen	RT Window 4 1 4 1	Positive 21 Positive 26 Positive 18	2.02 182.02 1.03 194.02 1.02 152.04	10 15 25	0	0	3.5 3.5 3.5		Minimum window Peak width Min. baseline peak width Desired scans per peak:	0.3 <u>+</u> min	1
	Name Actonifen Actonifen	RT Window 4 1 4 1 4 1	Postive 21: Postive 26 Postive 18 Postive 20	2 02 182 02 1 03 194 02	10 15	0	0 0 0 0	3.5 3.5		Minimum window Peak width Min. baseline peak width Desired scans per peak: SRM Resolution	03 ± min 30 ± 1	sec
	Name Actorifien Actoriation Activitation Activitation Alachier Alachier	RT Window 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1	Postive 21: Postive 26- Postive 18 Postive 20 Postive 16 Postive 18	2.02 182.02 4.03 194.02 1.02 152.04 8.05 181.04 1.07 146.06 8.08 160.07	10 15 25 8 12 10	0 0 0 0 0	0 0 0 0 0 0	3.5 3.5 3.5 3.5 3.5 3.5 3.5		Minimum window Peak width Min. baseline peak width Desired scans per peak: SRM Resolution SEt resolution for eacl	03 ± min 30 ± 1	sec
	Name Actorifen Actorifen Acrinathin Acrinathin Acriathin Active Active Active Active Active	RT Window 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1	Postive 210 Postive 26- Postive 18 Postive 200 Postive 16 Postive 180 Postive 290	2.02 182.02 4.03 194.02 1.02 152.04 3.05 181.04 1.07 146.06 3.08 160.07 2.91 222.92	10 15 25 8 12 10 20	0 0 0 0 0 0 0 0	0 0 0 0 0 0 0	3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5		Minimum window Peak width Min. baseline peak width Desired scans per peak: SRM Resolution	03 th min 30 th 10 10 th unique transitio	sec
	Name Actorifen Actorifen Actorifen Actoriten Actoriten Actoriten Alachior Alachior Alachior Alachion Aladhin Aladhin	RT Window 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1	Postive 21: Postive 26/ Postive 18 Postive 20 Postive 16 Postive 18 Postive 23 Postive 230	2.02 182.02 4.03 194.02 1.02 152.04 8.05 181.04 1.07 146.06 8.08 160.07 2.91 222.92 2.91 257.91	10 15 25 8 12 10 20 20	0 0 0 0 0 0 0 0	0 0 0 0 0 0 0 0 0 0	3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5		Minimum window Peak width Min. baseline peak width Desired scans per peak. SRM Resolution C Set resolution for eact Precursor (Q1): [Normal	0.3 1 min 30 1 t 10 1 t unique transitio	iec in scan
	Name Actorifen Actorifen Actorifen Actoristri Actoristri Actoristica Actoristica Actoristica Actoristri Actoristica	RT Window 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1	Postive 211 Postive 26- Postive 181 Postive 200 Postive 161 Postive 161 Postive 230 Postive 230 Postive 230 Postive 230	2.02 182.02 4.03 194.02 1.02 152.04 8.05 181.04 1.07 146.06 8.08 160.07 2.91 222.92 2.91 257.91 3.19 147.1	10 15 25 8 12 10 20 20 15			3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5		Minimum window Peak width Min. baseline peak width Desired scars per peak. SRM Resolution IT Set resolution for eacl Precursor (Q1): Normal Product (Q3): Normal	0.3 1 min 30 1 t 10 1 t unique transitio	sec
	Name Aclorifen Aclorifen Aclorifen Aconathin Aschlor Alachlor Addin Addin Addin Addin Addin Addin Addin Addin Addin Addin Addin	RT Window 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1	Postive 211 Postive 26 Postive 26 Postive 200 Postive 200 Postive 16 Postive 210 Postive 230 Postive 230 Postive 230 Postive 230	2.02 182.02 4.03 194.02 1.02 152.04 8.05 181.04 1.07 146.06 8.08 160.07 2.91 222.92 2.91 257.91	10 15 25 8 12 10 20 20	0 0 0 0 0 0 0 0	0 0 0 0 0 0 0 0 0 0	3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5		Minimum window Peak width Min. baseline peak width Desired scans per peak. SRM Resolution F Set resolution for each Precursor (21): Normal Product (23): Normal Product (23): Normal	0.3 min 30 min 10 min unique transitio	n scan
	Name Actorifen Actorifen Actorifen Actoristri Actoristri Actoristica Actoristica Actoristica Actoristri Actoristica	RT Window 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1	Postive 211 Postive 26- Postive 181 Postive 200 Postive 161 Postive 161 Postive 230 Postive 230 Postive 230 Postive 230	2.02 182.02 4.03 194.02 1.02 152.04 8.05 181.04 1.07 146.06 8.08 160.07 2.91 222.92 2.91 257.91 3.19 147.1	10 15 25 8 12 10 20 20 15		0 0 0 0 0 0 0 0 0	3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5		Minimum window Peak width Min. baseline peak width Desired scars per peak. SRM Resolution IT Set resolution for eacl Precursor (Q1): Normal Product (Q3): Normal	0.3 min 30 min 10 min unique transitio	n scan
	Name Aclorifen Aclorifen Aclorifen Aconathin Aschlor Alachlor Addin Addin Addin Addin Addin Addin Addin Addin Addin Addin Addin	RT Window 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1	Postive 211 Postive 26 Postive 26 Postive 200 Postive 200 Postive 16 Postive 210 Postive 230 Postive 230 Postive 230 Postive 230	2.02 182.02 4.03 194.02 1.02 152.04 8.05 181.04 1.07 146.06 8.08 160.07 2.91 222.92 2.91 257.91 3.19 147.1	10 15 25 8 12 10 20 20 15			3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5		Minimum window Peak width Min. baseline peak width Desired scam per peak: SRM Resolution for each Precursor (Q1): Normal Precursor (Q1): Normal Acquisition options Callon for asymmetric	0.3 d min 3.0 d min 10 d min unique transition unique transition unique transition	n scan vi scan vi vi vi vi vi vi vi vi vi vi
	Name Aclorifen Aclorifen Aclorifen Aconathin Aschlor Alachlor Addin Addin Addin Addin Addin Addin Addin Addin Addin Addin Addin	RT Window 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1	Postive 211 Postive 26 Postive 26 Postive 200 Postive 200 Postive 16 Postive 210 Postive 230 Postive 230 Postive 230 Postive 230	2.02 182.02 4.03 194.02 1.02 152.04 8.05 181.04 1.07 146.06 8.08 160.07 2.91 222.92 2.91 257.91 3.19 147.1	10 15 25 8 12 10 20 20 15			3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5		Minimum window Peak width Min. baseline peak width Denired scara per peak. SRM Resolution Focuror (21): [Normal Product (23): [Normal Product (23): [Normal Product (23): [Normal Product (24): [Normal Product (24): [Normal Product (25): [Normal Product (25): [Normal Product (26): [Norm	0.3 d min 3.0 d min 10 d min unique transition unique transition unique transition	n scan
	Name Aclorifen Aclorifen Aclorifen Aconathin Aschlor Alachlor Addin Addin Addin Addin Addin Addin Addin Addin Addin Addin Addin	RT Window 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1	Postive 211 Postive 26 Postive 26 Postive 200 Postive 200 Postive 16 Postive 210 Postive 230 Postive 230 Postive 230 Postive 230	2.02 182.02 4.03 194.02 1.02 152.04 8.05 181.04 1.07 146.06 8.08 160.07 2.91 222.92 2.91 257.91 3.19 147.1	10 15 25 8 12 10 20 20 15			3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5		Minimum window Peak widdh Min, baseline peak width Desired scars per peak. SPM Resolution Precursor (01): [Normal Product (02): [Nor	03 ± min 30 ± 1 10 ± unique transition unique transition sequisition wind	n scan v scan v priority
	Name Aclorifen Aclorifen Aclorifen Aconathin Aschlor Alachlor Addin Addin Addin Addin Addin Addin Addin Addin Addin Addin Addin	RT Window 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1 4 1	Postive 211 Postive 26 Postive 26 Postive 200 Postive 200 Postive 16 Postive 210 Postive 230 Postive 230 Postive 230 Postive 230	2.02 182.02 4.03 194.02 1.02 152.04 8.05 181.04 1.07 146.06 8.08 160.07 2.91 222.92 2.91 257.91 3.19 147.1	10 15 25 8 12 10 20 20 15			3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5 3.5		Minimum window Peak width Min. baseline peak width Denired scara per peak. SRM Resolution FSMR Resolution Focurror (21): [Normal Product (22): Allow down Allow dowell time priori	03 1 min 30 1 1 10 1 unique transitio unique transitio scaulation wind szation High t s range:	n scan

Figure 72. Changing Method Headings in SIMBridge

f. Click **Open** and the external method will be opened in the method editor. See Figure 73.



RS settings for MSDevic	e (TSQ Serex)				
TSQ Series					
Method Setup)				
Method type: Acou	ation - Timed • Use times	ecoustion methods to a	and the second states of	14 des	
Here and the literar	andel - Hereo - Cos Buel	acquestri merrode to a	cquestimes primiter :	instrument model for method	
MS transfer line temp :	200 12 To Ionization mod	e D		Model TSQ Dus Timed • 👔	
lon source temp.	250 10 C Cigashoe:	Nethane		Configured instrument TSQ Duo Timed	
Acquistion threshold		1.00 111 mL/mm		Configured instrument 13G Duo Timed	
	Linte us cidentes:	TUBL_12 erten			
Scans					
- 11 - 11 × 13	R (P N O R A F N	Scan type SRM	+ Link	to external file	E Desired scars per peak: 10.9
Name	RT Ion Polarity	Mass	Froduct Mass	Collsion (nergy	SRM Resolution
Rocken	4.00 Positive	Less than desired s	cans across peak pri	dicted	E Set resolution for each unique transition scen
Butrafol	4.00 Postive	• 123.64	75.03	15	Precursor (21) Normal
Captafol captan .	4.00 Postive	• 123.05	75.03	15	Product (Q2) Normal +
Heptenophos	4.00 Postive	124.01	89.01	10	Acquiation options
Denethoate EPTC	4.00 Positive 4.00 Positive	· 125 • 121.08	79 85.05	5	Also for approvate acquisitor variables
Feroropimorph	4.00 Postive	• 128.11	70.06	15	Allow dwell time prioritization
Ferpropriorph	4.00 Positive	• 128.11	110.05	15	folgen
Ferbuconacole	4.00 Postive	• 129.04	102.03	15	To be full scan with mass range 50-550
Keepon-nethyl	4.00 Postive	• 131.06	116.05	20	
Asinphos-ethyl	4.00 Postive	• 132.01	77.01	20	
Aprohoe-Methyl	4.00 Postive	• 132.02	77.02	20	Start time: 2009 nm. End time: 10004
Metazachior	4.00 Postive	· 133.05	117.04	20	
Heptachior epox		• 134.93	98.95	15	Instrument settings
Fondos	4.00 Postive	137.02	109.01	10	Ture file Ture file (a) [Internet]
Doofsi (1st, 2nd	- 4.00 Postive	• 138.97	110.97	20	
Fenatinol	4.00 Postive	138.01	111.01	15	Tune Re () Atstane_3E0 +
Daulfoton	4.00 Postive 4.00 Postive	• 142.01	109.01 57.99	10	Delector gan
Methodathion	4.00 Postive	• 144.90	57.39	10	Use last funed detector gain X 7.0-0
Fenalaguin	4.00 Postive	• 145.00	117.07	15	O Use specified detector part 2 1E-005
Ferpropidin	4.00 Postive	• 145.13	117.11	10	Emision current
Folpet met.	4.00 Postive	• 146.90	103.24	15	Use tune file emission current
	4.00 Postive	• 146.50	104.39	15	O Use specified emission current: 50.147 (LA)
Folpet met.		• 142.97	63.95	1	

- g. If you open an older model TSQ 8000 or a TSQ 8000 Evo (if you have a TSQ Duo system) method, the **Instrument Model for Method** box appears.
- h. Select your model from the drop-down menu in **Instrument Model for Method** in order to make the imported method compatible with your model.

i. In the **Name** column, enter the analyte name. If you have linked to an external file, the analyte name located in the first column of your linked file will appear in the first cell in the Name column. Right-click this window to search for an analyte within your method. See Figure 74. This function is useful if you need to edit an analyte in a complex method.

	MS transfer line temp.: 250				Model: TSQ 8000 E		0		
	lon source temp.: 300	☆ 'C Cligas type:	Methane	÷	Configured instrume	nt TSQ 8000 Ev	10		
	C Acquisition threshold: 100	Ci gas flow:	0.30 🛨 mLimin						
	Scans								
	21-21-21-24 × 🛍 🖏 👔	8 • • • • • • • • • • • • • • • • • • •	Scan type SRM	- Link Fir	nd Analyte				
Pight aligh the	Name RT	Ion Polarity	Window (min)	Pre-width (min)	Analyte to find:			in the	ion Energy
Right-click the ——	1,2,3-Trichlorobe	6.75 Postive 6.75 Postive	 ■ ■		1.2.3.4-Tetrachlorobenzene 1.2.3.5-Tetrachlorobenzene				_
Name Column	1,2.3-Trichlorobe	6.75 Postive	 1.3 1.3 		1.2.3-Trichlorobenzene				_
	1.2.4-Trichlorobe	7.38 Postive	 ■ ■		1.2.4-Trichlorobenzene 1.3.5-Tribromobenzene				
to Find an	1.2.4-Trichlorobe	7.38 Postive	1.1		1,3,5-Trichlorobenzene 1-Naphthylacetamide				
	1,2,4-Trichlorobe	7.38 Postive	· 1.3		2.3.5.6-Tetrachioroaniline 2.6-Dichlorobenzamide				
Analyte in a	Naphthalene	7.50 Positive	· 13		3. 5-Dichloraniline 3-Chloroaniline				
	Naphthalene	7.50 Positive	. 1.3	0	Acephate				
Method	Naphthalene	7.50 Postive	× 1.1	0	Acetamiprid Acetochior				
	3-Chloroanline	7.62 Positive	· 1.1	0	Acibenzolar-S-methyl Acionifen				
	3-Chioroaniine	7.62 Positive	1.3		Acrinathrin Akton				
	3 Chloroanline	7.62 Positive	1.3		Alachlor				
	Bhiolate	7.70 Positive	1.1		Allethrin				
	Ethiolate	7.70 Positive	1.1	-	Alidochlor Ametryne				
	Ethiolate	7.70 Positive	. 13		Aminocarb Amitraz				_
	Hexachlorbutadie	7.77 Positive	■ 1.3		Ancymidol				
	Hexachlorbutadie	7.77 Positive	1.3			1	Find Ca	ncel	_
	Hexachlorbutadie	7.77 Positive	- 1.3						
	1.3.5-Trichlorobe 1.3.5-Trichlorobe	7.78 Positive	 ▼ 1.1 ▼ 1.1 			0 179.9	74	38	
	1,3.5-Trichlorabe	7.78 Positive 7.78 Positive	 ■ ■			0 179.9	109	24	
	Methanidophos	7.93 Postive	- 13 - 13			0 141	64	18	
	Methamidophos	7.93 Positive	. 13			0 141	79	20	
	Methamidophos	7.93 Postive	I 13			0 141	94.8	8	
	Dichloryos	8.02 Postive	1.1			0 109	79	6	
	Dichlorvos	8.02 Postive	× 1.1			0 185	93	12	
	Dichlorvos	8.02 Positive	. 1.3	0	0	0 186.9	93	12	
	Country	0.05 0.05		0	0	0.00	20.1	10	

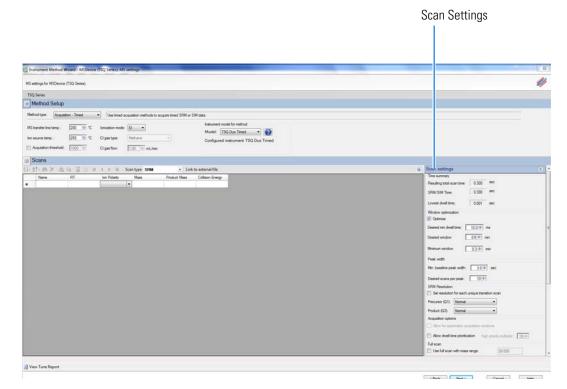
Figure 74. Finding an Analyte in a Method

- j. In the **RT** column, enter retention times for SRM methods. The retention time is the time it takes an analyte to pass from the column inlet to the detector.
- k. In the Window (min) column, set the acquisition times. Smaller acquisition windows increase sensitivity but can cause you to miss your peak if set too small. Changing the window size only affects sensitivity if it reduces the number of compounds analyzed in a segment. If the windows do not overlap, you will not notice an improvement by reducing the acquisition window.

Note The Window (min) column does not appear by default. The Method Editor displays it only when the window optimizer is not active.

- 1. In the **Mass** column, enter the precursor ion mass for SRM methods. For SIM methods, enter the mass of the ion you wish to monitor.
- m. In the **Product Mass** column, enter the product ion mass (for SRM methods). The product ion mass is the result of fragmenting the precursor ion with collision gas.
- 30. In the **Collision Energy** column, enter the collision energy (for SRM methods) in eV. The collision of a precursor ion with the collision gas at a particular energy fragments the precursor ions into product ions. By varying the collision energy, a single precursor ion can be fragmented into several product ions. An SRM transition is defined by a precursor ion being fragmented into a particular product ion mass using a certain collision energy.

- 31. Use the right-side panel in **Acquisition-Timed** mode to make the following adjustments to your method.
 - a. Use the **Scan Settings** panel to adjust the following settings. See Figure 75.
 - Figure 75. Acquisition-Timed Scan Settings



i. The **Window Optimization** pane allows access to the window optimizer settings. When the optimize button is checked, acquisition windows will be set automatically based on the acquisition window and dwell time targets set in this pane. For complex SRM or SIM methods, such as a method containing more than three hundred SRM transitions, this option will help insure a method is created that can achieve the requested scans per peak.

Algorithm Details: If the **Optimize** checkbox is checked, the SRM acquisition windows in the method are set to the **Desired Window** unless the **Desired Min Dwell Time** cannot be met with the number of **Requested Scans Per Peak**. If this occurs, then the acquisition windows are reduced until either the **Desired Min Dwell Time** is met, or the **Minimum Window** is reached. If the **Minimum Window** is reached first, the **Minimum Dwell Time** is reduced until the **Requested Scans Per Peak** is achievable. If the absolute minimum dwell time on the instrument, which is 0.5 ms, is reached before the requested points across the peak are achieved, the **Minimum Window** is lowered until the **Desired Scans Per Peak** criteria are met or until the absolute allowed minimum window on the instrument is reached, which is 0.24 min. In this very rare case you must reduce the number of **Desired Scans Per Peak**, increase the **Min Baseline Peak Width**, or reduce the number of transitions contained in your method before you will be allowed to save your method (see Chapter 7, "Troubleshooting.").

Settings: **Optimize** is checked by default. When Optimize is checked, you can adjust the settings in the pane to optimize your method. See Figure 75 on page 77. Remove the check from the box if you want to manually input your acquisition windows. The default values for the window optimizer should give reasonable results for normal methods. The default values are:

Desired Min Dwell Time—10 ms

Desired Window-0.6 min

Minimum Window-0.3 min

- Change the minimum dwell time using the Desired Min Dwell Time combo box. If your method has many transitions, you may want to reduce the desired minimum dwell time. Note that the wider the acquisition windows in your method, the shorter the average dwell time will be.
- Change the desired acquisition window in the **Desired Window** combo box. The desired window is the amount of time to scan for a transition around a given retention time to ensure that compound will be observed. The desired window can be set from 0.24–5 min. Set the window wide enough so that a retention time shift will not cause you to miss any compounds. Include extra time in this window if there is any uncertainty in compound retention times in the method. Note that the longer the dwell time for your compounds, the narrower your acquisition windows will be.
- Change the minimum acquisition window in the Minimum Window combo box. The minimum window can be set from 0.24–5 min. This is the smallest amount of time that should be scanned for a transition around a given retention time so that you are confident the compound will be observed. Set the minimum window to the lowest safe value to prevent compound retention times from drifting outside the acquisition window.

Note If the dwell time limit is reached and the minimum acceptable window is forced below the 0.24 min limit, the method will fail, and a smaller list must be used.

- ii. The **Time Summary** section reports the resulting total scan time, the SRM/SIM time, and lowest dwell time for you method. These values are for information only and not editable.
 - The Resulting Total Scan Time is the baseline peak width divided by the number of points desired across the peak. These values should be updated if your method requirements are different from the defaults.

- The SIM/SRM time is the total length of all SIM or SRM scans for each compound in your list. This will match the total scan time unless the method also has a full scan event.
- The Lowest Dwell Time is the actual lowest dwell time achieved by the method settings. When the Optimize check box is selected, if the actual lowest dwell time is considerably lower than the requested dwell time, then the minimum window has been reached, and if the actual lowest dwell time is considerably higher than the requested dwell time, then the requested window has been reached.
- iii. Under Peak Width, you can change the minimum baseline peak width and desired scans per peak. These values are used to calculate the SRM/SIM Time. The minimum baseline peak width should be set roughly to the shortest chromatographic peak time in your analysis.
- iv. Under SRM Resolution, check the Set Resolution for Each Unique Transition Scan checkbox if you want to adjust the Precursor (Q1) Resolution or Product (Q3) Resolution for individual analytes. If this box is not checked, the settings apply to all compounds in the method.
 - Use the Precursor (Q1) Resolution column to set the full width at 50% (FWHM) of the precursor mass for SRM scans. The combo-box options are Normal, Wide (1.5 FWHM), or Widest (2.5 FWHM) for each scan after a precursor mass has been entered. The default setting is Normal, which uses the tuned resolution values from the tune file specified in the Tune File Name column. Opening the precursor peak width increases the ion signal into the collision cell, which might improve the sensitivity of your method; however, it may also increase the background noise level.
 - Use the Product (Q3) Resolution column to set the full width at 50% (FWHM) of the product mass for SRM scans. The combo box options are Normal, Wide (1.5 FWHM), or Widest (2.5 FWHM) for each scan after a product mass has been entered. If the product mass is removed from a scan to run a SIM or FS scan event, the Product (Q3) Resolution column returns to the default value of Normal, which uses the tuned peak width at 10% or 50% as specified in the Tune File Name column. Selecting Wide (1.5 FWHM) or Widest (2.5 FWHM) increases the product ion peak width to 1.5 and 2.5 amu FWHM respectively, resulting in an increase of ions reaching the detector. The increase in ions at the detector may improve the sensitivity of your method; however, it may also increase the background noise level, as well as introduce isotopic interference from +/- 1 amu ions.

Note To achieve the highest transition speeds capable on the instrument, the resolution of all transitions must be set to Wide or Widest for both Q1 and Q3 resolution.

Note When the window optimizer is activated the **Window**, **Pre-width**, and **Post-width** columns are no longer available in the compound list editor. If your method is linked to a file or imported from a file with values in these columns, the values are stored in memory and can be recovered by deactivating the window optimizer.

v. Under Acquisition Options, select the Allow for Asymmetric Acquisitions check box to add extra time to the beginning or end of an acquisition window without affecting other timing in your scan. When you select this option, Pre-width and Post-width columns are added to your method. Enter the extra times in these columns.

Note This option is only available when the method optimizer is not active.

Select the **Allow Dwell Time Prioritization** check box to increase the dwell times for selected scans. The choices for each scan are **Normal** or **High**. Giving a scan high priority increases its dwell time by the value you set in the **High Priority Multiplier** box.

vi. Under Full Scan, you indicate if a Full Scan is to be run along with SIM and SRM. The Mass Range, Scan Time, Start and End Time can be set after the Use Full Scan box is checked. The full scan time will reduce the SRM/SIM time without increasing the total scan time. If you only want to use full scan for part of your method, you can enter full-scan start and end times.

Tip Click the arrow at the top right of the **Scan Settings** panel to collapse or expand the panel.

- b. Use the options in the **Instrument Settings** panel to further adjust your method.
 - i. Use Tune File to select a tune file or files to be used for this scan.
 - ii. In the Detector Gain area, set the detector gain. Select the Use Last Tuned Detector Gain option to indicate that you want to use the detector gain set in Auto Tune. If you do not need to use the gain set in Auto Tune, then you can set the gain manually. Higher gains give larger signals, but may shorten the lifetime of the detector when concentrated samples are detected. To manually set the detector gain, select the Use Specified Detector Gain radio button and enter the desired value in the Detector gain box.
 - iii. Use the **Emission Current** box to set the emission current used during the acquisition. For optimal analytical performance and stability, use the emission current at which the system was tuned. However, if you want to use a different emission current, select the Use Specified Emission Current radio button and enter a value in the Emission Current (μ A) box. A high emission current will lead to the production of more ions, but the interaction of too many ions in the source can cause a degradation in the resolution and signal. The margin of error is ± 0.5 μ A.

Tip Click the arrow at the top right of the **Instrument Settings** panel to collapse or expand the panel.

Subtrument Method Waard - MSDe 15 Instrument 11 MS settings for MSDevice (TSQ Series) Settings TSQ Serie A Method Setup Nethod type Acquisition - Timed • 200 10 TC Ioni de [] • Model TSQ Dus Timed

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Figure 76. Acquisition-Timed Instrument Settings

32. Click Next. The Instrument Method Wizard – Completion page opens. See Figure 77.

Figure 77. Instrument Method Wizard – Completion

ave entered all data required to create an instrument method.		
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Deception		
Network 150 PC 1 Genetro Tes 55 25 14 2 6 34 PM Owned by 150		
Press Finish to open the instrument method in the Orromatography Studio.		
	< Back Prish	Cancel H

33. Enter any comments or notes for your method and click Finish.

34. The New Instrument Method (Instrument Method) Chromeleon Chromatography Studio window opens. Click the **Save** icon to save your method. See Figure 78.

Figure 78.	Saving	Methods
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a Instrument Method			

- 35. The Save Instrument Method dialog box opens. Save your method in Chromeleon Local > Instrument Method. See Figure 79.
 - Figure 79. Finding the Instrument Method Folder

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Name	A	Туре	Date Modified	Comment	
Frankl Instrument Dat Instrument Mei Instrument Sec	hod	Folder Folder Folder Folder	5/8/2014 3:25 PM 7/17/2014 2:20 PM 8/3/2014 2:13 PM 8/3/2014 2:15 PM		
Object name: Channel:					

36. Enter your method in the **Object Name** box and click **Save**. Your method is now saved.

37. You can view all instrument methods on your system by opening the Chromeleon Console and selecting **Data** from the left menu. See Figure 80.

Figure 80. Viewing all Instrument Methods

sta		41				Instrument Method		
0 *	film	Y	Name -	Type	Date Modified		Comment	
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E Chenesoniaca			21p IDL study on 15m column	Instrument Method	6/18/2014 1:45 PM	25p IDL whydy		
8 D Instrument Data			25g IDL study on 30m column	Instrument Method	6/13/2014 11:18 AM	28g IDL study		
D Instrument Method			12 8270 method David	Instrument Method	6/10/2014 E 19 AM	David's 8270 DC split adjustment 20 method		
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			EI SRM method for OFN_amit	Instrument Method	6/51/2014 12:13 PM			
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			FS 1pg OFN SN tests for Duo	Instrument Method	8/3/2014 2 13 PM	FS SN test for 1pg OFN on column		
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			no solvent delay IDL testing	Instrument Method	6/13/2014 10:12 AM	no solvent delay 100fg run		
			Performance 7-22-14	Instrument Method	7(22/2014 12:19 PM	Performance		
			Performance A 7-22-14	Instrument Method	7:22/2014 12:40 PM	Performance		
			SRM Spece Splitless 7-22-14	Instrument Method	B/5/2014 1.18 PM	FullScen Scan Splitlese		
			test method	Instrument Method	\$/5/2014/2/12 PM			
			trial method for TriPlus classic	Instrument Method	7/22/2014 12:25 PM			
			TSQ 8000 suitability test method	Instrument Method	6/11/2014 11:17 AM	David's suitability test procedure		

38. To edit a method, select it from the list. The method opens in the Chromatography Studio. See Figure 81.

Figure 81. Editing a Method

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	K MS settings	
Overview	TSQ Series	
	Method Setup	
Sampler	Method type: Acquisition - Timed Use timed acquisition methods to acquire timed SFM or SIM data	
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rad seven	Acquisition threshold: 1000 🖶 Cligas flow: 1.00 🕂 el_Hain	
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	Name RT Ion Polarity Mass Product Mass Collision Evergy	Time summary
		Resulting total scan time: 0.300 MC
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		Product (03): Nomal ·
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39. Select a device from the left menu and edit its parameters as described in this chapter.

Running a Sequence

This section provides a quick introduction to running a sequence on a TSQ Duo MS. For detailed information, refer to the Chromeleon document set or help file.

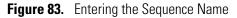
To create and run a sequence

 Go to Start > All Programs > Thermo Chromeleon. The Chromeleon Console opens. From the top menu, choose Create > Create New Sequence. See Figure 82.

film Ŷ TSQ Duo ReadyToDownload 250 [°C] 200 [°C] 208 [°C] 000 150 ["C] 190 ["C] 100 ["C] True 0.2 [V] 0.0 [V] 0 15.2 (M) 65.6 (HV) 0.0 (µA) 0.00 (A) True 15.0 [V] 70.0 [KV] 50.0 [µA] 000 0.1 [V] 0.4 [V] 0.1 [V] 0.0[V] 0.0[V] 0.0[V] 000 False 0.0 8.5 M 8.8 [V] 1677 [M42] 0.000 [V] 0 0 0.1 [V] -0.5 [V] [V] 0.0 [V] 0.8-8 0.152 [VHz] 0.152 [V] 0.0 [V] 0.151 [V] 0.1 [V] 8 0.5 (1) 0.0 [V] 1836 [kHz] 0.017 [V] 0 0.016 [M] 0 42M 0.0[V] 0 0.059 [V] 0 0.059 [V] 0 058 (V) 0.4 (V) 1635 [V] (0.01-2 [V] 0.0 [V]

Figure 82. Creating a New Sequence in the Chromeleon Console

 The New Sequence Wizard opens. In the Pattern for Injection Name box, enter your sequence name. See Figure 83.



nknown Injection Generate injection:	is s of type "Unknown"					Ĩ
		Rack View «	Pattern for Inje sample Number of Viai Injections per Start Position: Injection Volum	ls: 1 Vial: 1		[1105] [1999]
quence Preview						:
Chromatogram	Name	Туре	Level	Position	Volume [µL]	Instrument Method
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3. Click **Next**. The **Methods & Reporting** window opens. Browse to the instrument method you want to associate to your sequence. See Figure 84.

Figure 84. Associating an Instrument Method to a Sequence

TNew Sequence Wizard	and the second	4	8 23
Methods & Reporting			
Specify methods and rep	orting preferences		
Method Selection			
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Processing Method:		Brow	se
Defaults			
l			
Report Template:		Brows	se 🔻
View Settings:		Brows	se 💌
Channel:		*	
			1
		<< Back Next >>	Cancel

- 4. Click **Next**. The General Sequence Setting window opens. Add comments about your new sequence if desired and click **Finish**.
- 5. The Save Sequence window opens. See Figure 85.

				_
Add a detailed comment for your nev	Look in: 📴 ChromeleonLocal		- 4) • 🔇 • 🖄 🛗 🔢
	Name	Туре	Date Modified	Comment
	🛅 frankl	Folder	5/8/2014 3:25 PM	
Comment:	lnstrument Data	Folder	7/17/2014 2:20 PM	
sample	Instrument Method Instrument Sequence	Folder Folder	8/5/2014 2:12 PM 8/3/2014 2:15 PM	
	Object name:			
	Object name: Channel:			

Figure 85. Saving a Sequence

6. Navigate to ChromelonLocal > Instrument Sequence. Enter your sequence name in the Object Name box and click Save. See Figure 86.

New Sequence Wizard	Save Sequence			8 8 8
General Sequence Settings				
Add a detailed comment for your nev	Look in: 🛅 Instrument Sequence		- 🥥	- 🔾 - 🖄 📋
	Name	Туре	Date Modified	Comment
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Comment:	🗂 2fg IDL study	Sequence	6/18/2014 1:46 PM	2fg IDL study
sample	🗂 15m IDL study	Sequence	6/24/2014 5:15 PM	2fg IDL study
sampie	🗂 100fg OFN full scan IDL test	Sequence	6/13/2014 11:18 AM	100fg no sol
	🗂 100fg OFN full scan no solvent delay	Sequence	6/11/2014 6:25 PM	100fg OFN
	🗂 column performance test mix using	Sequence	7/22/2014 5:37 PM	8270 metho
	🗂 effect of Ar gas pressure	Sequence	6/27/2014 8:48 AM	sequence fo
	🗂 effect of Ar gas pressure - Copy	Sequence	8/3/2014 2:03 PM	sequence fo
	FS 1pg OFN tests for SN on Duo	Sequence	8/5/2014 1:17 PM	
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	Full Scan split 7-22-14 Sequence	Sequence	7/22/2014 5:31 PM	
	📅 Full Scan Splitless 7-22-14 Sequence	Sequence	7/23/2014 2:43 PM	
	🗊 Full Scan Splitless 7-23-14 Sequence	Sequence	7/23/2014 5:47 PM	
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	Reformance mix test	Sequence	6/12/2014 9·35 AM	Performanc
	Object name: test seq			
	Channel:			
	Object type: Sequence			
				Save Ca

7. Open the Chromeleon Console and click **Data** form the left menu and choose your sequence from the **Chromeleon Local > Instrument Sequence** folder in the left menu. See Figure 87.

Data	41						test seg					
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Figure 87. Opening a Sequence in the Chromeleon Console

8. Enter your sample names and sequence parameters in the table. See Figure 88.

Figure 88. Entering Sequence Parameters

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9. Click Start.

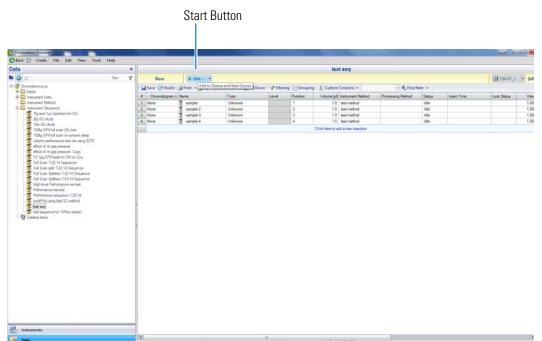
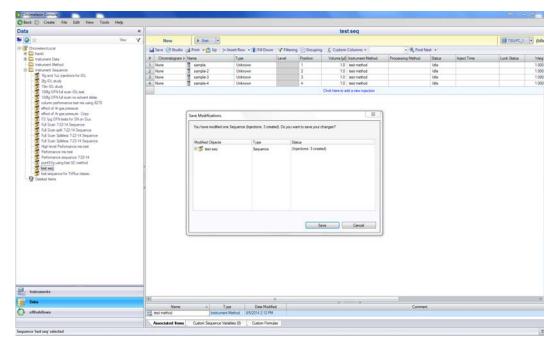


Figure 89. Starting a Sequence

10. The **Save Modifications** dialog box opens. If you are satisfied with your sequence, click **Save**.





11. Your run begins. You can click Stop if you want to stop the run. See Figure 91.

Figure 91. Stopping a Run

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12. When the run is complete, refer to the Chromeleon user documentation or help files for instructions on analyzing your data.

Optimizing Your Method

If you are not getting the expected results from your method, you can modify it using the following procedures for better results.

Contents

- Changing the Chromatographic Separation
- Finding the Best Way to Make an Injection
- Improving the Way You Prepare Samples
- Changing the Dwell Time or Scan Rate
- Narrowing the Mass Range
- Adjusting the Transfer Line and Ion Source Temperature
- Modifying an Automatic Tune

Changing the Chromatographic Separation

Peak shapes are defined by the chromatographic conditions. If your peak is too wide, too narrow, or not symmetrical enough, then changing the chromatographic conditions are the best way to improve your method. Start by changing the GC carrier gas flow or oven temperatures (the initial temperature, initial hold time, ramp temperature, final temperature for that ramp, and hold time at that final temperature). These temperatures can be adjusted for each ramp.

Oven changes are strongly dependent on the nature of the compounds you are analyzing. At some point, the GC oven has to be above the boiling point of the compounds you are looking for. If the GC oven is not at the boiling point, the compounds will not volatilize and they will become immobilized. You can change ramp rates to separate coeluting peaks. Refer to the GC user documentation for further suggestions on optimizing chromatographic separation.

Finding the Best Way to Make an Injection

Adjusting the way you get the sample from the needle into the column can sometimes improve the results of your data. Try modifying the autosampler method, injecting a different amount of liquid, adjusting the injector port temperature or flow, or changing the speed of your injection. You can also try using a hot or cold needle injection. In some cases, adjusting your injection port liners may give you better results. (For detailed instructions, refer to the user guide for your autosampler.)

Improving the Way You Prepare Samples

Although sample preparation adds time and expense to the overall analysis, a more focused method can give you better results. Try extracting your sample in a solvent that increases the solubility of the analytes of interest, but does not increase the solubility of the other compounds. If your method allows it, try switching solvents.

You can also use or change the phase of a solid phase extraction cartridge, which gives you similar results as changing a solvent. You can affect the way you prepare samples by changing the type of cartridge you are using.

Changing the Dwell Time or Scan Rate

The precision of your data depends on how well you define your chromatographic peak. Typically, you get good precision when sampling ten times across the chromatographic peak. In a full scan analysis, increasing the scan rate increases the number of times you have sampled across the peak. However, increasing the scan rate too much results in mass spectral noise, which decreases your analytical precision.

In a SIM or SRM analysis, reducing the dwell time will have a similar effect to increasing the scan rate in full scan. The dwell time is a measure of how long the instrument will average a SIM or SRM event. To optimize your scan rate or dwell time, select a value that gives you 8-12 points across a chromatographic peak. In timed acquisition mode this optimization is done automatically, providing you with the longest dwell time or slowest scan rates possible such that all compounds receive the desired scans across a chromatographic peak.

* To prioritize the dwell times of the problem analytes

- 1. In the Acquisition Options area, check Allow Dwell Time Prioritization. See Figure 92.
- 2. Set the High Priority Multiplier to the desired value.
- 3. A new Dwell Time Priority option appears in your scan list. The choice are Normal or High. Choosing High multiplies the dwell times by the value set in the previous step.

Note The total scan time does not change. If prioritizing some dwell times reduces others to less the lowest allowed dwell time for your method, try increasing the acquisition windows.

Figure 92. Setting the Dwell Time Prioritization

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	Diflubenzuron		Postive	-	1.30	0	0 141	113	12		-	Lowest dwell time; 0.001 sec
	1.2.3.4-Tetrachio		Postive	-	1.30	0	0 215.8	108	38	Nomal	-	
	1,2,3,4-Tetrachio		Postive	-	1.30	0	0 215.8	144.8	24	Nomal	-	Window optimization
	1,2,3,4-Tetrachio		Postive	-	1.30	0	0 215.8	180.9	16	Nomal	-	P Optimize
	Dichlobeni		Postive	-	1.30	0	0 170.9	99.9	24	Nomal	-	Desired min dwell time: 10.0 + ms
	Dichlobeni		Postive	-	1.30	0	0 170.9	136	14	Nomal	-	Desired window 0.6 - min
	Dichlobeni		Postive	-	1.30	0	0 172.8	99.8 54	24	Nomal	-	
	Maleic hydrazide		Postive	-		0				Nomal	2	Minimum window 0.3 1 min
	Maleic hydrazide		Positive	-	1.30	0	0 112	54	18	Nomal	-	Peak width
	Maleic hydrazide		Positive	-	1.30	0	0 112	82	8	Nomal	-	
	Dichlomid		Positive	-	1.30	0	0 108.2	93	16	Nomal	-	Min. baseline peak width: 3.0 + sec
	Dichlomid		Positive	-	1.30	0	0 172	108.1	6	Nomal	-	Desired scans per peak: 10-th
	Dichlomid		Positive	-	1.30	0		136.1	6	Nomal	-	SRM Resolution
	EPTC EPTC		Positive Positive	-	1.30	0	0 128.1	43.1	10	Nomal	-	Set resolution for each unique transition scan
	EPTC		Postive		1.30	0	0 189.1	43.1	6	Nomal	-	Precursor (Q1): Normal
	1,2,3,5-Tetrachlo		Postive	-	1.30	0	0 215.8	128.1	38	Nomal	2	
	1,2,3,5-Tetrachio		Postive	-	1.30	0	0 215.8	108	38	Nomal	2	Product (Q3): Normal
	1,2,3,5-Tetrachio		Postive	-	1.30	0	0 215.8	144.8	24	Nomal	4	Acquisition options
	1,2,3,5-Tetrachio Bohenvi		Postive		1.30	0	0 151.8	125.8	24	Nomal	4	Allow for asymmetric acquisition windows
	Bohenyl		Postive	-	1.30	0	0 151.8	125.8	24	Nomal	4	Allow dwell time prioritization High priority multiplier: 10-1
	Boheryl		Postive	-	1.30	0	0 154.1	127.4	30	Nomal	÷	Full scan
	Popamocath		Positive	-	1.30	0	0 58.1	42	20	Noma	÷.	Use full scan with mass range: 50-550
	Propamocarb		Positive	-	1.30	0	0 129.1	42 58.1	12	Noma		
	Propanocarb		Positive	-	1.30	0	0 188.2	58.1	8	Normal	i .	
		0.00			1.30		0 100.2	77	22		-	Start time: 2.00 - min. End time: 10.00 -

Narrowing the Mass Range

In a full scan analysis, by narrowing your mass range, you can look directly at the compounds of interest. However, if you are looking at a large number of compounds that have a broad range of mass fragments, a wide mass range makes sense. To narrow the mass range, refine your scan parameters to a smaller number. A narrower mass range also allows you to decrease the scan rate and get the same chromatographic peak sampling. By breaking your MS method into groups, you can create compound-specific MS settings to optimize your data.

Adjusting the Transfer Line and Ion Source Temperature

If your transfer line temperature is set too low, the less volatile compounds may get stuck in the transfer line and never make it into the ion source. On the other hand, if your transfer line is too hot, you could damage the column or cause a thermal breakdown in the compounds you are analyzing. Typically, aim for the transfer line temperature to be 10 °C (18 °F) over the highest boiling point of the compounds of interest, but no higher than the maximum safe operating temperature of the column.

Use the highest ion source temperature setting that allows adequate response for targeted analytes in order to improve response for high-boiling compounds and prevent column bleed and matrix compounds from dirtying the ion source.

Note The transfer line temperature and ion source temperature should be similar to avoid contaminating the ion source.

Optimizing an SRM Method

The following actions might improve your results when using an SRM method:

- Use timed SRM to optimize segments automatically and possibly improve dwell time and number of points across the peak.
- Reduce the Retention Time window if possible. If your sample's retention time is well known the scan window can be reduced. If this is in a high-density compound area it can dramatically improve the ion signal and number of points across the peak.
- Use AutoSRM to optimize collision energy and retention times. This can reduce errors in collision energy and improve confidence in retention times to allow shorter windows.
- Choose the optimum fragment ion for study if allowed by your method. Some fragments are higher in intensity and have less interference associated with their transition.

Modifying an Automatic Tune

The MS instrument control software includes a utility that uses certain parameters in the tune types to optimize system performance when generating a tune file. Follow these procedures:

- To modify an automatic tune
- To modify advanced tune settings
- To modify an automatic tune
- 1. Open the TSQ Series Dashboard and click Tune Types.
- 2. In the Tune Types dialog box, select a tune type to edit and click **Copy**. See Figure 93.

Figure 93. Tune Types Dialog Box

Tune Types		8 X
El Diagnostics Only (built-in) El Initial Tune (built-in) El SRM Quick Tune (built-in) El SRM Tune (built-in) El Standard Quick Tune (built-in) El Standard Tune (built-in)	•	New Edit
El Target Tune (built-in) El Tune Check (built-in) El SRM Factory Tune (built-in)	Ŧ	Rename
Runs a complete set of diagnostics and generate a report. No tuning is performed. Starts with last saved tune.	*	Delete Close

3. The Tune Type Editor opens. Configure the parameters under the **General** tab in the Tune Type Editor.

Figure 94. Configuring General Tuning Parameters

lit Tune Type		2	X				
Name:	EI Diagnostics Only	Save	;				
Description:	Runs a complete set of diagnostics and generate a report. No tuning is performed. Starts with last saved tune.						
Categories:	El SRM Tune, El Standard Tune, Factory						
	Tune and diagnostics Diagnostics only Show advanced succession Targets Detector Diagnostics Report ename prefix: Diagnostics	ettings					
Starting tune f	ile: (Last Saved)						
Mass calibra	tion n mass calibration						

a. **Name**—Enter a name for your tune type.

Note Create the new tune type name now. You will not be able to edit the tune type name later. Also, you will not be able to save new parameters to a built-in tune.

- b. **Description**—Enter details or notes about your tune type.
- c. **Categories**—This is an optional field that allows you to display a subset of tunes under Auto Tune on the dashboard. This is convenient for separating your most frequently used tunes.
- d. **Type**—Select the **Tune and Diagnostics** option to run a tune with diagnostics or select the **Diagnostics Only** option if you are creating a diagnostics test.
- e. **Output Tune Filename Prefix**—Enter a prefix to be added to the title of your tune report. It is a good idea to use a descriptive prefix for your tune type. For example, if you are always running BFB reports, you could enter BFB here to distinguish it from other reports you are generating.
- f. Starting Tune File—Select a tune file that your tune type will be based on:

Note If you select a tune file with a prefix-only name rather than a tune file with a date-time stamp, the most recent tune file of with that prefix name will be loaded at the start of each tune.

- Select **Factory** to use a default factory-made file on the instrument that can be used to begin tuning an instrument with a clean ion source.
- Select **Last Saved** to use a tune file saved on the instrument by the most recent successful automatic tune.
- Select a specific tune file if you have a reliable tune file you want to use as a starting point for a new tune file.
- g. **Perform Mass Calibration**—Select this to enable the system to recalibrate all of the masses during a tune.
- h. **Check Mass Calibration**—Select this to enable the system to confirm that your mass calibration is correct rather than performing a mass calibration.

Note If Tune and Diagnostics is selected, you can select only one Mass Calibration option.

4. Select the **Ion Source** tab to configure the ion source as follows:

Name:	El Initial Tune							Save
Description:	Requires a clean in: quadrupole voltage	strument. Sta to ixed value	rts with Factory is.	/ tune. Sets de	tector gain to 3x1	0^5. Sets Repeller t	to 0.5 V and	Cancel
Cate <mark>gories</mark> :							=	Print
Type:	Tune and diagno	ostics (Diagnostics	only		V Show	v advanced setting	S
General	Ion Source Target	s Detect	or Diagnos	tics Report	Q1 Lenses	Q1 Resolution	Q3 Resolution	Q3 Lenses
lonization m	node & ion polarity:	El+	•					
		-						
	Cl gas type:	Methane		-				
	CI gas type: CI gas flow:	Methane	mL/min	*				
Emission cu	CI gas flow:		mL/min	Ψ				
Emission cu Electron en	CI gas flow:	0.3						
Electron en	CI gas flow:	0.3 Default	• 50	μA				
Electron en Electron ler	CI gas flow: urrent: nergy:	0.3 Default Default	 ▼ 50 ▼ 70 	μA eV				

Figure 95. Configuring the Ion Source Parameters

- a. Ionization Mode & Ion Polarity—Use this pull-down menu to select a mode:
 - Select **EI** + to run a tune in Electron Ionization (EI) mode.
- b. **Emission Current**—Defines the emission current that you use to run subsequent tunes, but not the emission current that is used for data acquisition.
 - **Tune File**—Select this option to use the value in the tune file that you selected on the General tab.
 - Default—Select this option to use the default emission current, which is 50 µA.
 - **Custom**—Select this option if you want to use a value other than 50 μ A when increasing or decreasing the sensitivity of the instrument. For the emission current, the default is 50 μ A. Tune with the same value you are planning to use for your analysis. The use of emission currents above 100 μ A may lead to the generation of too many ions in the source and a degradation in resolution.
- c. **Electron Energy**—Indicates the energy of the electrons that come off the filament.Lowering the electron energy will extend the lifetime of your filament.
 - Default—Select this option to use the default electron energy, which is 70 eV.
 - **Tune File**—Select this option to use the value in the tune file you selected in the General tab.
 - **Custom**—Select this option to set the energy of the electrons emitted by the filament. For example, you could change the voltage if you wanted to change the ionization efficiency and fragmentation of the sample. This is typically set to 70 V because the standard EI libraries are based on 70 eV electron beams.

Note Avoid reducing the electron energy to less than 70 eV. The calibration compound will not be sufficiently ionized for tuning or calibrating at low electron energies.

Note A custom electron energy can be selected in the instrument acquisition method even if it is not used for tuning.

- d. Electron Lens Positive Voltage—Allows the electrons to enter the ion volume.
 - **Default**—Select this option to use the default electron lens positive voltage, which is 15 V.
 - **Tune File**—Select this option to use the value used in the tune file that you selected on the General tab.
 - **Custom**—Select this option if you do not want to use the default value for the tune. For the electron lens positive voltage, the default is 15 V. You should tune with the same value you are planning to use for your analysis. This value affects the focusing of the electron beam into the source. If you change your electron energy from 70 eV, this value also changes. This voltage must always be at least 45 V above the voltage applied to the filament. The voltage applied to the filament is the same number, but the opposite sign, of the electron energy.

e. Electron Lens Negative Voltage.

- **Default**—Select this option to use the default electron lens negative voltage, which is -75 V.
- **Tune File**—Select this option to use the value used in the tune file that you selected on the General tab.
- **Custom**—Select this option if you do not want to use the default value for the tune. For the electron lens negative voltage, the default is -75 V. Tune with the same value you are planning to use for your analysis. This value affects how well the electrons are kept from entering the ion source when they are not supposed to. If you change your electron energy from 70 eV, this value also changes. Ensure this voltage is at least 5 V below the voltage applied to the filament.
- f. **Set Ion Source Temperature**—Select this check box to enable the temperature field. Then enter a value between 0 and 350 °C. The default is 200 °C. The optimal temperature is determined by the molecular structure and weight of the compounds you are analyzing. Heavier compounds require a higher temperature. Tune at the same temperature you will use to run your samples. You should set the temperature as high as possible to keep the ion source clean and maintain the right amount of sensitivity.

Tip If you tune regularly between running sample sets, you can save time waiting for the temperatures to equilibrate by setting the tune temperatures to the same temperature used in your acquisition method.

5. Click the **Targets** tab to configure how you want to tune your targets. Target tuning is used to adjust the way the mass spectrometer tunes to meet regulatory requirements. Otherwise, this is an optional setting.

Figure 96. Configuring Tuning Targets

ame:	El Initial T	une			Save
escription:			ninstrument. Starts with Factory tune. Sets detector gain to 3x10^5. Sets Repeller to 0 V, Q1 nd Q3 Voltage to -3 V. Higher resolution and lower sensitivity than El Standard Tune.	1	Cancel
ategories:	Factory				Print
ype:	Tune a	and dia	Ignostics 🔘 Diagnostics only 📃 Show advance	ed settings	
ieneral	lon Source	Tar	gets Detector Diagnostics Report		
Tune targ	et ion ratios	11.10	gets Delector Diagnostica Fréport		
Tune targ	et ion ratios Ratio	11.0			
		%	(25 to 51.5 amu)		
arget mass	Ratio				
arget mass 50 amu:	Ratio	%	(25 to 51.5 amu)		
arget mass 50 amu: 69 amu:	Ratio 1.1 100	%	(25 to 51.5 amu) (51.5 to 78.5 amu)		
arget mass 50 amu: 69 amu: 131 amu:	Ratio 1.1 100 90	% %	(25 to 51.5 amu) (51.5 to 78.5 amu) (78.5 to 132.5 amu)		

Tune Target Ion Ratios—Select this check box to adjust the ratios based on the results from an injection of tuning compound.

6. Click the **Detector** tab to configure the detectors. See Figure 97.

Figure 97. Configuring the Detectors

Edit Tune Type		? ×
Name:	El Initial Tune	Save
Description:	Requires a clean instrument. Starts with Factory tune. Sets detector gain to 3x10^5. Sets Repeller to 0 V, Q1 Voltage to -3 V and Q3 Voltage to -3 V. Higher resolution and lower sensitivity than El Standard Tune.	Cancel
Categories:	Factory	Print
Туре:	Tune and diagnostics O Diagnostics only	
General Io	n Source Targets Detector Diagnostics Report	
👿 Set initial o	letector gain: 3 × 10 ⁵	
Tune dete	ctor	
Adjust det	ector sensitivity	
Mass:	69 v Intensity: 10,000,000	

a. Set Initial Detector Gain—Select this option to set the electron multiplier gain that will be used in tuning the instrument. The gain is the number of electrons generated for every ion that strikes the detector. This is typically set at 3×10^5 electrons per ion. Gains larger than this will generate more electrons per ion, but both the analyte ion and the noise ion signals will be larger. You can also tune to lower gain values, which

decreases the signal strength. Lower values also increase the chance that an ion will not be detected.

- b. **Tune Detector**—Select this check box to tune the detector. As the electron multiplier ages, the voltage required for a given gain increases. Depending on your sample load and if your system is leak tight (oxygen is bad for the detector), you should not have to perform this tune very often. Limit to 1–2 times a year.
- c. Adjust Detector Sensitivity—Select this check box to tune the detector to generate a consistent area count of a calibration gas ion for the tune report. Because the intensity of the cal gas varies depending on the atmospheric pressure and temperature of the lab, this option results in larger variation in the analytical runs, as compared to using a fixed detector gain. If you use this function, set the initial detector gain to 3×10^5 electrons per ion.
 - Mass—Select the calibration gas mass you want to use.
 - Intensity—Enter the intensity you want to see on the tune report.
- 7. Click the **Diagnostics** tab and select a test to confirm the operational ability of the system. See Figure 98.

Figure 98. Setting Diagnostics

Edit Tune Type					? ×
Name: Description: Categories; Type:	El Initial Tune Requires a clean instru Voltage to -3 V and Q3 Factory Tune and diagnosti	ment. Starts with Factory tune. Sets de Voltage to -3 V. Higher resolution and cs Diagnostics only	etector gain to 3x10^5. Sets I lower sensitivity than EI Sta	s Repeller to 0 V, Q1	Save Cancel Print
Communic Detector 0 Filament C Uon Guide V Leak Chee Lens Chet Lens Cont Power Suj Q1 Freque Q1 Transs Q2 Freque Q3 Freque Q3 RF/D0 SRM Effic Temperati	heck Frequency Check ck inuity Check pply Check ancy Check System Check inission Check ancy Check ancy Check System Check iency Check iency Check	Detector Diagnostics Repo	nt		
					.#

8. Click the **Report** tab configure how you want to view your data

Figure 99. Selecting Data Report Options

Edit Tune Type		? ×
Name:	El Initial Tune	
Name:		Save
Description:	Requires a clean instrument. Starts with Factory tune. Sets detector gain to 3x10^5. Sets Repeller to 0 V, Q1 Voltage to -3 V and Q3 Voltage to -3 V. Higher resolution and lower sensitivity than EI Standard Tune.	Cancel
Categories:	Factory	Print
Туре:	Tune and diagnostics Image: Diagnostics only Image: Diagnostics only <td></td>	
General lo	In Source Targets Detector Diagnostics Report	
📄 Acquire fu	II-scan spectrum	
Acquisiti	on threshold: 3.000	
SIM masses:	69 • 131 • 219 • 414 • 502 •	

- a. Acquire Full-Scan Spectrum—Select this check box to display the full spectrum on your tune report.
- b. **Acquisition Threshold**—Enter a minimum peak height for the full scan spectrum on the tune report. If your peak has an intensity that is below this threshold, it will not be stored.
- c. **SIM Masses**—Select masses to be displayed on your tune report. You can select a maximum of five masses, one from each list. At least one mass must be displayed on the report.

Tip Factory tunes cannot be altered. If a newly created tune does not perform as expected, you can always go back to a factory tune, copy it, and change the settings as described above. Save the new tune with a new name before closing the Edit Tune Type dialog box.

- 9. Once you are finished configuring all the tabs, click **Save** to save the tune type. Your new tune type appears in the Tune Types window.
- 10. Click **Close** to return to the dashboard. You can select the tune type in the automatic tune window.
- 11. If you want to rename a tune you created, open the Tune Types dialog box by clicking **Tune Types** on the TSQ Series Dashboard.

Note You cannot rename a factory tune.

a. Select **Rename** in the Tune Types dialog box.

Figure 100. Renaming a Tune Type

Tune Types		? ×
EI Standard No Before EI Standard Quick Tune EI Standard Tune Just Lens Check Leak Check Diag Leak Check Lens Contin only MODIFIED EI STANDARD My tune type	•	New Edit Copy Rename
Leak check diag	*	Delete Close

b. The **Rename Tune Type** dialog box opens. Assign a valid new name to your tune. You will not be able enter a name that is an invalid file name or already in use. See Figure 101.

Figure 101. Changing a Tune File Name

Rename tune type - Leak	Check Diag	X
New name for tune type:	Leak Check Diag	
	Ok <u>C</u> ancel	

c. Click OK.

Note The following instructions refer to advanced tuning settings that most users will never need to access. Contact your local Thermo Fisher Scientific service organization before adjusting these settings.

To modify advanced tune settings

1. To configure the **Q1 Lenses**, you must select the **Show Advanced Settings** check box. The lens tune is the main portion of the tune algorithm. In this section you can choose which components to tune, which mass to optimize for the tune, what the range of allowed values is, how to move through the range, how much the signal must change for a new value to be selected, and what (if anything) can be done about the resolution of the peak and any errors that occur. Select the **Tune Lenses** check box to access the data table. See Figure 102.

Figure 102. Tuning the Q1 Lenses

Name	e: El Init	al Tune												Sa	ve
Descr									0^5. Sets Repel an El Standard				÷ (Can	cel
Categ	gories: Facto	Factory													
Type:	: • Tu	ne and diagno	stics	Dia	aqnostics	only			🔽 S	how	advance	d settings	s		
			. D.I.		Diagnost	tice F	Report	Q1 Lenses	Q1 Resolutio		Q3 Res				
Gene	ral Ion Sou	ce Targets	s Dete	CLOF	Diagnosi		Jebour	G I Lenses	Grincooldio		Q3 Res	olution	Q3	Lenses	
	eral Ion Sou une lenses	ce Targets	s Dete	ctor	Diagnosi	100	iepoir]	QT Lenses	GT HOSPILIE	•• (Q3 Resi	olution	Q3	Lenses	
Gener	1	ce Targets Mass	s Dete Start	Stop	Step	Max. Width	Measure		On Error		Q3 Hes	olution	Q3	_	
	une lenses	1 -	I			Max.	Measure					olution	Q3	_	Í
	une lenses Device	Mass	Start	Stop	Step	Max. Width	Measure at %	Threshold	On Error			olution	Q3I	_	Í
	Device Repeller	Mass 219 69	Start 0	Stop 0	Step 0	Max. Width 2	Measure at %	Threshold	On Error Continue				Q3I	_	
	Device Repeller Lens 1	Mass 219 69 olt 502	Start 0 -50	Stop 0 -50	Step 0 0	Max. Width 2 2	Measure at % 50 50	Threshold 1 1	On Error Continue Continue		Q3 Hes		Q3I	_	Í
	Device Repeller Lens 1 Ion Guide V	Mass 219 69 olt 502 olt 219	Start 0 -50 -15	Stop 0 -50 -1	Step 0 0 0.25	Max. Width 2 2 2	Measure at % 50 50 50	Threshold 1 1 1	On Error Continue Continue Continue		Q3 Hes		Q3I	_	Í
	Device Repeller Lens 1 Ion Guide V Ion Guide V	Mass 219 69 olt 502 olt 219 olt 69	Start 0 -50 -15 -10	Stop 0 -50 -1 1	Step 0 0.25 0.2	Max. Width 2 2 2 2 2	Measure at % 50 50 50 50	Threshold 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	On Error Continue Continue Continue Continue				Q3I	_	Í

- a. **Device**—Click in this box to select the component you want to tune according to the settings in the other columns.
 - i. **Repeller**—Controls how the repeller pushes the ions out of the ionization region. The voltage applied to this component has a very strong effect on the energy of the ion beam, which has a strong effect on the resolution and the intensity. The lower the voltage, the better the resolution. However, higher voltages will prevent ions from striking the repeller surface, which leads to better robustness. A clean system typically tunes from 3 to 8 V, although a dirty system may have the repeller climb as high as 12.5 V. Do not set the repeller any higher.
 - ii. Lens 1—Controls Lens 1, which is the first of three lenses that the ions see as they leave the ionization region. These three lenses act as a focusing element to maximize the ion beam intensity that is entering the ion guide. This field is typically set between -35 and -50 V.
 - iii. **Lens 2**—Controls Lens 2, which is the second of three lenses that the ions see as they leave the ionization region. These three lenses act as a focusing element to maximize the ion beam intensity that is entering the ion guide. This field is typically set between 0 and -15 V.
 - iv. **Lens 3**—Controls Lens 3, which is the last of three lenses that the ions see as they leave the ionization region. These three lenses act as a focusing element to maximize the ion beam intensity that is entering the ion guide. This field is typically set between -30 and -35 V.
 - v. **Ion Guide Voltage**—Controls the ion guide's DC offset voltage. It can potentially help focus the ions into the quadrupole while ensuring that neutral noise is eliminated. The voltage on this component is mass dependent and

should be set at several different masses. This field is typically set between +1 and -15 V, depending on the mass of the ion.

- vi. **Ion Guide RF q**—Controls the ion guide's RF q (amplitude). This device is mass dependent and should be set at several different masses. This field can be tuned between 0 and 2. This device determines the ion guide RF amplitude tune value which is given as the RF q * (m/z).
- vii. **Ion Guide RF**—(This setting is maintained for compatibility with legacy tune types. For new tune types, Ion Guide RF q is the recommended setting.) Controls the ion guide's RF voltage. It can potentially help focus the ions into the quadrupole while ensuring that neutral noise is eliminated. The voltage on this component is mass dependent and should be set at several different masses. This field is typically set between 0 and +5 V, depending on the mass of the ion.
- viii. **Q1 Voltage**—Controls the voltage that pulls the ions into the quadrupole. The voltage applied to this component has a very strong effect on the energy of the ion beam, which has a strong effect on the resolution and the intensity. The lower the voltage, the better the resolution. However, higher voltages will pull more ions into the quads, which leads to better signal. This field is typically set between 0 and -5 V.
- ix. **Q1 Resolution**—Adjusts the ratio of the quadrupole DC and RF voltages to create the resolution required for your analysis. You can set the desired peak width at a given mass and whether you measure the width at 10% or 50% of the peak height. Because there is no static DC voltage involved, the start, stop, and step values are not used.
- b. Mass—Click in this box to select the ion to be used for tuning.
- c. **Start**—Enter the starting voltage for the tune. The start voltage must always be less than the stop voltage. For example, -35 is smaller than 0.
- d. **Stop**—Enter the final voltage for the tune.
- e. **Step**—Enter the increment for the tuning range. For example, if you tune from 0-50 in increments of 10 V, then you would set the **Step** field to 10.

Note If the start and stop values are the same value, the device will be set to that value and ignore the step size. In some tunes, devices are set to a value before tuning them in another line. This presetting is done because ramping devices do not allow the tune to run at values higher than those chosen for higher masses for the same device.

- f. Max Width—Enter the maximum allowable width of the ion during the tune.
- g. **Measure at %**—Click in this box to select the location on the peak where you want to measure the maximum width.
 - **10**—Measures the width at 10% of the peak height.
 - **50**—Measures the width at 50% of the peak height.

h. **Threshold**—Enter the change in intensity that has to occur when the tune selects a new voltage. For example, if you set this field to 1.1, the tune will not select a new voltage for that component unless the intensity is 110% of the old intensity. If you set this field to 1, anytime the new voltage has an intensity larger than the old intensity, the tune will select the new intensity.

Note If you choose a value less than 1 for ramped devices, the tune steps back in energy until the lower intensity is detected. For example, if you choose 0.8, the device value lower in energy than the device value with the maximum response that gives an intensity closest to 80% of the maximum intensity is chosen.

- i. On Error—Click in this box to select how to handle an error in the tune.
 - **Fail**—Stops the tune when an error occurs or if no tune points are found meeting the tune criteria.
 - **Continue**—Allows the tune to continue on to the next device when an error occurs.
- j. **Entrance lens offset** and **Exit lens offset**—Changing these voltages affects ion acceleration. The default levels will give the best performance; changing the voltages will affect peak resolution.
- 2. Click the **Q1 Resolution** tab to configure the resolution of your tune. You must select the **Show Advanced Settings** check box to access this tab. Selecting this option tunes the resolution of Quadrupole 1 by itself. This resolution tunes the system at 100 and 1000 amu/s scan rates. You can tune the resolution during a lens tune or you can tune the resolution by itself.

Figure 103. Configuring Q1 Resolution

	EII	nitial Tune						Save	
Descri		Requires a clean instrument. Starts with Factory tune. Sets detector gain to 3x10^5. Sets Repeller to 0.5 V and quadrupole voltage to ixed values.							
Categ	ories:							Print	
-	0	Tune and diagnostic	s 🔘 Diagno:	stics only		Show advanced	acttings		
Type:		Tune and diagnostic	s O Diagno:	SUCS ONLY			acturiya		
Gener	ral Ion So	ource Targets	Detector Diag	gnostics Report Q1 L	enses Q1 Re	solution Q3 Resol	lution Q	3 Lenses	
					enses Q1 Res	solution Q3 Resol	lution Q	3 Lenses	
	ral Ion So ne resolution		Detector Diag		enses Q1 Re	solution Q3 Resol	lution Q	3 Lenses	
					enses Q1 Re	solution Q3 Resol	lution Q	13 Lenses	
	ne resolution	ı 🕅 Tı	une high scan rate:	10000 amu/sec	enses Q1 Re	solution Q3 Resol	lution Q	13 Lenses	
	ne resolution	Peak Width	une high scan rate: Measure at %	10000 amu/sec	enses Q1 Re	solution Q3 Resol	lution Q	13 Lenses	
	Mass	Peak Width 0.7	une high scan rate: Measure at %	10000 amu/sec Lens Tune Relation Before and After	enses Q1 Re	solution Q3 Resol	lution Q	13 Lenses	
	Mass 69 219	Peak Width 0.7 0.7	Measure at % 50 50	10000 amu/sec Lens Tune Relation Before and After Before and After	enses Q1 Re	solution Q3 Resol	lution Q	13 Lenses	

Note Changing the Q1 lenses or resolution affects the Q3 lenses and resolution.

a. Mass—Click in this box to select the ion to be used for tuning.

- b. **Peak Width**—Enter the target peak width.
- c. **Measure at %**—Click in this box to select the location on the peak where you want to measure the target peak width.
 - **10**—Measures the width at 10% of the peak height.
 - **50**—Measures the width at 50% of the peak height.
- d. **Lens Tune Relation**—Click in this box to set the occurrence of the resolution tune parameters before or after a lens tune.
 - **Before and After**—Uses the same resolution parameters before and after a lens tune.
 - **Before**—Uses the resolution parameters before a lens tune.
 - After—Uses the resolution parameters after a lens tune.
- 3. Select the **Q3 Lenses** tab to configure the Q3 lenses. You must select the **Show Advanced Settings** check box to access this tab. The lens tune is the main portion of the tune algorithm. In this section you can choose which components to tune, which mass to optimize for the tune, what the range of allowed values are, how to move through the range, how much the signal must change for a new value to be selected, and what (if anything) must be done about the resolution of the peak and any errors that occur.

Figure 104. Tuning the Q3 Lenses

Edit Tune	Туре						-				? ×
Name: El Initial Tune Description: Requires a clean instrument. Starts with Factory quadrupole voltage to ixed values.							ts detector	r gain to 3x1(0^5. Sets Repel	llerto 0.5 V and	Save Cancel
Catego	nies:					Print					
Type:	Type: Tune and diagnostics Diagnostics only Show advanced settings							js			
Genera	al Ion Source	Targets	Dete	ctor	Diagnost	ics F	Report	Q1 Lenses	Q1 Resolutio	Q3 Resolution	Q3 Lenses
📝 Tun	ne lenses										
	Device	Mass	Start	Stop	Step	Max. Width	Measure at %	Threshold	On Error		
+	Q3 Voltage	502	-5	-5	0	2	50	1	Continue		
	Q3 Voltage	219	-4.35	-4.35	0	2	50	1	Continue		
	Q3 Voltage	69	-4	-4	0	2	50	1	Continue		
*											
	ice lens offset:	5 Vo	lts E	Exit lens o	offset: -	1	Volts				
-MS/	MS/MS Voltages										
		Volts									
	69.00	-2.50									
	219.00	-3.00									
	502.00	-7.00									

- a. Q3 Voltage—Controls the voltage that pulls the ions into the quadrupole. The voltage applied to this component will have a very strong effect on the energy of the ion beam, which will have a strong effect on the resolution and the intensity. The lower the voltage, the better the resolution. However, higher voltages will pull more ions into the quads, which leads to better signal. This field is typically set between 0 and -5 V.
- b. **Resolution**—Adjusts the ratio of the quadrupole DC and RF voltages to create the resolution required for your analysis. You can set the desired peak width at a given mass and whether you measure the width at 10% or 50% of the peak height. Because there is no static DC voltage involved, the start, stop, and step values are not used.
- c. Mass—Click in this box to select the ion to be used for tuning.
- d. **Start**—Enter the starting voltage for the tune. The start voltage must always be less than the stop voltage. For example, -35 is smaller than 0.
- e. **Stop**—Enter the final voltage for the tune.
- f. **Step**—Enter the increment for the tuning range. For example, if you tune from 0-50 in increments of 10 V, then you would set the **Step** field to 10.
- g. Max Width—Enter the maximum allowable width of the ion during the tune.
- h. **Measure at %**—Click in this box to select the location on the peak at which you want to measure the maximum width.
 - 10—Measures the width at 10% of the peak height.
 - 50—Measures the width at 50% of the peak height.
- i. **Threshold**—Enter the change in intensity that has to occur when the tune selects a new voltage. For example, if you set this field to 1.1, the tune will not select a new voltage for that component, unless the intensity is 110% of the old intensity. If you set this field to 1, anytime the new voltage has an intensity larger than the old intensity, the tune will select the new intensity.
- j. **On Error**—Click in this box to select how to handle an error in the tune.
 - Fail—Stops the tune when an error occurs.
 - **Continue**—Allows the tune to continue on to the next device when an error occurs.
- k. **Entrance lens offset** and **Exit lens offset**—Changing these voltages affects ion acceleration. The default levels will give the best performance; changing the voltages will affect peak resolution.
- 1. **MS/MS Voltages**—These overwrite the Q3 voltage tune values when the instrument is in SRM mode. These values are not tuned and are added to the collision energy used in the SRM scan.
- 4. Click the **Q3 Resolution** tab to configure the resolution of your tune. You must select the **Show Advanced Settings** check box to access this tab. Select this option to tune the resolution of Quadrupole 3 by itself. This resolution will tune the system at 100 and

1000 amu/s scan rates. You may also tune at a higher scan rate. You can tune the resolution during a lens tune or you can tune the resolution by itself. See Figure 105.

Figure 105. Configuring the Q3 Resolution

Edit Tu	ne Type				Later later				? <mark>×</mark>
Nan	Name: El Initial Tune								Save
Des	cription:	Requires quadrupo	Cancel						
Cate	Categories:								Print
Тур	e:	Tune	and diagnostics	Diagnos	stics only		Show	advanced settings	3
Ger	neral lo	n Source	Targets [Detector Diag	nostics Report (Q1 Lenses Q1	Resolution	Q3 Resolution	Q3 Lenses
V	Tune resol	ution	🔲 Tun	e high scan rate:	10000 amu/sec				
	Mas	s	Peak Width	Measure at %	Lens Tune Relation				
+	69		0.96	10	After				
	219		0.96	10	After				
	502		0.96	10	After				
*									

- a. Mass—Click in this box to select the ion to be used for tuning.
- b. Peak Width—Enter the target peak width.
- c. **Measure at %**—Click in this box to select the location on the peak at which you want to measure the target peak width.
 - 10—Measures the width at 10% of the peak height.
 - 50—Measures the width at 50% of the peak height.
- d. **Lens Tune Relation**—Click in this box to set the occurrence of the resolution tune parameters before or after a lens tune.
 - **Before and After**—Uses the same resolution parameters before and after a lens tune.
 - Before—Uses the resolution parameters before a lens tune.
 - After—Uses the resolution parameters after a lens tune.
- 5. **Tune High Scan Rate**—Select this check box to tune the resolution at a scan rate in addition to the 100 and 1000 amu/s default scan rates.

Computer Settings

Your TSQ Duo system includes a computer with all the software needed to operate the instrument.

If you replace the computer in the future, use the information in this chapter to change the computer settings so your TSQ Duo system and the associated Thermo Scientific software will run correctly. Instructions for installing Thermo Scientific software are included on the software CD shipped with your instrument.

Contents

- System Requirements
- Computer Settings
- Excluding the Xcalibur Directory from Virus Scan

System Requirements

The TSQ Duo system requires a Windows 7 32-bit or 64-bit operating system. See "System Requirements" on page xi for a complete list of system requirements.

Computer Settings

The following computer settings will help prevent communication interruptions in your TSQ Duo system.

- To use the power save feature
- 1. From the Start menu choose Control Panel | System and Security | Power Options.
- 2. On the left side, click Change When the Computer Sleeps.
- 3. For **Put the Computer to Sleep**, select **Never**. See Figure 106. You may set the display to turn off at any time. It will not affect the performance of your TSQ Duo system.



🕒 🖉 🖗 🕨 Control Panel 🔸 Hardware and Sound 🔸 Power Options 🔸 Edit Plan Settings							
Change settings for the plan: Balanced Choose the sleep and display settings that you want your computer to use.							
Turn off the display: Put the computer to sleep:	1 hour 30 minutes Never						
Change advanced power setting Restore default settings for this							

✤ To use the virtual memory feature

- 1. From the Start menu, choose Control Panel | System and Security | System.
- 2. On the left side of the window, click Advanced System Settings.
- 3. In the System Properties dialog box, click Settings.
- 4. In the Performance Options dialog box, click the **Advanced** tab, and then click **Change** to open the Virtual Memory window.
- Confirm that Automatically Manage Paging File Size for All Drives check box is selected. See Figure 107.

Figure 107. Virtual Memory Settings

System Properties	Virtual Memory
Computer Name Hardware Advanced System Protection Remote You must be logged on as an Administrator to make most of these changes. Performance Visual effects, processor scheduling, memory usage, and virtual memory Settings	Automatically manage paging file size for all drives Paging file size for each drive Drive [Volume Labe] Paging File Size (MB) C: [OSDisk] System managed
User Profiles Desktop settings related to your logon Settings	Selected drive: C: [OSDisk] Space available: 227140 MB Custom size: Initial size (MB): Maximum size (MB):
Startup and Recovery System startup, system failure, and debugging information	System managed size No paging file Set
Settings Environment Variables	Total paging file size for all drives Minimum allowed: 16 MB Recommended: 30 19 MB Currently allocated: 2669 MB
OK Cancel Apply	OK Cancel

✤ To confirm the network card firewall settings

- 1. From the Start menu, choose Control Panel | Network and Internet | System and Security | Windows Firewall | Advanced Settings.
- 2. In the Actions Panel on the right, (see Figure), select Properties.
- 3. In the Windows Firewall with Advanced Security on Local Computer Properties window, select Domain Profile | State | Customize.

Figure 108. Firewall Settings for the TSQ Duo System Computer

Windows Firewall with Advanced Security			
File Action View Help			
Windows Firewall with Advance Windows Firewall with Advance Commettion Rules Monitoring Monitoring Windows Firewall with Monitoring Windows Firewall with Monitoring Windows Firewall Windows Firewall	Windows Fiewall with Advanced Security on Local Computer Pro Domain Profile Provate Profile Public Profile (Piece Settings Specify behavior for when a computer is connected to its corporate domain. Safe Preval state: On (seconnected to its corporate Indownd connections: Block (idefault) Outbound connections: Block (idefault) Protected network connections: Customize Protected network connections: Customize Settings Specify logging settings for toubleshooting. Learn more about these settings OK Cancel Acopy toubleshooting and activity A currently applied frewall and connection security vies and security associations for	Actions Windows Firewall with Advanced Security on Local ▲ Import Policy Restore Default Policy Diagnose / Repair View Refresh Properties Help 	Actions Panel

4. In the **Protected Connections for the Domain Profile** window, make sure that any instrument checkboxes are not checked.

Figure 109. Setting the Network Connections

1.00

		th Advanced Security on Local Computer	Actions	
Contection Rules	Private Profile Windows Free kibound come Cutbound come	Windows Firewall with Advanced Security on Local Computer Pro	Windows Firewall with Advanced Security on Local. Import Policy Restore Orlaw Policy Diagnose / Repair View Refersh Properties Help	•
	View and cr Create frewal rul t is authenticated blocked unless the blocks them.	Learn non about these settings OK Cancel OK Cancel	Π.	

Excluding the Xcalibur Directory from Virus Scan

If you are using anti-virus software on your computer, exclude the following folders from scanning:

- C:\Program Files\Thermo\
- C:\Thermo\
- C:\Xcalibur\

Failure to exclude these folders might cause interruptions in data acquisition. If you must, for security reasons, scan all system files, set the virus scans so that they occur when you are not using the instrument to acquire data.

Troubleshooting

This section contains information to help you diagnose problems within your data. Sometimes, your experience as a scientist will enable you to look at your data and detect that something is wrong either with the instrument or your sample. This chapter describes the most common indications of a problem with a baseline, peak or result.

Contents

- Setting Instrument Conditions for Troubleshooting
- Checking Air/Water Spectra
- Diagnostics Checks
- How to Know When Your System Needs Maintenance
- Investigating Baseline Issues
- Investigating Peak Issues
- Investigating Results Issues

If there is an issue with the hardware, see the troubleshooting section of the hardware manual.



CAUTION ELECTRICAL SHOCK HAZARD: When troubleshooting any issue that requires removing a cover on the instrument, power-off and vent the instrument to avoid possible injury.

You can begin troubleshooting by running a tune with diagnostics on the instrument. If you have good ion intensities, good peak shapes, and no air leak, you might want to look first at the GC, autosampler, or carrier gas.

If you have air leaks, locate and address them. Pay particular attention to the transfer line ferrule, vent valve knob, front panel, gas quality, carrier gas vacuum compensation, and vacuum interlock on the mass spectrometer, as well as the inlet on the GC.

If your intensities are too low, make sure carrier vacuum compensation is turned on.

IMPORTANT When inserting a cold ion source cartridge such as after cleaning or when switching between EI and CI modes, the ion source and lens stack will expand as the source cartridge heats, often pushing the ion volume and lenses away from the rear of the instrument where they are firmly held by the RF Lens spring contacts. To avoid intermittent electrical contacts to the lenses, you should insert the ion source cartridge, wait 30 min for it to get to temperature, then remove and reinsert it. See the hardware manual for instructions on cleaning and inserting the ion source cartridge.

If you have no ions in full-scan or SIM mode, do the following:

- Switch filaments.
- Confirm the starting tune file is appropriate for ionization mode or source cleanliness level.
- Confirm the correct ion volume is inserted (EI or CI).
- Verify the ion source is correctly installed.
- Run diagnostic checks (Diagnostics Only tune type)
- Clean the ion source.

If you do not see ions in CI mode:

- Confirm that the CI ion volume is installed.
- Confirm that the reagent gas is connected and turned on.

If you do not see any peaks in SRM scans, do the following:

- Verify the collision gas supply.
- Run diagnostics checks (Diagnostics Only tune type).
- Run a full-scan method to check if your system is functional.

Setting Instrument Conditions for Troubleshooting

Before troubleshooting the TSQ Duo system, set the instrument to the conditions in this section in order to compare your system more accurately to the values in the section. All troubleshooting should be performed in EI mode. Once EI mode is working, check CI conditions if relevant.

IMPORTANT Use only Nitrile Cleanroom gloves when touching ion source components. Other types of gloves deposit contaminants on the source components that affect system performance. See the *TSQ Duo Spare Parts Guide* for ordering information.

• Clean the ion source cartridge. See the TSQ Duo Hardware Manual for instructions.

- Install a 15 m \times 0.25 mm \times 0.25 μm GC column (If using a different column, pressure readings may vary.)
- Ion Source Temperature 200 °C
- Transfer Line Temperature 250 °C
- Vacuum Compensation On
- Column Flow Rate 1.5 mL/min
- Foreline Pressure < 100 mTorr

Note Foreline Pressure is a function of how long the interior of the manifold has been exposed to the atmosphere, the pumping capacity of the turbo pump, length of the foreline hose, and other criteria. As an example, if the system has been recently exposed to atmosphere, the foreline pressure will be above the expected value. Foreline pressure is not set by the user, but the value above needs to be reached prior to troubleshooting.

- Pump down the system according to the time recommended for the turbomolecular pump installed on it.
- Air/Water Check Water (*m*/*z* 18/69) < 300%

Once you have applied the settings in this section, and have allowed the TSQ Duo system to equilibrate, run an EI diagnostic tune even if you cannot see any ion intensities.

Checking Air/Water Spectra

Before running additional diagnostics, check the air/water spectra of your TSQ Duo system in the TSQ Series Manual Tune utility and use the information in this section to troubleshoot your system.

- * To check Air/Water spectra on the TSQ Duo system
- 1. Open the TSQ Series Dashboard. Check that all the Status indicators are green and that the turbo pump is set at 100%.
- 2. Select TSQ Series Manual Tune and check the Air/Water spectrum. See Figure 110.

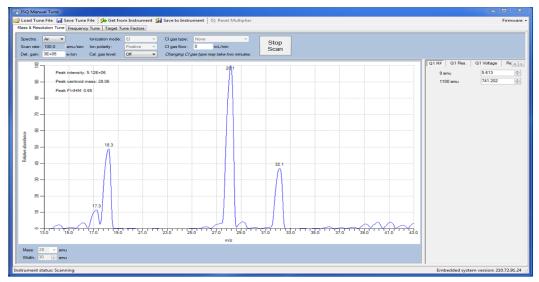


Figure 110. Typical TSQ Duo Air/Water Spectrum for a System with Helium Carrier Gas

The correct settings should be:

- Detector Gain = 3e5
- Peak Intensity > 3e6
- Normal ions are present and in acceptable ratios:
 - 18 (Water) 20–300% of N₂
 - 28 (N₂) Reference (base) Peak
 - 32 (O₂) 10-40% of N₂
 - 40 (Argon) <10% of N₂

Note Several factors strongly influence the ion ratios listed above: tune file, gas flow, gas quality, temperature, and time from pump down.

3. Using hydrogen as a carrier gas changes the air/water spectrum on the TSQ Duo system. It general more background peaks due to the increased reactivity of the hydrogen gas with the components inside of the sample path. See Figure 111 for an image of a typical TSQ Duo air/water spectrum when hydrogen is used as a carrier gas.

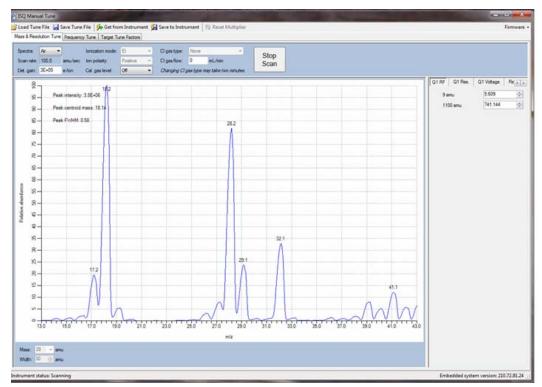


Figure 111. Typical TSQ Duo Air/Water Spectrum for a System with Hydrogen Carrier Gas

The following conditions can cause changes in the air/water spectrum on the TSQ Duo instrument.

- 1. Standard detector gain is equal to 3e5, but this can vary depending on customer defined tunes.
- 2. As the instrument pumps down over time, the ratio of 18/28 will change as m/z 18 decreases with m/z 28 remaining constant. This eventually changes m/z 28 to the base peak.
- 3. Changing components of the system such as the column, ion source, or gas supply affects the different masses present in the air/water spectra.
- 4. Maximum intensity may vary based on different instrument parameters, such as changing the column flow, or accessories.

If any of the previous conditions are not met and a leak is suspected as the root cause, follow "An air leak has been detected" in the *"Investigating Vacuum Issues*" section of the **TSQ Duo Hardware Manual** or "How to Know When Your System Needs Maintenance" on page 121.

The next several images show a typical air/water spectrum for several common problems.

1. Figure 112 is an example of an air/water spectrum of a system with a potential air leak.

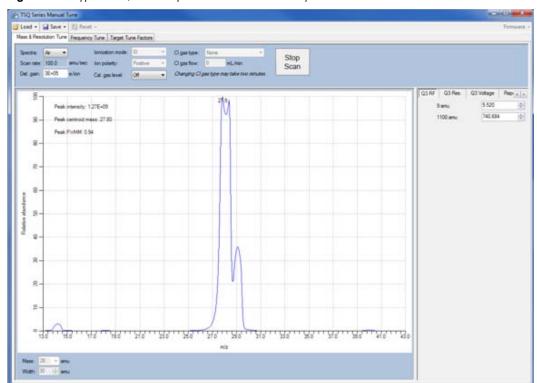
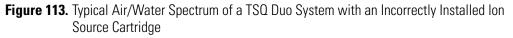
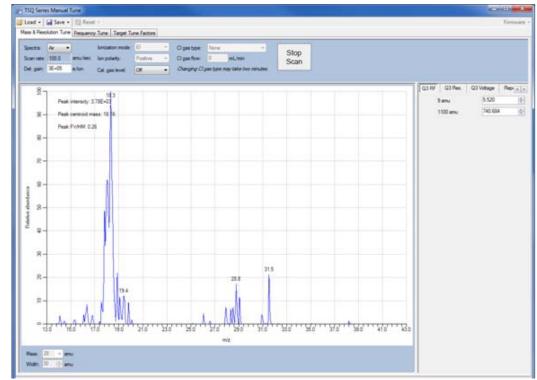


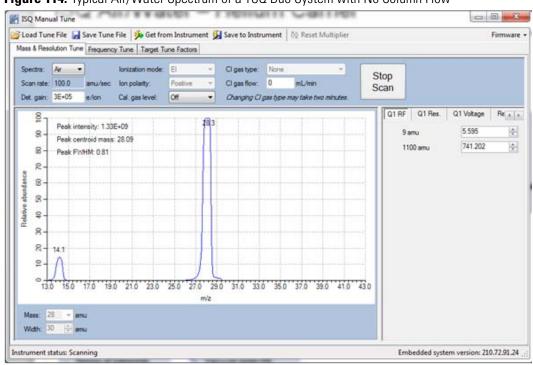
Figure 112. Typical Air/Water Spectrum of a TSQ Duo System with an Air Leak

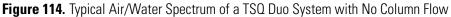
2. Figure 113 is an example of a system with an incorrectly installed ion source.





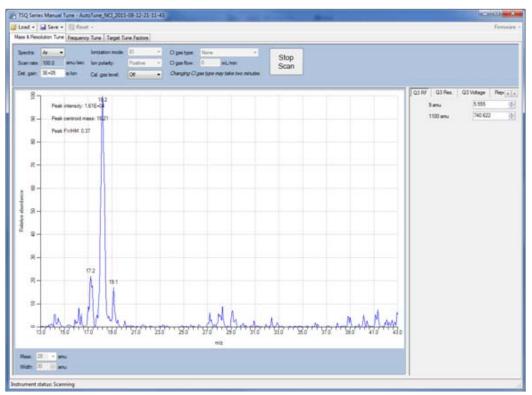
3. Figure 114 is an example of a system with no column flow.





4. Figure 115 is an example of a spectrum with a CI tune file run with an EI ion source.

Figure 115. Typical Air/Water Spectrum of a TSQ Duo System with an El Ion Source Run with a Cl Tune File



Diagnostics Checks

The following table provides you with possible corrective actions you can take after failing a diagnostics check on your system.

Note If you are unsure about what to do if your system fails a diagnostic, please contact your field service engineer.

Diagnostic	Cause of Failure	Corrective Action
Communication Check	Unknown	Contact your Field Service Engineer.
Frequency Check	RF Dip is not correct	Run a full tune that will correct for any RF dip problems.
		Ensure source cartridge is installed.
		Contact your Field Service Engineer.
SRM Efficiency Check	Collision gas is not supplied correctly	Check collision gas tank and pressure.
	Q1 is poorly tuned	Run an EI SRM tune (not a quick tune).
Lens Continuity Check	Ion source not inserted properly	Remove and reinsert ion source.
	Lens plate and/or springs are damaged	Contact your Field Service Engineer.
	A wire is disconnected or shorted inside the manifold	Contact your Field Service Engineer.

 Table 2.
 Diagnostics and Corrective Actions

How to Know When Your System Needs Maintenance

Typically, you will notice that your instrument needs maintenance when you are analyzing your data on the computer. Figure 116 shows a normal full scan background on a TSQ Duo system.

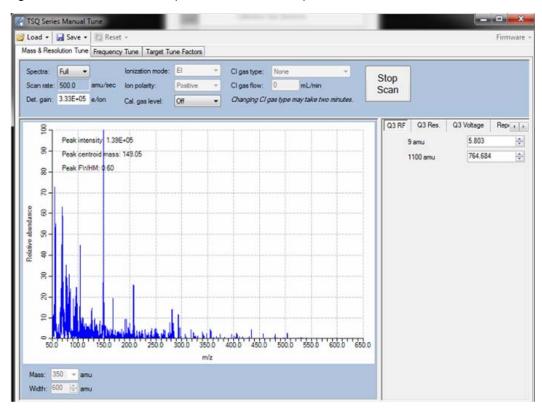


Figure 116. Normal Full Scan Spectrum of a TSQ Duo System

A normal system should have the following conditions met;

- Detector gain 3e5
- Maximum intensity < 1e6
- There should be exponential decay for background noise.
- There should be no extraneous peaks indicating contamination.

If you run a sample with Perfluorotributylamine (PFTBA), the tuning compound, the spectrum should look like Figure 117 below.

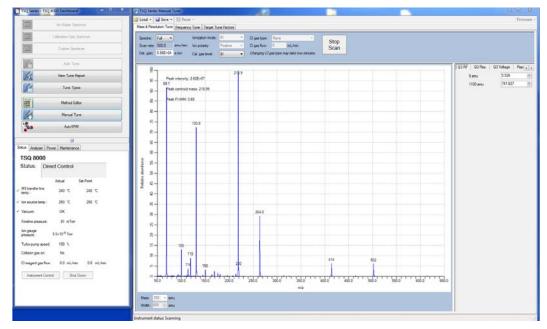


Figure 117. Normal PFTBA Profile Spectrum

A normal PFTBA profile spectrum should meet the conditions below.

- All status indicators on the TSQ Series Dashboard are green.
- Turbo Pump = 100%
- Intensity >1.5e7 (15,000,000)
- The ions m/z 69, 131, 219, 414, and 502 are all present and in the correct relative ratios.
 - *m/z* 69 or *m/z* 219 is the base peak
 - *m*/*z* 131 and *m*/*z* 219 between 60-99%
 - *m/z* 414 and *m/z* 502 between 2-10%
 - *m/z* 100, 119, and 264 are also present and cleanly separated from any noise.
- Mass assignments are correct. Review the tune report for true mass assignment values.
- No extraneous peaks indicating contamination are present.

Some of the most common reasons and indications that your system needs maintenance are as follows:

• **Contamination**—Excessive background in your mass spectra usually indicates that your instrument is contaminated. Use the mass spectrum in the table below to understand the origin of the contamination. Cleaning solvent peaks usually indicate that the ion source cartridge was installed before it completely dried.

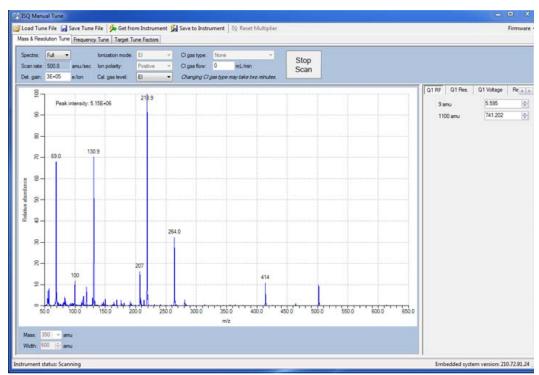
- **Fingerprints**—A series of mass peaks in your data that are 14 amu apart usually indicates fingerprint or other hydrocarbon contamination. To avoid fingerprints, wear clean, lint-free gloves when performing any type of maintenance on a component in the vacuum manifold of the instrument.
- Air Leaks—Higher than normal vacuum pressure or poor sensitivity usually indicates an air leak. Check the last o-ring or ferrule that you installed.

Table 3. Common Contaminants

lons (<i>m/z)</i> To Monitor	Compound	Possible Source
13, 14, 15, 16, 17, 29, 41, 57	Methane	CI gas
18, 28, 32, 44 or 14, 16, 19	H ₂ O, N ₂ , O ₂ , CO ₂ or N, O	Residual air and water, air leaks, outgassing from Vespel ferrules
69, 100, 119, 131, 169, 181, 214, 219, 264, 376, 414, 426, 464, 502, 576, 614	PFTBA and related ions	PFTBA (tuning compound)
31	Methanol	Cleaning solvent
43, 58	Acetone	Cleaning solvent
78	Benzene	Cleaning solvent
91, 92	Toluene or xylene	Cleaning solvent
105, 106	Xylene	Cleaning solvent
151, 153	Trichloroethane	Cleaning solvent
149	Plasticizer (phthalates)	Use of vinyl or plastic gloves
Peaks spaced 14 amu apart	Hydrocarbons	Fingerprints, foreline pump oil, or other hydrocarbons

Figure 118 shows an abnormal PFTBA profile spectrum.





In Figure 118, the abnormal results are as follows:

- Maximum intensity < 1.5e7 (15,000,0000)
- m/z 207 is prominent, indicating column bleed
- Many small contamination peaks present

Investigating Baseline Issues

Behavior	Characteristic	Cause	Remedy
Drifting Baseline	General	Stationary phase has accumulated in column	Replace the column or cut off the end of the column.
		Chromatographic baseline is high	Replace the column or cut off the end of the column.
		Carrier gas pressure is too low	Check for leaks in injector or flow path. Replace the carrier gas cylinder if it is empty or low. Increase the pressure if maximum injector pressure in method is greater than carrier line pressure set by regulator. Make sure the vacuum compensation is on.
		Carrier gas flow is drifting	Check for leaks in injector or flow path. Check the carrier gas tank.
		Impurities have accumulated in column	Run solvent blank to remove impieties. If impurities persistent after multiple solvent blanks: Inject solvent from a different source. Change syringe, liner and septum. Clean injector. Check impurity levels in your gas. Use the correct gas purity and filter.
	Falling	Carrier gas leak in the system	Perform a leak test and tighten the connections to the carrier gas line if leak is found.
		Column is baking out	Wait for the column to stabilize.
	Rising	Impurities have accumulated in column	Check impurity levels in the gas source. Use correct gas purity.
	Abnormal rise in baseline when oven temperature is high	Impurities have accumulated in column	Recondition or replace the column.

Behavior	Characteristic	Cause	Remedy
Noisy Chromatographic Peaks	General	Excessive column bleed at high oven temperatures	Reduce the column temperature. Bake out the column. Install a high-temperature column.
			Install oxygen filters in carrier gas line. Check pneumatic and inlet systems for leaks. Use correct gas purity with low oxygen content.
		Column is contaminated or damaged	Condition or replace the column.
		Oven temperature is higher than column's maximum allowable temperature	Reduce oven temperature to the maximum allowable temperature of the column.
		Leak at column fittings	Find leak. Tighten fittings if loose. Replace ferrule if overtightened.
			Transfer line temperature is not set too low.

Investigating Peak Issues

 Table 4.
 Troubleshooting Peak Issues in Your Data (Sheet 1 of 3)

Behavior	Characteristic	Cause	Remedy
Broadening General	General	Column higher than optimum of column	Reduce the flow. Make sure vacuum compensation is turned on.
		Column flow lower than optimum of column	Increase the flow.
		Split flow is too low for split injection	Increase the flow to 40-50 ml/min.
	Performance of the column has degraded	Test the column at the optimum flow rate.	
		Injector is dirty	Clean or replace the liner.
		Ion source is dirty	Clean the ion source and tune the instrument.
		Column is not far enough into the transferline	The GC column does not extend into the MS source. Use the column measuring tool to confirm column length. If the end of the column is inside the transfer line, an excessive amount of GC effluent will contact the inside wall.
Fronting	General	Column is overloaded	Decrease the injected amount and/or analyte concentrations. Increase the split ratio. Use a column with a thicker film.

Behavior	Characteristic	Cause	Remedy
Tailing	Sample peaks	Column degradation is causing activity	Inject a test mixture and evaluate the column. Replace column if necessary.
		Liner is dirty	Clean or replace the liner.
		Ion source is dirty	Clean the ion source and tune the instrument.
		Glass wool or inlet liner is causing activity	Replace wool with fresh silanized wool and install a clean inlet liner.
		Inlet temperature is too low	Increase the temperature of the inlet.
		Column connections are poor or obstructed	Reconnect the column inlet.
		Stationary phase is not appropriate for your target compounds	Replace the column and choose a more appropriate phase for your analysis.
		Final hold oven temperature is too low	crease the column/oven temperature. Do not exceed the recommended maximum temperature for the stationary phase.
		Transferline temperature is too low	If tailing occurs on late eluting compounds, it is likely the source or transferline temperature is too low.
		Source temperature is too low	If tailing occurs on late eluting compounds, it is likely the source or transferline temperature is too low.
		Poor column characterization	See Changing the Column for information about checking for leaks and column flow.

 Table 4.
 Troubleshooting Peak Issues in Your Data, continued (Sheet 2 of 3)

Behavior	Characteristic	Cause	Remedy
Ghost Peaks	General	Incomplete elution of previous sample	Increase the final oven program temperature or total run time. Increase the column flow rate.
		Carrier gas is contaminated	Replace the gas cylinder or filter.
		Laboratory glassware has caused contamination	Ensure the glassware is clean and contaminant-free.
		Injected sample has decomposed	Decrease the injection port temperature. Use the on-column injection technique.
		Injection solution has matrix present	Adequately clean up your sample prior to injection.
		Inlet or pneumatics are contaminated	Remove the column and bake out the inlet. Use a high-quality septum. Replace the split vent filter. Install an in-line filter between the pneumatics and the inlet.
Missing Peaks	Baseline or background present	Column is broken	Replace the column.
		Column flow is incorrect	Make sure the septa are sealing. Make sure vacuum compensation is turned on.
		Backflush settings are incorrect	Set backflush to off until after injection.
		Column position in S/SL injector is incorrect (too high)	Check the position of the column.
	No baseline or background present	Poor or missing electrical connection	Check the cable connections.
		Mass spectrometer is not collecting data	Make sure the tune file is correct. Verify that the Busy light is on during acquisition. Make sure the filament is not burned out. Close <i>Xcalibur</i> , open <i>Instrument</i> <i>Configuration</i> , press the Reset button on the mass spectrometer, wait ten seconds, close <i>Instrument Configuration</i> .

Table 4. Troubleshooting Peak Issues in Your Data, continued (Sheet 3 of 3)

Behavior	Characteristic	Cause	Remedy
Low Reproducibility of Peaks Area	General	Detector gain is set too low	Retune the gain. Increase the electron multiplier voltage. Increase the target ion count.
		Concentration is not compatible with the dynamic range of the detection system	Verify that the sample concentration is suitable for the system.
		Chromatogram and spectrum are blank	Make sure the tune file is correct. Verify that the Busy light is on during acquisition. Make sure the filament is not burned out.
		Injection technique is not appropriate	Use a different injection technique.
		Injection parameters are not appropriate	Verify the injection temperature and flow rates.
		Sample injection technique is not reproducible	Evaluate the sample preparation sequences. Compare the results with a series of standard injections.
		Syringe or septum is leaking	Check and replace the syringe and/or septum at regular intervals.
		Injection port is leaking	Check the column connections. Run a leak check.
		Injection technique is not suitable	Carefully meter the injected amount. Use a clean, good-quality syringe.
		Ion source is dirty	Clean the ion source and retune the instrument.
		Split flow or ratio control is inadequate	Monitor the flow. Replace the in-line filter.

Investigating Results Issues

Behavior	Characteristic	Cause	Remedy
Poor Sensitivity	With increased retention time	Carrier gas flow rate is too low	Increase the carrier gas flow rate. Locate and remove possible obstructions in the carrier gas line. Check the septum for leaks. Check the injector/column ferrules for leaks.
	With normal	GC carrier gas line has leaks	Run a leak test and correct leaks.
	retention time	Syringe is leaking during injection	Replace syringe or piston seals, if necessary.
		Split injection temperature is too low	Increase the temperature of the injector.
		Voltage is not reaching the lens.	Replace the lens plate and springs if damaged. Remove debris or broken pieces in the manifold. Run a lens check diagnostic. Check the connection by removing or inserting the ion source.
Retention Times	Low reproducibility	DCC is drifting or unstable	Monitor the column pressure or flow. Check and replace the controller, if necessary.
		Injection technique is inadequate	Pick injection technique suitable for the injector and liner you are using.
		Vaporization size of sample inject larger than volume of liner	Reduce the injected amount and/or volume.
		Column temperature is unstable	Check the main oven door and cooling flap. Monitor the column temperature.

Index

A

acquisition methods, purpose of, 65 AutoTune, modifying, 95 Autotune, using, 31

B

baseline (troubleshooting) drifting, 126 baseline issues, investigating, 126

C

centroided data, about, 70 chromatographic separation, changing, 91 column changing, 15 leak checking, 19 temperature, 21 compliance FCC vi regulatory v WEEE vii Configuring the TSQ Duo, 6 contaminants, common, 124 contamination, indications, 123

D

data result issues, investigating, 131 diagnostics, selecting tests to run, 100 documentation survey xxi

E

electromagnetic compatibility vi

F

FCC compliance vi

filament voltage, setting, 97 filament, turning off and on, 72 fingerprint, indications, 124 full-scan mode, about, 69

G

GC creating a method, 61 ghost peaks, troubleshooting, 130

I

ion source temperature, setting, 66, 98

L

leak, indications, 124 leaks, checking for, 19

Μ

maintenance, settings, 10 mass range, narrowing, 93 methods acquisition, 65 creating, 53 purpose of, 53

Ρ

peak height, minimum, 66 peak issues, investigating, 128 peaks (troubleshooting) broadening, 128 fronting, 128 ghost, 130 none, 130 tailing, 129 profiled data, about, 70

R

regulatory compliance v results (troubleshooting) low reproducibility of peaks area, 131 poor sensitivity, 132 retention times, 132 retention times, troubleshooting, 132 routine maintenance causes, 121

S

safety standards v scan rate, optimizing, 92 scans, running, 67 sensitivity, troubleshooting, 132 survey link xxi

Т

tailing peaks, troubleshooting, 129 temperature of the ion source, 66 temperature of the transfer line, 93 temperature of transfer line, 66 transfer line temperature, setting, 66 transfer line, about, 66 troubleshooting about, 113 baseline issues, 126 peak issues, 128 results issues, 131 tune report, 46 TSQ Duo changing the column, 15 configuring, 6 creating a method, 53 TSQ Series AutoTune, about, 95 tune reports example report, 46 printing, 46 settings, 46 tune types adding, 42 CI- tune, 38 daily tune check, 33 Daily Tune, 37 daily tune, 31 EI default tune, 37 EI full tune, 33 tuning using Autotune, 31 with TSQ Series Autotune, 31

V

voltage of filament, setting, 97

W

WEEE compliance vii